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1 **Adding a diversity of legumes to a crop decision-support system: maintaining satisfactory accuracy**
2 **while keeping the model simple**

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22

23 **Abstract**

24 In a context of economic and environmental concerns in agriculture, legumes appear to be suitable
25 alternative crops to diversify current cropping systems and reduce their dependence on synthetic
26 nitrogen (N) fertiliser and protein from imported soya bean. However, legume-based cropping systems

27 may increase N losses through nitrate leaching if the N available from legumes does not coincide with
28 subsequent crop requirements. To help agricultural advisers manage N in these systems, we adapted
29 the decision-support system Syst’N®, designed to assess N losses in cropping systems, to simulate
30 three annual and one perennial legume crops: pea, faba bean, soya bean and lucerne. To this end, we
31 adapted and simplified existing submodels of legume functioning to include them in Syst’N, to keep
32 the latter simple. We adapted the submodels “BNF” (i.e. biological N fixation) from the STICS model
33 and “dormancy” from the CropSyst model. We also added the ability to enter the flowering date to
34 improve predictions (improvement in N fixation’s rRMSE from 57% to 41% and EF from 0.57 to 0.77).
35 The equations and associated parameter set developed for the four legume crops yielded satisfying
36 predictions of crop biomass (rMBE = 9%, EF = 0.82, rRMSE = 39%) and N content (rMBE = 5%, EF = 0.76,
37 rRMSE = 37%). These performances support the philosophy of Syst’N® that requires minimising the
38 number of additional parameters for users when representing new crops or processes.

39

40 **Keywords:** nitrogen, legume crops, decision-support system, biological N fixation, model

41

42 **1. Introduction**

43 The agricultural revolution after 1945 led to a major intensification of French agriculture due to the
44 increasingly intensive use of mechanisation and chemically synthesised inputs, which was enabled by
45 technical progress. This intensification and the organisation of agriculture into sectors have fostered
46 specialisation of production systems with a geographical separation between crop and livestock
47 production and increasingly shorter cropping sequences (Ferrant, 2009). These changes, based on
48 choosing crop species that maximise profit in the short term, led to the abandonment of species that
49 provided recognised agronomic and ecosystem services such as legumes (Schott et al., 2010).
50 Moreover, historical choices of public policies (e.g. subsidies for cereal production), research and
51 engineering triggered the marginalisation of legumes (Magrini et al., 2016). Since the 1990s, however,
52 growing awareness of negative externalities associated with cereal-based intensive systems (e.g.

53 environmental pollution, greenhouse gas emissions, loss of biodiversity) has prompted some
54 stakeholders in the agricultural sector to seek more economical and environmentally friendly practices
55 (Voisin et al., 2013). In this context, legumes appear to be good alternative crops to diversify current
56 cropping systems and reduce their dependence on synthetic N fertiliser and protein from imported
57 soya bean (Carof et al., 2019). Comparing N fixed by legumes vs. industrial sources, Crews and Peoples
58 (2004) showed that developing legume-based cropping systems could lead to more sustainable
59 agriculture. Indeed, legumes have the unique ability to fix N from the air (N_2), thus reducing their need
60 for N fertilisation; in Europe, legume crops are usually not fertilised (Cernay *et al.*, 2016). Biological N
61 fixation (BNF) by grain/forage legumes also benefits subsequent crops by leaving more N in the soil
62 than cereals do (Herridge et al., 1995) and through mineralisation of N-rich plant residues incorporated
63 into the soil after crop harvest, thus increasing soil N fertility (Justes et al., 2001). However, legume-
64 based cropping systems may increase N losses through nitrate leaching if the N available from legumes
65 does not coincide with subsequent crop N requirements (Muller et al., 1993; Cellier et al., 2015; Plaza-
66 Bonilla et al., 2015; Voisin et al., 2015). Legumes differ in their N-fixation efficiency, which depends on
67 the response of their BNF to abiotic factors (e.g. soil mineral N content, temperature, soil water
68 content) and their physiological functioning (Guinet et al., 2018). Therefore, legume BNF and N pre-
69 crop effects must be quantified over time to improve the design of sustainable cropping systems that
70 include legumes. Doing so requires improving quantification of benefits for subsequent crops from
71 legume BNF and its influence on N losses, and thus environmental pollution.

72 In this context, agricultural stakeholders need to design new cropping systems using decision-making
73 tools that consider legumes and that can estimate their environmental impacts. Since the 1960s, many
74 crop models have been developed to address a variety of objectives, but only a few are directly used
75 by practitioners to simulate N dynamics in legume-based cropping systems. This arises from a lack of
76 user-friendly interfaces, low availability of input parameters and failure to consider users' constraints
77 and knowledge (Prost et al., 2012). Meanwhile, users in the environmental field and agricultural

78 advisers use indicators or simple tools that fail to simulate N-loss dynamics and to understand
79 weaknesses of cropping systems.

80 As an operational tool for quantifying N losses at the multi-year scale of a cropping system, Syst'N®
81 predicts N dynamics under the most common crops in France. Syst'N is used mainly by agricultural
82 advisers for regions with strong environmental issues (especially nitrate emissions); thus, it must
83 estimate N losses and help discuss and design more sustainable cropping systems. Stakeholders
84 involved in water and agricultural management who use Syst'N need it to be applicable to a variety of
85 cropping systems that include less common crops, especially legumes (as grain or forage). In the first
86 version of Syst'N (2013), pea (*Pisum sativum* L.) was the only legume crop parameterised.
87 Consequently, there was a real need to extend the scope of Syst'N for users and adapt it to new
88 cropping systems with a wider variety of legume crops.

89 Accordingly, this study developed, evaluated and validated Syst'N predictions for several common
90 legume species: faba bean (*Vicia faba* L.), soya bean (*Glycine max* L.) and lucerne (*Medicago sativa* L.).
91 Pea was parametrised again for the new equations added to predict the other legume species
92 considered, and predictions for pea were also evaluated. This adaptation of the model is crucial
93 because satisfactory simulation of growth and N uptake is essential to meet Syst'N's ultimate goal,
94 which is to estimate N emissions to the environment.

95 We adapted Syst'N by choosing and adapting generic equations that could simulate N dynamics for
96 several legumes without decreasing its user-friendly aspect. The design criteria for adapting Syst'N to
97 four legumes included: (i) the ability to simulate growth and N uptake of annual and perennial legumes,
98 (ii) the ability to simulate soil N dynamics and content during and after a crop cycle sufficiently well
99 (e.g. residual N effects of legumes), (iii) avoiding the need for users to provide additional input data,
100 (iv) limiting the number of additional parameters needed for new crops and (v) adapting equations
101 from updated knowledge and models. We assumed that adapting Syst'N to a wide range of legumes
102 would meet these criteria.

103

104 **2. Materials and methods**

105 **2.1. The crop model Syst'N**

106 Syst'N is a model-based decision-support system (DSS) developed to estimate N losses in cropping
107 systems and promote better N management in rural areas. The French National Research Institute for
108 Agriculture, Food and Environment (INRAE) and eight French agricultural institutes developed Syst'N
109 together. The objective was to meet the needs of stakeholders involved in water and agricultural
110 management for an operational tool that could predict N losses at the multi-year scale of cropping
111 systems. Since 2005, Syst'N has been co-designed with end-users to adapt it further to their needs
112 (Cerf et al., 2012), in an iterative process with agricultural advisers, research and development
113 institutes, applied research institutes and environmental agencies.

114 The Syst'N version described by Dupas et al. (2015) added a soil-crop model to simulate soil N
115 transformations, crop growth, N uptake, water balance and N losses towards water (e.g. nitrate) and
116 air (e.g. ammonia, N₂ and nitrous oxide) on a daily time step. The input data required include the crop
117 sequence, agricultural management practices, soil and climate. The equations in the biophysical model
118 combine existing validated and published submodels from STICS (Brisson et al., 2003) for water and
119 nitrate balances in soils, AZOFERT (Machet et al., 2004; 2017) for N mineralisation of soils and crop
120 residues, AZODYN (Jeuffroy and Recous, 1999) for crop N uptake, NOE (Hénault et al., 2005) for N₂ and
121 nitrous oxide emissions and VOLT'AIR (Génermont and Cellier, 1997) for ammonia emissions.

122 Syst'N attempts to chart a middle course between complex simulation models such as STICS (Brisson
123 et al., 1998), which simulate many processes but require many input data, and more simple indicators
124 commonly used by agricultural practitioners (e.g. N balance, N-use efficiency) but which have relatively
125 little ability to explain phenomena. Therefore, the equations added to Syst'N were selected to conform
126 to the input data that are generally available to targeted end-users, hence keeping Syst'N user-friendly.

127 **2.2. Experimental data**

128 Most of the parameters initially used to calibrate the new legumes (lucerne, faba bean and soya bean)
129 were derived from previously published studies. The data used to calibrate and evaluate Syst'N were

130 collected in French field experiments, in contrasting pedoclimatic contexts and with different
131 experimental treatments, such as various amounts of N fertilisation or irrigation applied (Table 1; see
132 Table S1 for more details). The objective was to find data with the highest diversity in experimental
133 treatments, including sowing date, amounts of N fertiliser and irrigation applied, as well as cutting
134 dates for lucerne as a perennial crop. Ultimately, fertiliser applications could be tested only for pea,
135 faba bean and lucerne. Site-year-management (SYM) units that covered a wide range of crop-
136 management practices with high measurement frequency were chosen for the calibration dataset (6,
137 6, 9 and 17 for pea, faba bean, lucerne and soya bean, respectively), and the remaining SYM units
138 formed the evaluation dataset (12, 3, 32 and 9 SYM, respectively).

139 **2.3. Adaptation and calibration method**

140 To begin adapting Syst’N, equations and corresponding parameters that represented plant processes
141 (e.g. phenology, biomass accumulation, BNF) were chosen (Fig. 1). First, specific characteristics of each
142 legume crop were identified. If the existing Syst’N equations could not simulate these characteristics
143 or the resulting N dynamics satisfactorily, the equations were supplemented or modified based on
144 those in existing models. Initial parameters for the crops were set based on a literature review of
145 existing models and plant physiology articles, or directly on measurement datasets if parameter values
146 were not found in the literature.

147 We then graphically and statistically tested these initial equations and parameters using calibration
148 datasets (Table 1). If predictions were unsatisfactory, some parameters were selected according to a
149 set of criteria (e.g. sensitivity of variables of interest to the parameter, uncertainty) and tested for a
150 range of values to improve prediction accuracy by optimising statistical indicators of accuracy.

151 **2.4. Model assessment**

152 As we were adding several legumes to Syst’N, we assessed it by considering important plant variables,
153 such as aboveground biomass and N uptake, as well as BNF and soil mineral N content when datasets
154 included them. Predicted target variables of Syst’N (i.e. N losses) were not evaluated because they

155 were not measured in experiments and because only plant-related equations and parameters were
156 modified in this study.

157 Model performance (with the optimised parameter set) was evaluated graphically as dynamics over
158 the crop cycle and plots of predictions vs. observations. It was quantified by calculating mean bias error
159 (MBE) and its relative value (rMBE); root mean square error (RMSE) and its relative value (rRMSE); and
160 model efficiency (EF) using data collected at harvest and throughout crop growth. These indicators are
161 commonly used to calibrate and evaluate agronomic models (Dumont et al., 2012).

$$162 \quad \text{MBE} = \frac{1}{N} \sum_{i=1}^N (P_i - O_i) \quad (1)$$

$$163 \quad \text{rMBE} = \frac{\text{MBE}}{\bar{O}} \times 100 \quad (2)$$

$$164 \quad \text{RMSE} = \sqrt{\frac{1}{N} \sum_{i=1}^N (O_i - P_i)^2} \quad (3)$$

$$165 \quad \text{rRMSE} = \frac{\text{RMSE}}{\bar{O}} \times 100 \quad (4)$$

$$166 \quad \text{EF} = 1 - \frac{\sum_{i=1}^N (O_i - P_i)^2}{\sum_{i=1}^N (O_i - \bar{O})^2} \quad (5)$$

167 where O_i and P_i are observed and simulated values for the i^{th} measurement, N is the number of these
168 paired values and \bar{O} is the mean of observed values.

169 MBE and rMBE indicate whether a model underestimates (negative values) or overestimates (positive
170 value) a given variable. Nevertheless, bias is not a sufficient measurement of the quality of a model:
171 low bias can result from small prediction errors in all situations or instead from large prediction errors
172 that compensate each other. Hence, the RMSE was also calculated (expressed in the same unit as the
173 given variable) to identify this latter problem by squaring the difference between simulations and
174 observations. Doing so, however, gives more weight to larger errors, which requires caution when
175 interpreting RMSE, because a large RMSE can result from only one or two major differences. The rRMSE
176 is more relevant for comparing variables with different units. Finally, EF assesses the overall
177 performance of a model by comparing it to the mean of observations. EF ranges from $-\infty$ to 1, which

178 facilitates interpretation: negative EF indicates that the model predicts no better than the mean of
179 observations, whereas EF = 1 indicates a model with perfect accuracy. EF is relevant for comparing
180 models or for comparing the same model's simulations of the same dataset using different sets of
181 parameters/equations. Analysing these five statistical indicators from different groups of correlated
182 indicators give a robust assessment of model prediction accuracy, as recommended by Yang et
183 al.(2014). Like for Beaudoin et al. (2008) and Constantin et al. (2015a, 2015b), Syst'N predictions were
184 considered satisfactory when EF exceeded 0.50 and rMBE was lower than 10% (Table 2). A 20%
185 threshold was used for rRMSE, like for Thiebeau (2019a; 2019b), although a higher threshold (40%)
186 was used to evaluate STICS predictions for lucerne (Strullu et al., 2020).

187

188 **3. Results**

189 **3.1. Model description and parameterisation for legume crops**

190 **3.1.1. Mineralisation of crop residues**

191 Syst'N already used a double exponential equation from the AZOFERT tool to simulate net N
192 mineralisation from crop residues (Machet et al., 2017):

$$193 \quad N = N_{RO} \times (a_N - b_{NE}^{-kt} - c_{NE}^{-lt}) \quad (6)$$

194 where N is amount of N mineralised; N_{RO} is the initial amount of N in crop residues and a_N , b_N , c_N , l and
195 k are coefficients that describe the mineralisation kinetics of N.

196 Before this study, the equation was parameterised for a variety of legume residues. The same
197 parameterisation was used in this study. Inspired by the model TNT2 (Casal et al., 2019) for grazed or
198 cut perennial crops (including lucerne in Syst'N), an additional N sequestration pool was added during
199 crop development depending on grazing or cutting practices. After crop harvest or cover crop
200 destruction, this pool of organic N mineralises.

201 **3.1.2. Biological nitrogen fixation**

202 As shown by the systematic literature review and quantitative statistical analysis of Anglade et al.
203 (2015), legume species differ significantly in their median percentages of N derived from the

204 atmosphere (%Ndfa), due mainly to the species' sensitivity to abiotic factors (e.g. soil mineral N
205 content, soil water content, temperature) and to their phenological characteristics (e.g. phenological
206 stages, nodule establishment rate) (Guinet et al., 2018; Pampana et al., 2018). In the first version of
207 Syst'N, the equation used to predict BNF was based on the AFISOL-Pea model, but it was adapted only
208 for pea. In the present study, it was necessary to consider the diversity of BNF processes among legume
209 species. The most common method for estimating %Ndfa of legume species in mechanistic crop
210 models is to calculate a potential or maximum BNF rate which is then decreased by the influence of
211 environmental factors: in their review, Liu et al., (2011) identified nine simulation models (Sinclair
212 Model, EPIC, Hurley Pasture Model, Schwinning Model, CROPGRO, SOILN, APSIM, Soussana Model and
213 STICS) that simulated legume BNF. Each model considered one or more of the following factors: soil
214 temperature, soil-plant water, soil-plant N, plant carbon and phenological stage. Some BNF models are
215 calibrated and parameterised for only one legume species, whereas others can simulate a wide range
216 of legume species with different characteristics and geographic ranges.

217 To adapt Syst'N to a variety of legume species (grain and forage), we chose to add the BNF submodel
218 of STICS (Brisson et al., 2009; Corre-Hellou et al., 2007), because it (i) is adapted to the French
219 pedoclimatic context, (ii) is adapted for a wide range of legumes (e.g. field pea, soya bean, faba bean,
220 lucerne) and (iii) considers all the environmental factors mentioned except carbon dynamics (ignored
221 by Syst'N). The BNF equations of STICS were modified slightly to conform to the input data available in
222 Syst'N (i.e. using air temperature instead of soil temperature for the temperature-weighting factor)
223 and to the variables already calculated by Syst'N (beginning of nodulation associated with the
224 beginning of BNF, nodule death associated with the final stage of seed abortion) (Fig. 2). The
225 parameters specific to these equations, which also resulted from parameterisation of STICS, varied
226 among species (Table 3).

227 3.1.3. Phenology

228 As for most crops, legume phenology is controlled mainly by air temperature and photoperiod. As a
229 simple crop model, Syst'N calculates phenological stages using only sums of growing degree-days.

230 According to their photo-thermal sensitivity, legumes can be classified into three main categories:
231 photoperiod-insensitive, day-neutral and photoperiod-sensitive (short-day or long-day) (Roberts and
232 Summerfield, 1987). As many studies observed, sensitivity to photoperiod is even more important for
233 legume species, because it drives their transition to the reproductive stage (i.e. flowering) (Baranger
234 et al., 2010; Confalone, 2008; Iannucci et al., 2008; Schneider and Huyghe, 2015). Because flowering
235 stops (e.g. root growth), modifies (e.g. remobilising N, BNF) or triggers (e.g. plant senescence) some
236 plant processes, and thus influences N dynamics, it seemed important for Syst’N to predict flowering
237 time accurately. However, simulating photoperiod and its effects on crop phenology would (i) be too
238 complex for such a DSS, (ii) require new input data (e.g. latitude) and (iii) require new parameters for
239 the photoperiod sensitivity for each crop, which seems difficult to define. Thus, we chose to add the
240 actual flowering date as optional input data. Indeed, a survey of Syst’N users indicated that they often
241 observe it and thus could provide it as input data. When flowering date is provided, Syst’N calculates
242 all phenological stages from it; if not, it calculates stages from the crop parameter that represents the
243 number of degree days from sowing to flowering. When parametrising annual legumes, we decided to
244 distinguish soya bean of types 00 and I, as their stages differ greatly.

245 **3.1.4. Perennial legumes**

246 We modified Syst’N greatly to capture specific characteristics of perennial crops, such as several cuts
247 and regrowth sequences. In the first version of Syst’N, an equation from TNT2 (Casal et al., 2019) that
248 estimated the change in plant biomass and N content when cutting grassland (Nitschelm et al., 2018)
249 was used and adapted to represent other perennial crops such as lucerne. The residual aboveground
250 biomass that remains in the field after a cut is calculated from the cutting height. Nevertheless, plant
251 growth differs between seedlings and crop regrowth (after harvest or dormancy), as observed by
252 Thiébeau et al. (2011) and Lemaire et al. (1985) for lucerne, due to crop defoliation and storage of
253 reserves in roots, which supports regrowth after cutting or after winter. To keep Syst’N relatively
254 simple, it contains two parameter sets to simulate seedlings and crop regrowth separately instead of

255 using complex equations to represent reserve management by perennial crops, as in STICS versions for
256 perennial crops (Strullu et al., 2014; 2020).

257 Some vigorous perennial legumes, such as lucerne, become dormant during winter (Undersander et
258 al., 1997). Dormancy is triggered mainly in autumn by two factors: photoperiod and temperature
259 (Mauriès, 2003). As mentioned, simulating photoperiod would require new input data from users and
260 make Syst’N more complex. An equation from CropSyst (Confalonieri and Bechini, 2013) was a good
261 compromise to add dormancy to Syst’N in a relatively simple way (Fig. 3). This equation was adapted
262 slightly in Syst’N to fit with its existing equations of biomass growth and N uptake. The parameters
263 used in these equations were calculated from the optimisation measurement dataset. For simulated
264 plant N content, this adaptation delays crop regrowth after winter and resets crop-stage variables,
265 thus avoiding the unexpected early physiological maturity predicted without the dormancy equation
266 (Fig. 4).

267 3.2. Results of model evaluation

268 The final parameters for legumes in Syst’N were set by the calibration method described (Table S2).

269 3.2.1. Calculating phenological stages from the actual flowering date

270 To evaluate effects of calculating phenological stages from the observed flowering date on the
271 accuracy of Syst’N predictions, the statistical indicators were calculated for predictions based on a
272 dataset with or without the actual flowering date. The latter dataset was smaller than the
273 measurement dataset, as it contained only the [site-year-management](#) (SYM) units in which flowering
274 had been observed. The accuracy of predicted total aboveground plant N (Fig. 5C and 5D) content and
275 BNF (Fig. 5E and 5F) increased, and that of predicted aboveground biomass remained good (Fig. 5A
276 and 5B), which indicated the relevance of calculating phenological stages from the actual flowering
277 date (as input data), as flowering is an important stage for BNF and N uptake. The rMBE were low (-6%
278 to 19%) and showed Syst’N’s tendency to overestimate total aboveground plant N (rMBE = 19%, Fig.
279 5C) but underestimate BNF (rMBE = -6%, Fig. 5E). However, the relatively good EF (0.73-0.85) of

280 predictions of all plant variables confirmed that Syst'N predicted growth and N assimilation (uptake
281 plus BNF) throughout the crop cycle (from juvenile to mature stages) better when stages were
282 calculated from the actual flowering date. All rRMSE were satisfactory (< 41%).

283 3.2.2. Accuracy of predictions over the crop cycle

284 3.2.2.1. Aboveground biomass

285 Predicted biomass agreed with observations throughout each legume crop's growing season for both
286 calibration and evaluation datasets ($EF > 0.75$, $9\% < rMBE < 13\%$), considering a dataset with or without
287 flowering date (Fig. 6A and 6B). Positive rMBE showed Syst'N's tendency to overestimate aboveground
288 biomass, especially for lucerne with the calibration dataset (data not shown). Prediction error was high
289 (rRMSE of 40% for both datasets) due to senescence defoliation and the loss of biomass related to
290 respiration during dormancy (Justes et al., 2002), which Syst'N ignores for all crops. Hence, Syst'N
291 assumes that aboveground biomass does not change during dormancy. Nevertheless, it can be
292 considered a time-lag in growth dynamics that does not influence Syst'N prediction, as they became
293 more accurate after dormancy (Fig. 7).

294 For each legume crop, all statistical indicators were good: rMBE was low (i.e. $-13\% < rMBE < 20\%$), EF
295 was satisfactory (> 0.59) and rRMSE was rather satisfactory (< 48%) (Table 4).

296 3.2.2.2. Plant nitrogen content

297 3.2.2.2.1. Total aboveground nitrogen content

298 Total aboveground N content was predicted accurately using both the calibration and evaluation
299 datasets considering a dataset with or without flowering date for evaluation (Fig. 6C and 6D): rMBE
300 was low (19% and 5%, respectively), EF was high (> 0.70) and rRMSE was satisfactory (< 40%).
301 Prediction of aboveground N content was satisfactory for faba bean and pea, with good rMBE (< 16%),
302 high EF (0.87) and low rRMSE (< 25%) (Table 4). Despite lower EF (0.61) and higher rRMSE (44%),
303 predictions for lucerne using the evaluation dataset remained satisfactory, with a slight overestimation
304 of the dynamics. In contrast, rMBE (23%), EF (0.48) and rRMSE (44%) for soya bean showed a tendency
305 for Syst'N to overestimate aboveground N content (Table 4).

306 **3.2.2.2.2. Aboveground nitrogen fixation**

307 The parameters related to BNF defined for each crop (Table 3) yielded good predictions (Fig. 6E and
308 Fig. 6F), with a high EF (> 0.78) and low rMBE ($-13\% < \text{rMBE} < 6\%$) for pea, faba bean and soya bean
309 (Table 4). Because observed data were not available for lucerne, we compared predictions to data from
310 the literature (Kelner et al., 1997; Schneider and Huyghe, 2015; Wivstad et al., 1987; Xie et al., 2015).
311 They showed highly variable %Ndfa of 20-90%, which seemed to be explained in part by variable
312 amounts of mineral N in the soil. These percentages were found in the predictions, with a mean %Ndfa
313 of 70% for unfertilised plots and 50% for plots that had received mineral N fertiliser. For the other
314 legume crops, this good accuracy was verified throughout growing season (EF=0.80, low rMBE). EF
315 remained high and bias remained low with the evaluation dataset, indicating that Syst’N was able to
316 reproduce the wide range of cumulative BNF at harvest (33-265 kg N.ha⁻¹).

317 **3.2.2.3. Soil water content and soil mineral nitrogen content**

318 Syst’N predicted soil water content better during evaluation (EF = 0.44) than calibration (EF = -2.38),
319 despite lower rMBE (-7% and 20%, respectively) and higher rRMSE (33% and 24%, respectively) (Fig.
320 8). Despite satisfactory indicators, we observed threshold effects. Syst’N had difficulty predicting the
321 wide range of soil water content observed (2.5-33.2%), as it was constrained by threshold values of
322 the pedotransfer classes used to define field capacity (θ_{FC}) and wilting point (θ_{WP}) as a function of soil
323 texture (9.1-26.4%).

324 Soil N content was predicted less well using the calibration and evaluation datasets, with negative EF
325 (-0.19 and -0.24, respectively) and high rRMSE (45% and 67%, respectively) (Fig. 8). The low EF (-0.76)
326 and high rRMSE (69%) for lucerne (Table 4) could be explained mainly by temporal shifts (data not
327 shown). First, nodule activation, which enables BNF, and moderate N uptake from the soil can occur
328 sooner than predicted. This delay in prediction by a few days may cause Syst’N to underestimate soil
329 N content at a given date significantly. Secondly, the simple equation used to simulate dormancy may
330 delay N uptake during biomass regrowth and cause Syst’N to overestimate soil N content for a few
331 weeks, which has little influence on predicted N losses.

332 3.2.3. Nitrogen effect of legumes on subsequent crops

333 Dynamics of N mineralisation after a legume crop were evaluated only for lucerne because of the
334 specific calculations in Syst’N for grazed or cut perennial crops. For the other crops, the N effect of
335 legumes is calculated directly with the AZOFERT equation, which has already been evaluated (Machet
336 et al., 2017).

337 Predicted N mineralisation in the soil after lucerne was compared to measurements from Justes et al.
338 (2001). In this experiment, mineralisation from humus and from residues returned to the soil after
339 lucerne destruction were differentiated. Dynamics of simulated N mineralisation from lucerne residues
340 and soil organic matter accurately reproduced observations for two years in two situations: lucerne
341 cut just before destruction (3% error between prediction (331 kg N ha⁻¹) and observation (340 kg N
342 ha⁻¹) after two years) and lucerne cut one week before destruction (8% error between prediction (368
343 kg N ha⁻¹) and observation (400 kg N ha⁻¹) after two years). Thus, with less than 10% error, predictions
344 of N mineralisation were satisfactory and confirmed Syst’N’s ability to represent long-term effects of
345 lucerne residues.

346

347 4. Discussion

348 4.1. A generic and user-friendly crop model able to simulate a diversity of legumes

349 This study showed that the generic equations of Syst’N can simulate a variety of legumes species: those
350 with cool-season adaptation (pea and faba bean), warm-season adaptation (soya bean) and a perennial
351 cycle (lucerne). The accuracy of predicted aboveground biomass and N assimilation (uptake plus BNF)
352 was satisfactory for all 4 legume species, with rMBE less than 10% (2-9%) and EF greater than 0.50
353 (0.74-0.81), which are consistent with the guidelines of Beaudoin et al., (2008) and Constantin et al.
354 (2015a, 2015b). Moreover, the accuracy of these predictions are similar to those for other species
355 parameterised in Syst’N (unpublished).

356 Prediction of BNF was improved by adding the actual flowering date as input data. Users in the field
357 can obtain the actual flowering date easily, which allows Syst’N to remain adapted to its purpose and

358 user-friendly. In addition, the flowering date can also be estimated using a simple phenology algorithm
359 (Schoving et al., 2020) and then used as an input variable for Syst’N.

360 **4.2. Water balance prediction**

361 Syst’N’s ability to simulate soil water and mineral N contents, both of which are used to calculate N
362 leaching, is less satisfactory than that for biological variables, as all statistical indicators except rMBE
363 lay above satisfactory thresholds. Our predictions of soil N content were similar to those of STICS with
364 faba bean (Falconnier et al., 2019), whose rMBE, EF and rRMSE were -10%, -1.42 and 51%, respectively,
365 using an evaluation dataset.

366 Syst’N predicted both low and high observed soil water contents poorly (Fig. 8) due to the pedotransfer
367 classes it uses to estimate θ_{FC} and θ_{WP} as a function of percentages of clay, silt and sand. Indeed, large
368 differences in soil texture can yield the same characteristic water contents, as observed for 9 site-year-
369 management (SYM) in this study (Fig. 8). For example, because silty-clay loam (30% clay, 62% silt, 8%
370 sand) and sandy-clay loam (14% clay, 32% silt, 54% sand) are considered to be the same in Syst’N’s
371 pedotransfer classes, both have θ_{FC} of 32.3% and θ_{WP} of 14.0%. Many studies have related soil texture
372 to water-retention characteristics of the soil (Gupta & Larson, 1979) using pedotransfer classes or
373 functions. Classes were used in the first version of Syst’N instead of functions based on the results of
374 Al Majou et al., (2007), who showed that “simple” texture-based pedotransfer classes allowed water-
375 retention characteristics to be predicted as accurately as with more sophisticated pedotransfer classes
376 or functions. However, using pedotransfer classes inevitably leads to threshold effects, which
377 decreases the accuracy of predictions of water balance and thus nitrate leaching. Tóth et al. (2015)
378 developed new hydraulic pedotransfer functions valid for the pedoclimatic context in France. The most
379 accurate predictions were obtained using the clay percentage, silt percentage, bulk density, organic
380 carbon and pH as parameters, all of which are available as input data in Syst’N. In future versions, using
381 pedotransfer functions could improve predictions of soil water content, in particular by overcoming
382 the threshold effect caused by pedotransfer classes. Soil water content has a strong influence on BNF
383 and soil N content, but also on water transfer and thus on N leaching, which is one of the main outputs

384 of the model. In addition, the parameters θ_{FC} and θ_{WP} could be used as input data if they are available.
385 Nevertheless, we have noted that the end-users of Syst'N do not often know the values of these
386 parameters.

387 **4.3. Specifying a large number of plant parameters**

388 Using the actual flowering date to calculate phenological stages and the new BNF equation required
389 adding 2 and 16 new plant parameters, respectively, specific to legume species. Thus, 77 agro-
390 physiological parameters for each species must now be defined to simulate a new legume crop in
391 Syst'N. Although some of these parameters require good knowledge of the agro-physiological
392 behaviour of the crop, particularly for N processes (e.g. parameters of N-dilution curves), many of them
393 can be approximated easily using knowledge of other crops already parameterised in Syst'N.
394 Moreover, despite the large number of plant parameters, Syst'N is easier to adapt than most other
395 models, such as STICS, which requires specifying over 200 crop- and cultivar-specific parameters;
396 however, it was developed by researchers to investigate a wider diversity of cropping systems and
397 consider many more processes (Brisson et al., 2009).

398 **4.4. Strong simplification of biological processes for perennial legumes**

399 Predictions highlight high overestimation of aboveground biomass and N content immediately after
400 dormancy due to the lack of equations to simulate senescence and mobilisation of reserves in Syst'N.
401 However, this trend is relatively brief: along with rapid biomass regrowth in the spring, reserves stored
402 in roots during autumn and winter are remobilised to shoots (Justes et al., 2002). The accuracy of
403 predicted aboveground biomass and N content increased after a few weeks and became satisfactory
404 during the cuts in spring. This simplification in Syst'N delays N uptake from the soil in the spring but
405 has little influence on soil N content during winter, a key variable used to predict nitrate leaching. As
406 shown by Strullu et al. (2014) for *Miscanthus x giganteus*, total aboveground N content decreases only
407 slightly in winter despite a strong decrease in aboveground biomass. Rapid remobilisation of N
408 temporarily stored in perennial organs has also been observed for other legumes, such as white clover
409 (*Trifolium repens* L.) (Robin et al., 1999), which supports this hypothesis. In the future, the simple

410 equation that maintains aboveground biomass and N content constant during winter could be applied
411 to other perennial crops in Syst'N.

412 As mentioned, Syst'N has two parameter sets to simulate seedlings and crop regrowth separately,
413 instead of using a complex model to represent reserve management by perennial crops, as in STICS
414 versions for perennial crops. These STICS versions, initially developed for *Miscanthus* reserves (Strullu
415 et al., 2014) and then for lucerne (Strullu et al., 2020), consider the N fluxes between perennial organs
416 (e.g. the taproot) and non-perennial organs (i.e. leaves, stems and roots) as a function of abiotic stress,
417 photoperiod and phenological stages using a single set of parameters, but they required defining more
418 compartments in the model and are thus complex. STICS version for lucerne resulted in EF values of
419 0.70 for aboveground biomass and 0.60 for N content, while Syst'N using less complex equations gave
420 similar EF (0.60 for both biomass and N content). However STICS predictions were slightly more
421 accurate (rRMSE of 31% vs 36% for Syst'N). So, despite being less accurate, the predictions of Syst'N
422 were reasonable for perennial crops given it uses simpler equations than STICS..

423 Comparing Syst'N's lucerne predictions to those of CropSyst highlights the difficulty in applying a model
424 to contrasting environments using a single set of crop parameters. Using CropSyst, Confalonieri and
425 Bechini (2013) obtained an rRMSE of 3-6% for predictions of total aboveground biomass after
426 calibrating crop parameters that partly considered local conditions, especially for dormancy and
427 development temperature. We used a submodel of CropSyst for dormancy in Syst'N, but due to the
428 small amount of measurement data over the period, Syst'N's parameters were not calibrated to local
429 conditions of the datasets, which may explain the lower rRMSE for Syst'N. Moreover, lucerne
430 parameters were calibrated based on a variety grown in northern France, and the influence of
431 decreasing photoperiod varies among lucerne varieties. Syst'N represents the effect of photoperiod
432 on crops indirectly using two fixed calendar dates that can stop growth during a period with a short
433 photoperiod. Unfortunately, this study could not test the ability of Syst'N and this parameter set to
434 simulate southern varieties of lucerne, which may start growing earlier in the spring.

435

436 **5. Conclusion**

437 Legumes can play a key role by providing N to cropping systems and thus enhancing the N self-
438 sufficiency of farms (Cellier et al., 2016; Vertès et al., 2010; 2015). Therefore, farmers and their advisers
439 increasingly attempt to insert legume species into their crop sequences; besides the effects on their
440 crop production that they can observe, they need results for and knowledge about N fluxes that they
441 cannot assess themselves, such as N supply to subsequent crops and emissions to the environment.
442 Adapting Syst’N to several legumes with different crop cycles and BNF capacities but keeping it
443 relatively simple was both a challenge and an expected development, given its utility to non-researcher
444 users.

445 We chose and combined equations from research crop models such as STICS and CropSyst to maintain
446 the simplicity of Syst’N for end users. Although some BNF equations required many parameters, users
447 need to supply only one additional parameter (flowering date) if they want to predict BNF. Based on
448 the literature, experiments and optimisation, we developed a set of parameters that provided
449 satisfactory predictions of legume aboveground biomass and N content. In contrast, Syst’N predicted
450 soil N content less well, mainly due to temporal shifts. Nevertheless, they remain relevant and are an
451 important prerequisite for simulating N emissions that are of particular interest to users. The next step
452 will be to improve predictions of soil N and water content, to evaluate the accuracy of Syst’N
453 predictions of N emissions (i.e. nitrate leaching, nitrous oxide emissions) and to do so at the scale of a
454 cropping sequence that includes legumes.

455

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462

463 **References**

- 464 Al Majou, H., Bruand, A., Duval, O., Cousin, I., 2007. Variation of the water-retention properties of
465 soils: Validity of class-pedotransfer functions. *Comptes Rendus Geoscience* 339, 632–639.
466 <https://doi.org/10.1016/j.crte.2007.07.005>
- 467 Anglade, J., Billen, G., Garnier, J., 2015. Relationships for estimating N₂ fixation in legumes: incidence
468 for N balance of legume-based cropping systems in Europe. *Ecosphere* 6, 37.
469 <https://doi.org/10.1890/ES14-00353.1>
- 470 Baranger, A., Nayel, M.-L., Lecomte, C., Hanocq, E., Moussart, A., Lejeune-Henaut, I., 2010. Quel
471 impact des innovations génétiques pour lever les facteurs limitant la production du pois
472 protéagineux? *Innovations Agronomiques* 11, 59–78.
- 473 Beaudoin, N., Launay, M., Sauboua, E., Ponsardin, G., Mary, B., 2008. Evaluation of the soil crop
474 model STICS over 8 years against the “on farm” database of Bruyères catchment. *European*
475 *Journal of Agronomy* 29, 46–57. <https://doi.org/10.1016/j.eja.2008.03.001>
- 476 Brisson, N., Gary, C., Justes, E., Roche, R., Mary, B., Ripoche, D., Zimmer, D., Sierra, J., Bertuzzi, P.,
477 Burger, P., Bussièrè, F., Cabidoche, Y.M., Cellier, P., Debaeke, P., Gaudillère, J.P., Hénault, C.,
478 Maraux, F., Seguin, B., Sinoquet, H., 2003. An overview of the crop model stics. *European*
479 *Journal of Agronomy* 18, 309–332. [https://doi.org/10.1016/S1161-0301\(02\)00110-7](https://doi.org/10.1016/S1161-0301(02)00110-7)
- 480 Brisson, N., Launay, M., Mary, B., Beaudoin, N., 2009. Conceptual basis, formalisations and
481 parameterization of the STICS crop model. Editions Quae.
- 482 Brisson, N., Mary, B., Ripoche, D., Hélène Jeuffroy, M., Ruget, F., Nicoullaud, B., Gate, P., Devienne-
483 Barret, F., ANTONIOLETTI, R., Durr, C., Richard, G., Beaudoin, N., Recous, S., Tayot, X., Plenet,
484 D., Cellier, P., Machet, J.-M., Marc Meynard, J., Delécolle, R., 1998. STICS: a generic model for
485 the simulation of crops and their water and nitrogen balances. I. Theory and
486 parameterization applied to wheat and corn. *Agronomie* 18, 311–346.
487 <https://doi.org/10.1051/agro:19980501>

488 Carof, M., Godinot, O., Ridier, A., 2019. Diversity of protein-crop management in western France.
489 Agronomy for Sustainable Development 39, 15. <https://doi.org/10.1007/s13593-019-0561-7>

490 Casal, L., Durand, P., Akkal-Corfini, N., Benhamou, C., Laurent, F., Salmon-Monviola, J., Ferrant, S.,
491 Probst, A., Probst, J.-L., Vertès, F., 2019. Reduction of stream nitrate concentrations by land
492 management in contrasted landscapes. Nutrient Cycling in Agroecosystems 114, 1–17.
493 <https://doi.org/10.1007/s10705-019-09985-0>

494 Cellier, P., Odoux, J.F., Thiébeau, P., Vertès, F., 2016. Ce que les légumineuses fourragères et
495 prairiales apportent à l'environnement. Fourrages 226, 87–94. [https://hal.archives-
496 ouvertes.fr/hal-01461116](https://hal.archives-ouvertes.fr/hal-01461116)

497 Cellier, P., Schneider, A., Thiébeau, P., Vertes, F., 2015. Impacts environnementaux de l'introduction
498 de légumineuses dans les systèmes de production, in: Les Légumineuses Pour Des Systèmes
499 Agricoles et Alimentaires. Editions Quae, 297-338.

500 Cerf, M., Jeuffroy, M.-H., Prost, L., Meynard, J.-M., 2012. Participatory design of agricultural decision
501 support tools: taking account of the use situations. Agronomy for Sustainable Development
502 32, 899–910. <https://doi.org/10.1007/s13593-012-0091-z>

503 Cernay, C., Makowski, D., Lescoat, P., Pelzer, E., 2017. Comparaison des performances de différentes
504 espèces de légumineuses à graines. Innovations Agronomiques 60, 1–21.
505 <https://doi.org/10.15454/1.513850748029309E12>

506 Confalone, A.E., 2008. Crecimiento y desarrollo del cultivo del haba (*Vicia faba* L.). Parametrización
507 del submodelo de fenología de cropgro-fababean. PhD Univ Santiago de Compostela. 175 p.

508 Confalonieri, R., Bechini, L., 2013. A preliminary evaluation of the simulation model CropSyst for
509 alfalfa. European Journal of Agronomy 21, 223–237.
510 <https://doi.org/10.1016/j.eja.2003.08.003>

511 Constantin, J., Dürr, C., Tribouillois, H., Justes, E., 2015a. Catch crop emergence success depends on
512 weather and soil seedbed conditions in interaction with sowing date: A simulation study

513 using the SIMPLE emergence model. *Field Crops Research* 176, 22–33.
514 <https://doi.org/10.1016/j.fcr.2015.02.017>

515 Constantin, J., Le Bas, C., Justes, E., 2015b. Large-scale assessment of optimal emergence and
516 destruction dates for cover crops to reduce nitrate leaching in temperate conditions using
517 the STICS soil–crop model. *European Journal of Agronomy* 69, 75–87.
518 <https://doi.org/10.1016/j.eja.2015.06.002>

519 Corre-Hellou, G., Brisson, N., Launay, M., Fustec, J., Crozat, Y., 2007. Effect of root depth penetration
520 on soil nitrogen competitive interactions and dry matter production in pea–barley intercrops
521 given different soil nitrogen supplies. *Field Crops Research* 103, 76–85.
522 <https://doi.org/10.1016/j.fcr.2007.04.008>

523 Crews, T.E., Peoples, M.B., 2004. Legume versus fertilizer sources of nitrogen: ecological tradeoffs
524 and human needs. *Agriculture, Ecosystems & Environment* 102, 279–297.
525 <https://doi.org/10.1016/j.agee.2003.09.018>

526 Dumont, B., Vancutsem, F., Seutin, B., Bodson, B., Destain, J.-P., Destain, M.-F., 2012. Simulation de la
527 croissance du blé à l’aide de modèles écophysiologicals: Synthèse bibliographique des
528 méthodes, potentialités et limitations. *Biotechnologie, Agronomie, Société et Environnement*
529 16, 376–386.

530 Dupas, R., Parnaudeau, V., Reau, R., Jeuffroy, M.-H., Durand, P., Gascuel-Oudou, C., 2015. Integrating
531 local knowledge and biophysical modeling to assess nitrate losses from cropping systems in
532 drinking water protection areas. *Environmental Modelling & Software* 69, 101–110.
533 <https://doi.org/10.1016/j.envsoft.2015.03.009>

534 Falconnier, G.N., Journet, E.-P., Bedoussac, L., Vermue, A., Chlébowski, F., Beaudoin, N., Justes, E.,
535 2019. Calibration and evaluation of the STICS soil-crop model for faba bean to explain
536 variability in yield and N₂ fixation. *European Journal of Agronomy* 104, 63–77.
537 <https://doi.org/10.1016/j.eja.2019.01.001>

538 Ferrant, S., 2009. Modélisation agro-hydrologique des transferts de nitrates à l'échelle des bassins
539 versants agricoles gascons (Thèse de Doctorat). PhD Université Toulouse III-Paul Sabatier.

540 Générmont, S., Cellier, P., 1997. A mechanistic model for estimating ammonia volatilization from
541 slurry applied to bare soil. *Agricultural and Forest Meteorology* 88, 145–167.
542 [https://doi.org/10.1016/S0168-1923\(97\)00044-0](https://doi.org/10.1016/S0168-1923(97)00044-0)

543 Guinet, M., Nicolardot, B., Revellin, C., Durey, V., Carlsson, G., Voisin, A.-S., 2018. Comparative effect
544 of inorganic N on plant growth and N₂ fixation of ten legume crops: towards a better
545 understanding of the differential response among species. *Plant and Soil* 432, 207–227.
546 <https://doi.org/10.1007/s11104-018-3788-1>

547 Gupta, S.C., Larson, W.E., 1979. Estimating soil water retention characteristics from particle size
548 distribution, organic matter percent, and bulk density. *Water Resources Research* 15, 1633–
549 1635. <https://doi.org/10.1029/WR015i006p01633>

550 Hénault, C., Bizouard, F., Laville, P., Gabrielle, B., Nicoulaud, B., Germon, J.C., Cellier, P., 2005.
551 Predicting in situ soil N₂O emission using NOE algorithm and soil database. *Global Change*
552 *Biology* 11, 115–127. <https://doi.org/10.1111/j.1365-2486.2004.00879.x>

553 Herridge, D.F., Marcellos, H., Felton, W.L., Turner, G.L., Peoples, M.B., 1995. Chickpea increases soil-
554 N fertility in cereal systems through nitrate sparing and N₂ fixation. *Soil Biol. Biochem.* 27,
555 545–551

556 Iannucci, A., Terribile, M.R., Martiniello, P., 2008. Effects of temperature and photoperiod on
557 flowering time of forage legumes in a Mediterranean environment. *Field Crops Research* 106,
558 156–162. <https://doi.org/10.1016/j.fcr.2007.11.005>

559 Jeuffroy, M.-H., Recous, S., 1999. Azodyn: a simple model simulating the date of nitrogen deficiency
560 for decision support in wheat fertilization. *European Journal of Agronomy* 10, 129–144.
561 [https://doi.org/10.1016/S1161-0301\(98\)00059-8](https://doi.org/10.1016/S1161-0301(98)00059-8)

562 Justes, E., Thiébeau, P., Avice, J., Lemaire, G., Volenec, J.J., Ourry, A., 2002. Influence of summer
563 sowing dates, N fertilization and irrigation on autumn VSP accumulation and dynamics of

564 spring regrowth in alfalfa (*Medicago sativa* L.). *Journal of Experimental Botany* 53, 111–121.
565 <https://doi.org/10.1093/jxb/53.366.111>

566 Justes, E., Thiébeau, P., Cattin, G., Larbre, D., Nicolardot, B., 2001. Libération d’azote après le
567 retournement de luzerne. Un effet sur deux campagnes. *Perspectives Agricoles* 264, 22-28.

568 Kelner, D.J., Vessey, J.K., Entz, M.H., 1997. The nitrogen dynamics of 1-, 2- and 3-year stands of alfalfa
569 in a cropping system. *Agriculture, Ecosystems & Environment* 64, 1–10.
570 [https://doi.org/10.1016/S0167-8809\(97\)00019-4](https://doi.org/10.1016/S0167-8809(97)00019-4)

571 Lemaire, G., Cruz, P., Gosse, G., Chartier, M., 1985. Etude des relations entre la dynamique de
572 prélèvement d’azote et la dynamique de croissance en matière sèche d’un peuplement de
573 luzerne (*Medicago sativa* L.). *Agronomie* 5, 685–692.

574 Liu, Y., Wu, L., Baddeley, J.A., Watson, C.A., 2011. Models of biological nitrogen fixation of legumes. A
575 review. *Agronomy Sust. Developm.* 31, 155–172. <https://doi.org/10.1051/agro/2010008>

576 Machet, J., Recous, S., Jeuffroy, M., Mary, B., Nicolardot, B., Parnaudeau, V., 2004. A dynamic version
577 of the predictive balance sheet method for fertiliser N advice. Presented at the Controlling
578 nitrogen flows and losses 12th Nitrogen Workshop, pp. 191–193.

579 Machet, J.-M., Dubrulle, P., Damay, N., Duval, R., Julien, J.-L., Recous, S., 2017. A Dynamic Decision-
580 Making Tool for Calculating the Optimal Rates of N Application for 40 Annual Crops While
581 Minimising the Residual Level of Mineral N at Harvest. *Agronomy* 7, 73.
582 <https://doi.org/10.3390/agronomy7040073>

583 Magrini, M.-B., Anton, M., Cholez, C., Corre-Hellou, G., Duc, G., Jeuffroy, M.-H., Meynard, J.-M.,
584 Pelzer, E., Voisin, A.-S., Walrand, S., 2016. Why are grain-legumes rarely present in cropping
585 systems despite their environmental and nutritional benefits? Analyzing lock-in in the French
586 agrifood system. *Ecological Economics* 126, 152–162.
587 <https://doi.org/10.1016/j.ecolecon.2016.03.024>

588 Mauriès, M., 2003. La luzerne: culture, récolte, conservation, utilisation. France Agricole Editions.

589 Muller, J.C., Denys, D., Thiébeau, P., 1993. Présence de légumineuses dans la succession de cultures:
590 luzerne et pois cultivés seuls ou en association, influence sur la dynamique de l'azote. *In*
591 *Matières organiques et agricultures*. Eds J. Decroux & J.C. Ignazi, 83-92.

592 Naudin, C., 2009. Nutrition azotée des associations Pois-Blé d'hiver (*Pisum sativum* L.–*Triticum*
593 *aestivum* L.): Analyse, modélisation et propositions de stratégies de gestion.

594 Nitschelm L, Parnaudeau V, Vertès F, van des Werf HMG, Corson MS, Viaud V, Aubin J, Walter C
595 (2018). Improving LCA estimates of nitrogen emissions during crop rotations at the scale of
596 an agricultural territory, *Environmental Science & Technology*, 52, 1330-1338.

597 Pampana, S., Masoni, A., Mariotti, M., Ercoli, L., Arduini, I., 2018. Nitrogen fixation of grain legumes
598 differs in response to nitrogen fertilisation. *Experimental Agriculture* 54, 66–82.
599 <https://doi.org/10.1017/S0014479716000685>

600 Parnaudeau, V., Reau, R., Dubrulle, P., 2012. Un outil d'évaluation des fuites d'azote vers
601 l'environnement à l'échelle du système de culture: le logiciel Syst'N. *Innovations*
602 *agronomiques* 21, 59–70.

603 Plaza-Bonilla, D., Nolot, J.-M., Raffailac, D., Justes, E., 2015. Cover crops mitigate nitrate leaching in
604 cropping systems including grain legumes: Field evidence and model simulations. *Agriculture,*
605 *Ecosystems & Environment* 212, 1–12. <https://doi.org/10.1016/j.agee.2015.06.014>

606 Prost, L., Cerf, M., Jeuffroy, M.-H., 2012. Lack of consideration for end-users during the design of
607 agronomic models. A review. *Agronomy for Sustainable Development* 32, 581–594.
608 <https://doi.org/10.1007/s13593-011-0059-4>

609 Rawls, W.J., Gish, T.J., Brakensiek, D.L., 1991. Estimating Soil Water Retention from Soil Physical
610 Properties and Characteristics, in: Stewart, B.A. (Ed.), *Advances in Soil Science: Volume 16*.
611 Springer New York, New York, NY, pp. 213–234. [https://doi.org/10.1007/978-1-4612-3144-](https://doi.org/10.1007/978-1-4612-3144-8_5)
612 [8_5](https://doi.org/10.1007/978-1-4612-3144-8_5)

613 Roberts, E., Summerfield, R.J., 1987. Measurement and prediction of flowering in annual crops, in:
614 Atherton, J.G. (Ed.), Manipulation of Flowering. Butterworth-Heinemann, 17–50.
615 <https://doi.org/10.1016/B978-0-407-00570-9.50007-7>

616 Robin, C., Corbel, G., Guinchard, M., Bazard, C., Guckert, A., 1999. Comportement hivernal et
617 repousse printanière du trèfle blanc: importance du cultivar. Fourrages 157, 21–32.
618 <https://hal.inrae.fr/hal-02694767>

619 Saxton, K.E., Rawls, W.J., 2006. Soil Water Characteristic Estimates by Texture and Organic Matter for
620 Hydrologic Solutions. Soil Science Society of America Journal 70, 1569–1578.
621 <https://doi.org/10.2136/sssaj2005.0117>

622 Schneider, A., Huyghe, C., 2015. Les légumineuses pour des systèmes agricoles et alimentaires.
623 Editions Quae. <https://hal.inrae.fr/hal-02800648>

624 Schneider A., Perrot T., Perrin A.-S., Le Gall C. (2019). Pre-cropping effects from grain legumes on
625 wheat and oilseed rape: nitrogen fluxes and productivity. European Conference on Crop
626 Diversification, Budapest, Hungary.18-21 September 2019.

627 Schneider A., Perrot T., Le Gall C., Perrin A.-S., Pontet C., Nourry F., Lacotte J.-C. (2021). Bénéfices et
628 risques liés aux flux azotés du segment de succession « précédent-suivant » avec pois,
629 féverole, lentille, colza et blé, RFL3, Rencontres Francophones des Légumineuses 3ème
630 édition, Angers (Webinars), 24-25 février 2021.

631 Schott, C., Mignolet, C., Meynard, J.-M., 2010. Les oléoprotéagineux dans les systèmes de culture :
632 évolution des assolements et des successions culturales depuis les années 1970 dans le
633 bassin de la Seine. <https://doi.org/10.1051/ocl.2010.0334>

634 Schoving, C., Stöckle, C.O., Colombet, C., Champolivier, L., Debaeke, P., Maury, P., 2020. Combining
635 Simple Phenotyping and Photothermal Algorithm for the Prediction of Soybean Phenology:
636 Application to a Range of Common Cultivars Grown in Europe. Frontiers in Plant Science 10,
637 1755. <https://doi.org/10.3389/fpls.2019.01755>

638 Schoving, C., Champolivier, L., Maury, P., Debaeke, P., 2022. Combining multi-environment trials and
639 crop simulation to evaluate the potential of early plantings of soybean crop in southern
640 France. *European Journal of Agronomy* **133**, 126439.
641 <https://doi.org/10.1016/j.eja.2021.126439>

642 Strullu, L., Beaudoin, N., de Cortàzar Atauri, I.G., Mary, B., 2014. Simulation of Biomass and Nitrogen
643 Dynamics in Perennial Organs and Shoots of *Miscanthus × Giganteus* Using the STICS Model.
644 *BioEnergy Research* 7, 1253–1269. <https://doi.org/10.1007/s12155-014-9462-4>

645 Strullu, L., Beaudoin, N., Thiébeau, P., Julier, B., Mary, B., Ruget, F., Ripoche, D., Rakotovololona, L.,
646 Louarn, G., 2020. Simulation using the STICS model of C&N dynamics in alfalfa from sowing
647 to crop destruction. *European Journal of Agronomy* 112, 125948.
648 <https://doi.org/10.1016/j.eja.2019.125948>

649 Thiébeau, P., 2019a. Mesurer l'épaisseur des résidus à la surface d'un sol pour estimer leur biomasse.
650 *Cahiers Agricultures* 28, 11. <https://doi.org/10.1051/cagri/2019011>

651 Thiebeau, P., 2019b. Relation entre taux de couverture du sol et biomasse de résidus de cultures: une
652 simplification prédictive est envisageable. *Cahiers Agricultures* 28, 30.
653 <https://doi.org/10.1051/cagri/2019031>

654 Thiébeau, P., Beaudoin, N., Justes, E., Allirand, J.M., Lemaire, G., 2011. Radiation use efficiency and
655 shoot:root dry matter partitioning in seedling growths and regrowth crops of lucerne
656 (*Medicago sativa* L.) after spring and autumn sowings. *European Journal of Agronomy* 35,
657 255–268. <https://doi.org/10.1016/j.eja.2011.07.002>

658 Thiébeau, P., Larbre, D., Usunier, J., Cattin, G., Parnaudeau, V., Justes, E., 2004. Effets d'apports de
659 lisier de porcs sur la production d'une luzerne et la dynamique de l'azote du sol. *Fourrages*
660 180, 511-525. <https://hal.inrae.fr/hal-02676239>

661 Thiébeau, P., Parnaudeau, V., Guy, P., 2003. Quel avenir pour la luzerne en France et en Europe?
662 *Courrier de l'environnement de l'INRA* 49, 29-46.

663 Tóth, B., Weynants, M., Nemes, A., Makó, A., Bilas, G., Tóth, G., 2015. New generation of hydraulic
664 pedotransfer functions for Europe. *European Journal of Soil Science* 66, 226–238.
665 <https://doi.org/10.1111/ejss.12192>

666 Undersander, D., Vassalotti, P., Cosgrove, D., 1997. *Alfalfa germination & growth*. University of
667 Wisconsin--Extension, Cooperative Extension.

668 Vertès, F., Jeuffroy, M.-H., Justes, E., Thiébeau, P., Corson, M., 2010. Connaître et maximiser les
669 bénéfices environnementaux liés à l'azote chez les légumineuses, à l'échelle de la culture, de
670 la rotation et de l'exploitation. *Innovations Agronomiques* 11, 25-44.

671 Vertès, F., Jeuffroy, M.-H., Louarn, G., Voisin, A.-S., Justes, E., 2015. Légumineuses et prairies
672 temporaires: des fournitures d'azote pour les rotations. *Fourrages* 223, 221–232.

673 Vocanson, A., 2006. Ex ante assessment of varietal innovations in winter pea (*Pisum sativum* L.):
674 Modelling approach at the plot and farm scales. PhD INAPG (AgroParisTech).

675 Voisin, A.-S., Cellier, P., Jeuffroy, M.-H., 2015. Fonctionnement de la symbiose fixatrice de N₂ des
676 légumineuses à graines: Impacts Agronomiques et Environnementaux. *Innovations
677 agronomiques* 43, 139–160.

678 Voisin, A.-S., Guéguen, J., Huyghe, C., Jeuffroy, M.-H., Magrini, M.-B., Meynard, J.M., Mougel, C.,
679 Pellerin, S., Pelzer, E., 2013. Les légumineuses dans l'Europe du XXI^e siècle: Quelle place dans
680 les systèmes agricoles et alimentaires actuels et futurs? Quels nouveaux défis pour la
681 recherche? *Innovations Agronomiques* 30, 283–312.

682 Voisin, A.-S., Salon, C., Munier-Jolain, N.G., Ney, B., 2002. Effect of mineral nitrogen on nitrogen
683 nutrition and biomass partitioning between the shoot and roots of pea (*Pisum sativum* L.).
684 *Plant and Soil* 242, 251–262. <https://doi.org/10.1023/A:1016214223900>

685 Wivstad, M., Mårtensson, A.M., Ljunggren, H.D., 1987. Field measurement of symbiotic nitrogen
686 fixation in an established lucerne ley using ¹⁵N and an acetylene reduction method. *Plant and
687 Soil* 97, 93–104. <https://doi.org/10.1007/BF02149828>

688 Xie, K., Li, X., He, F., Zhang, Y., WAN, L., David, B.H., Wang, D., Qin, Y., Gamal, M.A.F., 2015. Effect of
689 nitrogen fertilization on yield, N content, and nitrogen fixation of alfalfa and smooth
690 bromegrass grown alone or in mixture in greenhouse pots. *Journal of Integrative Agriculture*
691 14, 1864–1876. [https://doi.org/10.1016/S2095-3119\(15\)61150-9](https://doi.org/10.1016/S2095-3119(15)61150-9)
692 Yang, J.M., Yang, J.Y., Liu, S., Hoogenboom, G., 2014. An evaluation of the statistical methods for
693 testing the performance of crop models with observed data. *Agricultural Systems* 127, 81–
694 89. <https://doi.org/10.1016/j.agsy.2014.01.008>
695
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Table 1. Experimental data used to parameterise Syst'N

Species	Dataset	Total site-year-management units with available measurements						References
		Total	Leaf area index	Aboveground biomass	Aboveground N content	N fixed	Soil N content	
Lucerne (<i>Medicago sativa</i> L.)	Calibration	9	9	9	9		3	Thiébeau et al., 2011
	Evaluation	32	13	30	30		22	Justes et al., 2002, 2001; Lemaire et al., 1985; Thiébeau et al., 2011, 2004
Pea (<i>Pisum sativum</i> L.)	Calibration	6	4	6	6	6	6	Guinet et al., 2018
	Evaluation	12	8	12	9		12	Voisin et al., 2002; Corre-Hellou et al., 2009; Launay et al., 2009; Naudin, 2009; Pelzer et al., 2016
Faba bean (<i>Vicia faba</i> L.)	Calibration	6	4	6	6	6	6	Guinet et al., 2018
	Evaluation	3		3	3			Schneider et al., 2019; Schneider et al., 2021
Soya bean (<i>Glycine max</i> L.)	Calibration	17	12	16	16	16	4	Guinet et al., 2018
	Evaluation	9	0	9	9	0	0	Schoving et al., 2022

Corre-Hellou, G., Faure, M., Launay, M., Brisson, N., Crozat, Y., 2009. Adaptation of the STICS intercrop model to simulate crop growth and N accumulation in pea-barley intercrops. *Field Crops Research* 113, 72-81.

Launay, M., Brisson, N., Satger, S., Hauggaard-Nielsen, H., Corre-Hellou, G., Kasynova, E., Ruske, R., Jensen, E.S., Gooding, M.J., 2009. Exploring options for managing strategies for pea-barley intercropping using a modeling approach. *European Journal of Agronomy* 31, 85-98.

Naudin, C., 2009. Nutrition azotée des associations Pois-Blé d'hiver (*Pisum sativum* L.–*Triticum aestivum* L.): Analyse, modélisation et propositions de stratégies de gestion.

Pelzer, E., Bazot, M., Guichard, L., Jeuffroy, M.H., 2016. Crop Management Affects the Performance of a Winter Pea-Wheat Intercrop. *Agronomy Journal* 108, 1089-1100.

Voisin, A.S., Salon, C., Munier-Jolain, N.G., Ney, B., 2002. Quantitative effects of soil nitrate, growth potential and phenology on symbiotic nitrogen fixation of pea (*Pisum sativum* L.). *Plant and Soil* 243, 31-42.

Table 2. Indicators used to assess model accuracy. MBE=relative mean bias error, rMBE=relative mean bias error, RMSE=root mean square error, rRMSE=relative root mean square error, EF=model efficiency.

Indicator	Range of values	Ideal value	Satisfactory range
EF	$[-\infty ; 1]$	1	> 0.5
MBE	$[-\infty ; +\infty]$	0	
rMBE	$[-\infty ; +\infty]$	0	$ \text{rMBE} < 10\%$
RMSE	$[0 ; +\infty]$	0	
rRMSE	$[0 ; +\infty]$	0	$< 20\%$

Table 3. Parameters of the biological nitrogen fixation submodel added to Syst'N

Parameter name (unit)	Description	Crop							
		Lucerne		Faba bean		Pea		Soya bean	
		Initial	Regrowth	Winter	Spring	Winter	Spring	Type I	Type 00
CONC_N_NOD_SEUIL (kg N.ha ⁻¹ .mm ⁻¹ water)	Maximum soil nitrogen concentration threshold for nodule establishment	1.6	1.6	4.5	4.5	2.0	2.0	2.14	2.5
VIT_NOD (nodules.degree-day ⁻¹)	Nodule establishment rate	0.0037	0.0015	0.0068	0.0068	1.0	1.0	0.003	0.003
FIX_MAX_VEG (kg N.t DM ⁻¹)	Maximum nitrogen fixation capacity per t of dry matter produced before the grain-filling stage (DRG)	-	-	32	40	28	30	-	-
FIX_MAX_GR (kg N.t DM ⁻¹)	Maximum nitrogen fixation capacity per t of dry matter produced after the grain-filling stage (DRG)	-	-	17	17	9.5	9.5	-	-
FIX_MAX_CONST (kg N.day ⁻¹)	Constant capacity for maximum nitrogen fixation by the crop	6.0	6.0	-	-	-	-	6.0	6.0
TEMP_NOD_MIN (°C)	Minimum cardinal temperature for nodule activity	0	0	0	0	0	0	0	0
TEMP_NOD_OPT_B (°C)	Low optimal cardinal temperature for nodule activity	15	15	12	12	10	10	20	20
TEMP_NOD_OPT_H (°C)	High optimal cardinal temperature for nodule activity	25	25	25	25	25	25	36	36
TEMP_NOD_MAX (°C)	Maximum cardinal temperature for nodule activity	35	35	40	40	35	35	50	50
N_RAC_100 (kg N.ha ⁻¹ .cm ⁻¹ soil)	Nitrogen concentration threshold for full activity of nodules	0.12	0.12	1.6604	1.6604	0.3612	0.3612	1.6604	1.6604
N_RAC_0 (kg N.ha ⁻¹ .cm ⁻¹ soil)	Nitrogen concentration threshold that inhibits nodule activity	0.82	0.82	9.4759	9.4759	1.2058	1.2058	4.4804	4.4804
PROF_NOD_MAX (cm)	Maximum nodulation depth	40	40	30	30	30	30	40	40
ECARTTEMP_FIN_NOD (degree days)	Degree-day difference between the phenological reference stage and end of nodulation (FIN_NOD)	-100	0	550	550	1200	1316	700	530
ECARTTEMP_FSLA (degree days)	Degree-day difference between the phenological reference stage and final stage of seed abortion (FSLA)	300	300	800	1000	1200	1316	1000	750
ECARTTEMP_DRG (degree days)	Degree-day difference between the phenological reference stage and beginning of grain-filling (DRG)	0	0	250	250	200	150	400	400
SOMTEMP_DEBUT_FIX (degree days)	Sum of degree days from sowing to the beginning of fixation	250	0	340	310	235	235	320	320

Table 4. Statistical indicators of predictions of aboveground biomass, aboveground nitrogen (N) content and biological N fixation (BNF) for each legume crop with the evaluation dataset. EF=model efficiency, MBE=relative mean bias error, rMBE=relative mean bias error, RMSE=root mean square error, rRMSE=relative root mean square error.

Indicator	Prediction	Faba bean	Pea	Soya bean	Lucerne
EF	Biomass	0.82	0.90	0.71	0.59
	N content	0.87	0.87	0.48	0.61
	BNF	0.78	0.78	0.83	NA
	Soil water content	-28.71	0.44	-3.72	No data
	Soil N content	-0.2	0.49	-2.97	-0.76
MBE	Biomass	0.70	0.24	-0.65	0.43
	N content	16.21	0.08	31.52	5.63
	BNF	-9.24	-0.25	4.03	NA
	Soil water content	5.55	-0.75	5.21	No data
	Soil N content	-9.09	-5.93	-0.49	4.1
rMBE	Biomass	20%	5%	-13%	19%
	N content	16%	0%	23%	7%
	BNF	-13%	0%	6%	NA
	Soil water content	22%	-4%	22%	No data
	Soil N content	-14%	-11%	-1%	6%
RMSE	Biomass	0.94	1.28	1.59	1.08
	N content	20.45	32.99	59.83	36.19
	BNF	20.64	32.54	26.55	NA
	Soil water content	7.21	6.35	6.33	No data
	Soil N content	26.35	26.53	16.19	43.79
rRMSE	Biomass	27%	27%	33%	48%
	N content	20%	25%	44%	44%
	BNF	30%	45%	38%	NA
	Soil water content	29%	32%	27%	No data
	Soil N content	42%	50%	41%	69%

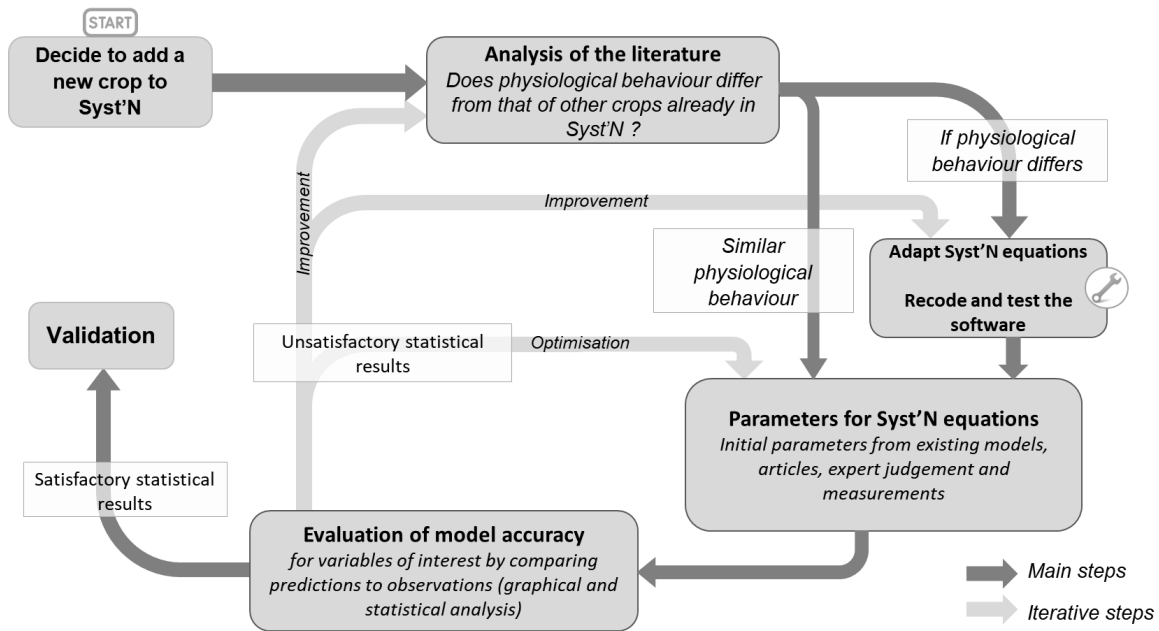
1 Table 4. Statistical indicators of predictions of aboveground biomass, aboveground nitrogen (N) content and
 2 biological N fixation (BNF) for each legume crop with the evaluation dataset. EF=model efficiency, MBE=relative
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	BNF	-13%	0%	6%	NA
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	N content	20%	25%	44%	44%
	BNF	30%	45%	38%	NA
	Soil water content	29%	32%	27%	No data
	Soil N content	42%	50%	41%	69%

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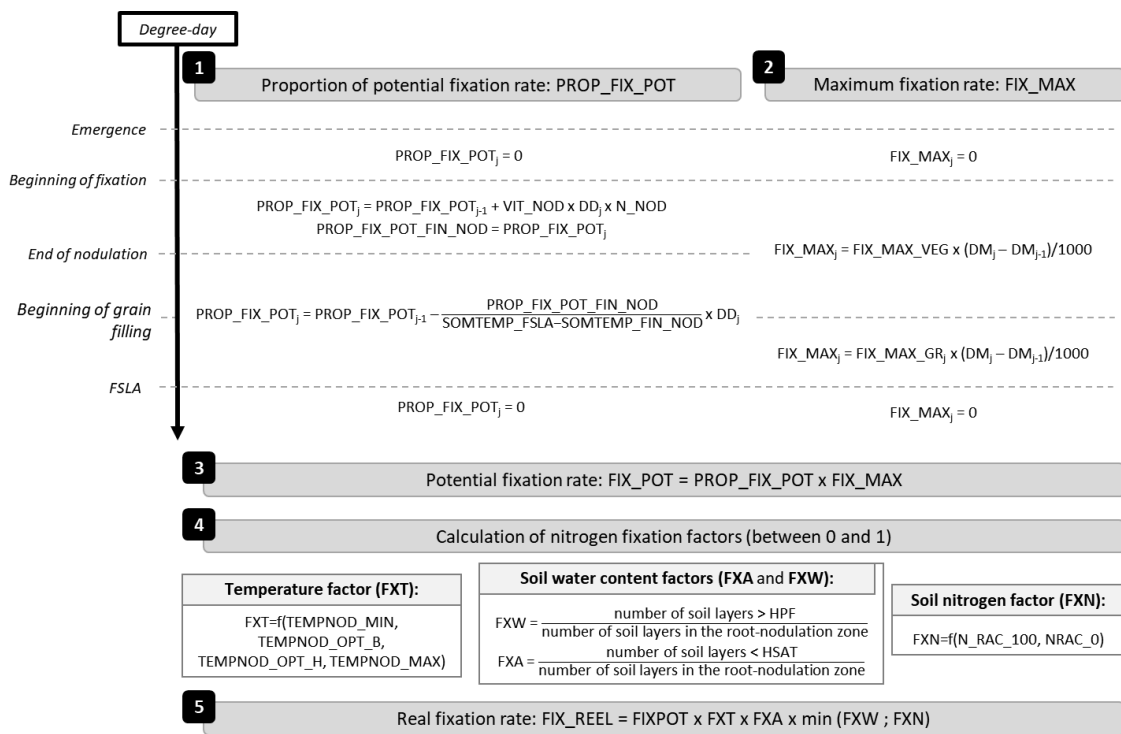
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9 Figure 1. Flowchart for adding a new crop to the Syst'N model

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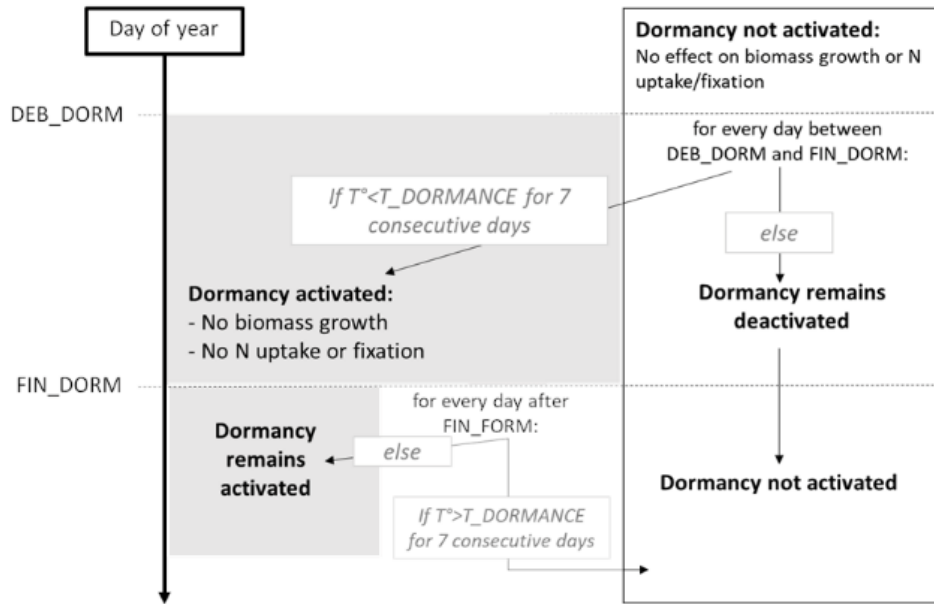


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12 Figure 2. Description of the biological nitrogen fixation (BNF) submodel added to Syst'N (adapted from the
13 STICS model). See Table 3 for definitions of variables not defined in the figure. Numbers in black boxes indicate
14 the order of the calculation steps.

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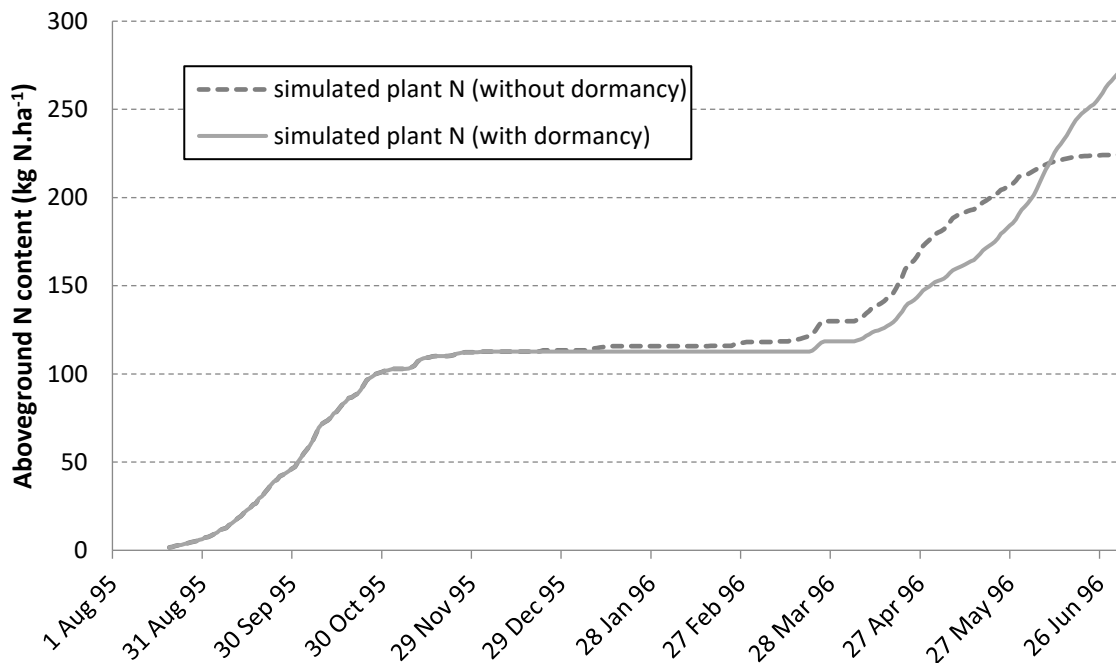
Parameter name (unit)	Description	Crop
DORMANCE (Boolean)	Indicates whether the crop can become dormant	Lucerne
DEB_DORM (date)	Beginning of the potential dormancy period	15 Nov
FIN_DORM (date)	End of the potential dormancy period	10 Mar
T_DORMANCE (°C)	Temperature below which crop dormancy begins, or above which dormancy ends, if maintained as a mean daily temperature for 7 consecutive days during the dormancy period, or after the dormancy period, respectively	5



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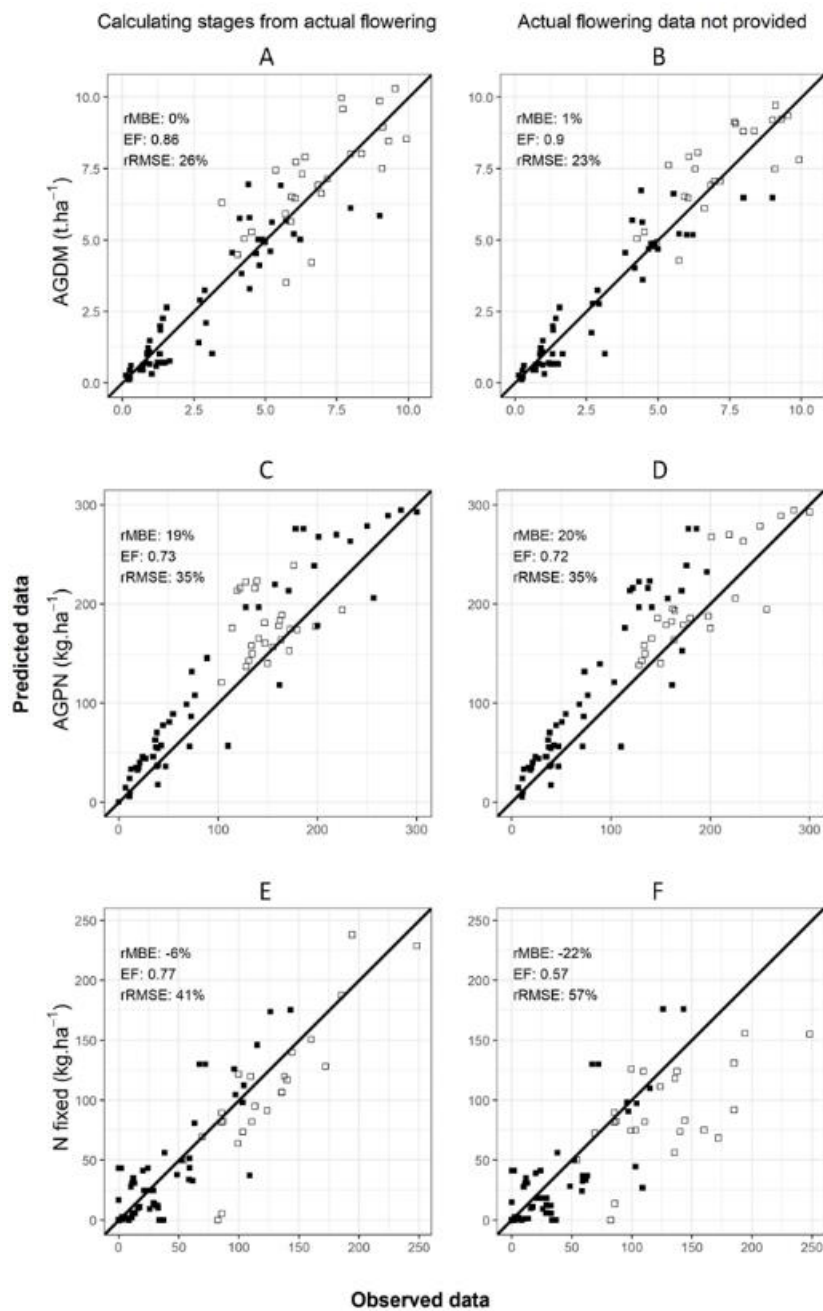
17 Figure 3. Description of the dormancy submodel added to Syst'N (adapted from that in the CropSyst model).

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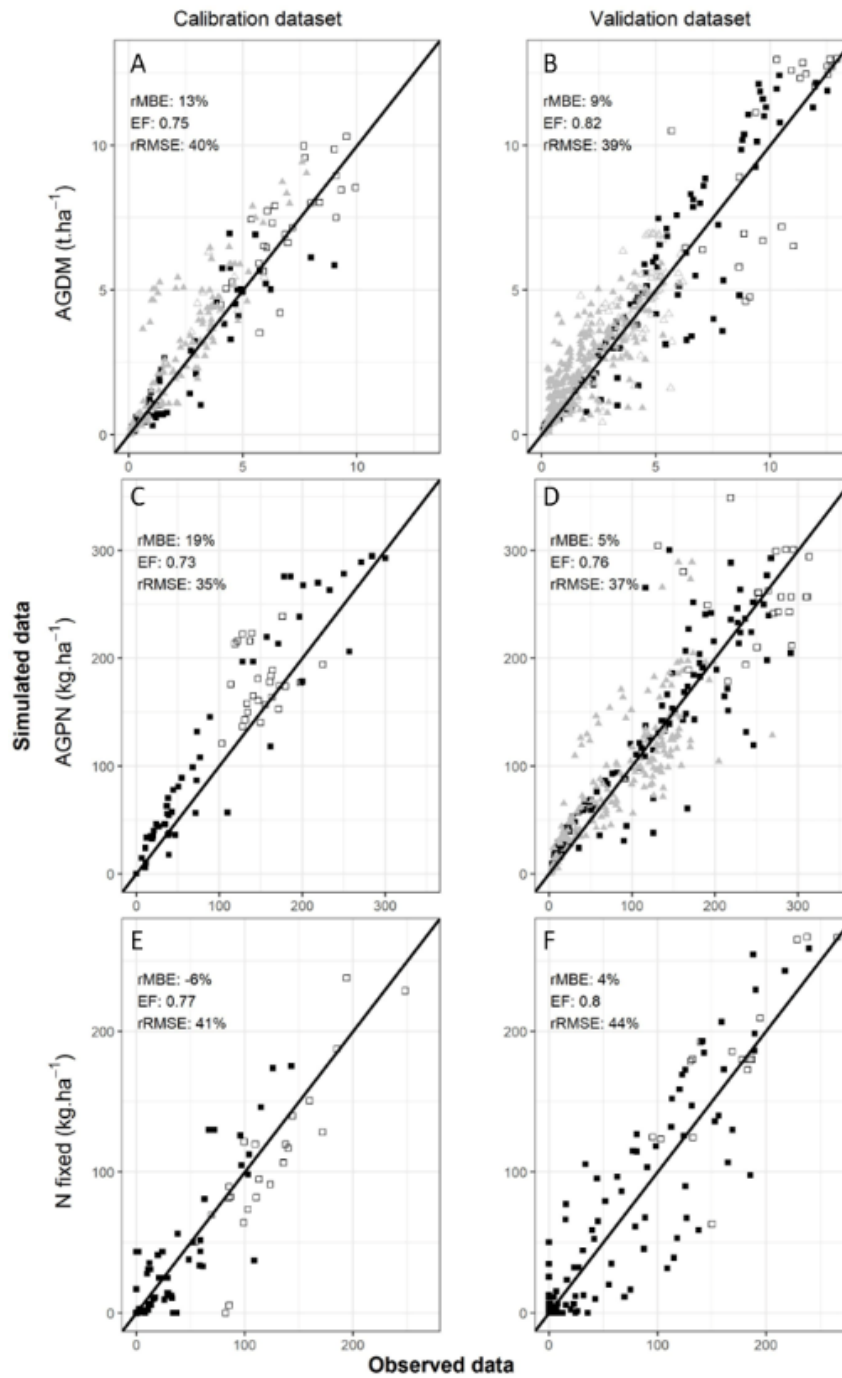
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20 Figure 4. Syst'N predictions of lucerne aboveground N content before and after adding equations to simulate
21 dormancy



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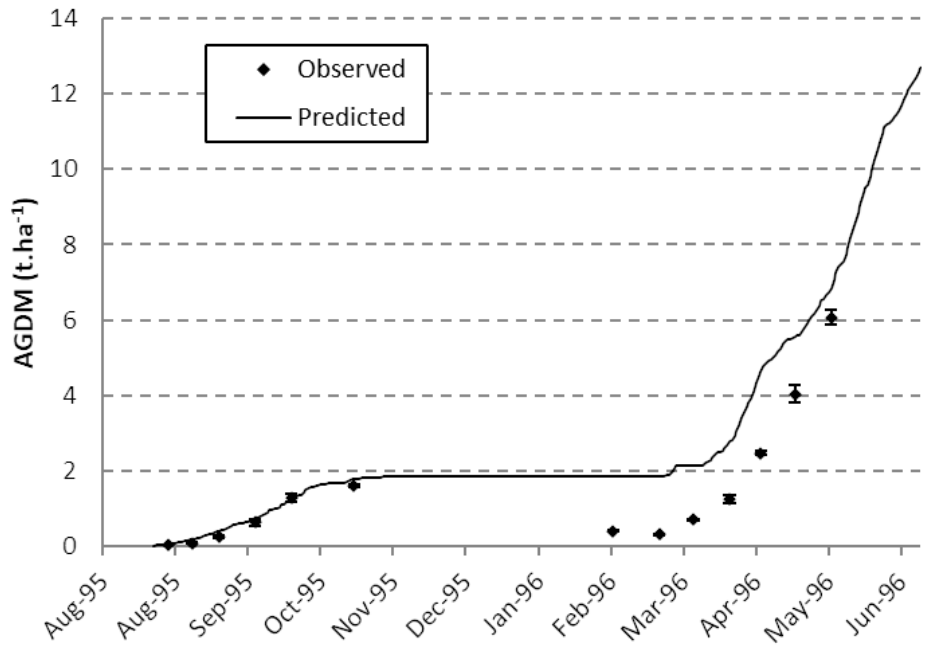
24 Figure 5. Comparison of aboveground dry matter (AGDM), aboveground plant nitrogen content
 25 (AGPN) and biological N fixation observed vs. that predicted by Syst'N with the calibration dataset,
 26 with or without calculating phenological stages from the actual flowering date. rMBE=relative mean
 27 bias error, EF=model efficiency, rRMSE=relative root mean square error. The black line is the 1:1 line.
 28 White symbols correspond to the end of the growing season. Black symbols correspond to earlier
 29 stages.



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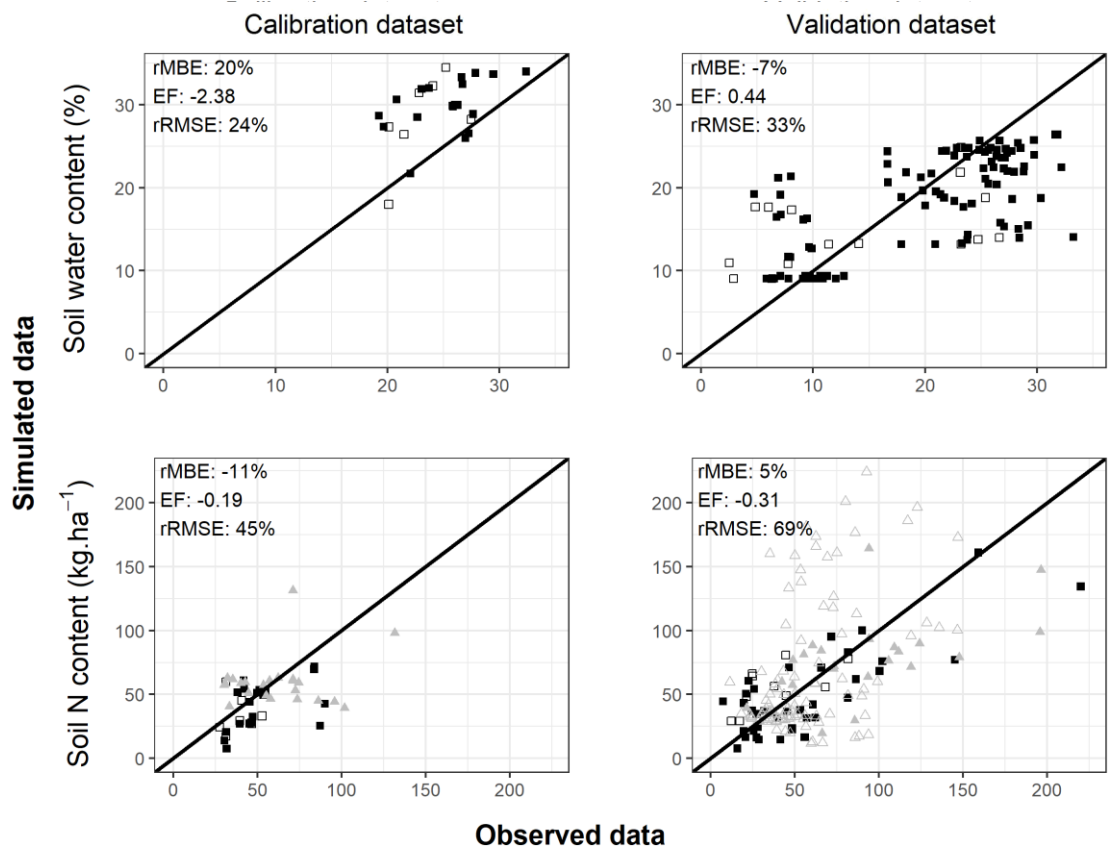
31 Figure 6. Comparison of aboveground dry matter (AGDM), aboveground plant nitrogen content (AGPN) and
 32 biological N fixation observed vs. that predicted by Syst'N with the calibration and evaluation datasets.
 33 rMBE=relative mean bias error, EF=model efficiency, rRMSE=relative root mean square error. The black line is
 34 the 1:1 line. White symbols correspond to the end of the growing season. Black symbols correspond to earlier
 35 stages.

36



37

38 Figure 7. Example of observed and predicted aboveground dry matter before and after dormancy using the
 39 evaluation dataset (site: Châlons-en-Champagne). Error bars indicate standard deviations.



40

41 Figure 8. Comparison of soil water content and soil nitrogen content observed and that predicted by Syst’N
 42 using calibration and evaluation datasets. rMBE=relative mean bias error, EF=model efficiency, rRMSE=relative
 43 root mean square error. The black line is the 1:1 line. White symbols correspond to the end of growing season.
 44 Black symbols correspond to earlier stages.

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