

Grassland-cropland rotation cycles in crop-livestock farming systems regulate priming effect potential in soils through modulation of microbial communities, composition of soil organic matter and abiotic soil properties

Marco Panettieri, Julien Guigue, Nicolas Chemidlin Prévost-Bouré, Mathieu Thevenot, Jean Lévêque, Cédric Le Guillou, Pierre-Alain Maron, Anne-Lise Santoni, Lionel Ranjard, Stéphane Mounier, et al.

► To cite this version:

Marco Panettieri, Julien Guigue, Nicolas Chemidlin Prévost-Bouré, Mathieu Thevenot, Jean Lévêque, et al.. Grassland-cropland rotation cycles in crop-livestock farming systems regulate priming effect potential in soils through modulation of microbial communities, composition of soil organic matter and abiotic soil properties. Agriculture, Ecosystems & Environment, 2020, 299, pp.106973. 10.1016/j.agee.2020.106973 . hal-03014052

HAL Id: hal-03014052 https://institut-agro-dijon.hal.science/hal-03014052v1

Submitted on 9 Mar 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- 1 Grassland-cropland rotation cycles in crop-livestock farming systems regulate priming effect
- 2 potential in soils through modulation of microbial communities, composition of soil organic
- 3 matter and abiotic soil properties.
- 4 Marco PANETTIERI^{a,b}, Julien GUIGUE^{a,c}, Nicolas CHEMIDLIN PREVOST-BOURÉ^d, Mathieu
- 5 THÉVENOT^a, Jean LÉVÊQUE^a, Cédric LE GUILLOU^d, Pierre-Alain MARON^d, Anne-Lise
- 6 SANTONI^a, Lionel RANJARD^d, Stéphane MOUNIER^e, Safya MENASSERI^f, Valérie VIAUD^f,
- 7 Olivier MATHIEU^a
- 8
- ^a Biogéosciences, UMR 6282 CNRS, Université Bourgogne Franche-Comté, 6 Boulevard Gabriel,
 21000 Dijon, France.
- ^b Present address: Museo Nacional de Ciencias Naturales (MNCN-CSIC), c/Serrano 115-B, 28006
 Madrid, Spain
- ^c Present address: Chair of Soil Science, Research Department Ecology and Ecosystem Management,
- 14 Technical University of Munich, 2 Emil-Ramann-Straße, 85354 Freising, Germany
- ^d INRA, UMR 1347 Agroécologie, F-21065 Dijon, France
- 16 ^e Université de Toulon, AMU, CNRS, IRD, MIO Toulon, CS 60583, 83041 Toulon Cedex 9
- 17 ^f INRA, UMR 1069 SAS, F-35042 Rennes, France
- 18
- 19 Corresponding author: Marco PANETTIERI ; ORCID : <u>https://orcid.org/0000-0003-4769-8955</u>
- 20 email : <u>marco.panettieri.soil@gmail.com</u>; <u>marco.panettieri@csic.es</u> Telephone +34665125289
- 21 Keywords: Soil Carbon; Water-Extractable Organic Carbon; Grassland; Cropland; Variance
- 22 Partitioning
- 23

24 Abstract

Soils can act as a carbon sink, and the chemical and biological transformation of vegetal litter into soil
organic matter (SOM) is widely influenced by land-use and other biogeochemical parameters.
However, the increase of new carbon inputs to soil has the potential to trigger the mineralization of
stabilized SOM, a process called priming effect.

The objective of this manuscript is to investigate, at a landscape scale (5km²), the factors influencing the susceptibility of SOM to priming effect. To achieve this objective, physical, chemical, and metagenomics analyses were conducted on 88 soil samples and successively combined with agronomical data and soil incubation for the quantification of carbon fluxes.

33 Variance partitioning models highlighted that priming effect is controlled by complex interactions of biotic and abiotic factors, which include soil chemistry, quality of SOM, shape and abundance of 34 microbial communities. Fluorescence properties of the dissolved organic matter has been found as a 35 36 strong descriptor for priming effect. Depending on the time of crop rotation devoted to grassland, two 37 different components leading to priming effect were identified. The introduction of grassland for 40-38 60% of the time of rotation achieved the lowest susceptibility to priming effect, and higher indexes of 39 microbial diversity, whereas higher or lower proportions of time of the rotation devoted to grassland resulted in an increase of priming effect and a decrease of bacterial evenness. 40

41

42 **1. Introduction**

43 A sustainable land management aims to guarantee adequate productivity in agriculture and, at the 44 same time, to provide several ecosystem services (Haines-Young and Potschin, 2010). Among 45 marketed ecosystem services, the production of food, fiber and biofuel are the most acknowledged 46 ones, but there are other non-marketed ecosystem services dealing with water and climate regulation, as well as cultural and aesthetic services (Swinton et al., 2007). Powlson et al. (2011) described soil 47 fertility and the accumulation of soil organic matter (SOM) among the most important services 48 provided by sustainable agricultural practices. There is a scientific consensus about the benefits 49 provided by higher SOM contents, and a major concern regarding the decline of SOM stocks 50 51 (Kirschbaum, 1995; Minasny et al., 2017; Six et al., 1998; Tiessen et al., 1994).

52 The contents of organic matter (OM) in a soil result from the differences between carbon (C) inputs, mainly as aboveground and belowground plant biomass or amendments, and outputs via SOM 53 54 decomposition by microorganisms, runoff and leaching of particulate and dissolved organic C. The actual stock of OM in a soil is thus regulated by transformation processes resulting to its stabilization, 55 or its mineralization. The microbial degradation of SOM is a function of its physical accessibility and 56 chemical composition (Kuzyakov and Blagodatskaya, 2015). The addition of exogenous organic 57 58 matter has been reported to alter SOM decomposition, either by boosting (Chen et al., 2014; Fontaine et al., 2003) or impeding it (Guenet et al., 2010) due to the so-called priming effect (PE) (Bingeman et 59 al., 1953; Kuzyakov et al., 2000). 60

A considerable effort is necessary to elucidate SOM dynamics and fluxes, especially PE, since they are 61 influenced by a plethora of biotic (interactions with plants, bacteria, fungi, micro fauna) (Tardy et al., 62 2015) and abiotic factors (content and type of minerals, water balance, atmospheric conditions) 63 (Dignac et al., 2017), acting from micro- to macro-scales (Kuzyakov and Blagodatskaya, 2015). The 64 fluxes of C in soil may differ strongly from one land-use to another, mediated by shifts in the 65 66 distribution of microbial communities and functionality, different types and amounts of C returned to soil, and various plant/soil interactions (Baumann et al., 2013; Dignac et al., 2017; Frank et al., 2002; 67 Tardy et al., 2015). Land-use can change SOM contents drastically, especially in the case of soils 68 69 devoted to intense anthropic activities (reclamation for urban or agricultural purposes) (Lal and 70 Stewart, 1990). The inclusion of grassland in crop rotation has been highlighted as a way to maintain or improve soil quality in agroecosystems (Glover et al., 2010; Lemaire et al., 2015) and to increase 71 72 soil C stocks (Smith, 2014; Solomon et al., 2007) because of the modification of both SOM accrual 73 and degradation dynamics (Panettieri et al., 2017). A considerable fraction of C fluxes resulting from 74 the priming effect is assumed to be driven by processes taking place in the rhizosphere (de Graaff et 75 al., 2014; Fontaine et al., 2003), and more extended and dense root networks are generally observed in grassland soils (Jackson et al., 1996; Panettieri et al., 2017). Therefore, the effects of grassland derived 76 77 C inputs on PE needs attention in order to estimate the potential effect of the introduction of grasslands into crop rotations on agroecosystems C balance. 78

79 Kuzyakov et al. (2000) described the mechanisms behind the increased C mineralization responsible of the "real priming effect", triggered by the addition of labile C sources and/or C-rich 80 81 rhizodeposition, among other factors. Recent studies have been focused on the PE produced by mining of nitrogen and preferential substrate use (Chen et al., 2014), energy starvation, microbial competition 82 and co-metabolisms (Louis et al., 2016; Tardy et al., 2015), or regulation by redox transformation of 83 soil minerals (Finley et al., 2018; Keiluweit et al., 2015). However, the quantification of the PE 84 85 mechanism is still limited by methodological uncertainties and by both spatial and temporal variability 86 in its occurrence (Bastida et al., 2019). Moreover, there are concerns about the accuracy of prediction 87 models for global C stocks in which the trigger of SOM mineralization provided by PE is not taken into account (Guenet et al., 2018). 88

Considering these scientific challenges, the present study has been designed to evaluate the effects of 89 land-use on PE at a landscape scale. Recent studies have been focused on the global assessment of PE 90 91 predictors (Bastida et al., 2019), but few studies have been targeted at the landscape scale (Craig et al., 92 2015; Helgason et al., 2014; Walz et al., 2017) and none of them included cropped soils. Unveiling the 93 magnitude and the dynamics of PE in agricultural landscapes is mandatory to calibrate land-use policies. The objective of the project was to identify the factors controlling PE, having a 94 95 comprehensive approach by integrating soil physical and chemical properties, indexes of soil 96 microbial communities, and agronomic practices in a 5-km² agricultural area in the region of Brittany 97 (France). The site is characterized by high density of pig farming and by the co-existence of mixed annual crops and grasslands (pastures or hayfields, being permanent or included in the rotations). We 98 hypothesized that (i) PE at the landscape scale is related to the community composition of soil 99 microorganisms interacting with the amount and type of OM, and that (ii) land-use management has a 100 101 significant impact on C fluxes associated with PE.

To test these hypotheses, soil samples were analyzed using a multidisciplinary approach in which soil
 physical, chemical, and biological variables were tested as descriptors of PE fluxes measured in
 incubation condition with addition of ¹³C-labeled wheat litter.

105

106 **2.** Materials and methods

107 2.1. Study area and soil sampling

Soil samples were collected from a 5 km² agricultural area, in the Brittany region (Kervidy-Naizin 108 109 catchment, N 47.99; W 2.84, France) in June 2013. About 85% of the surface of the sampling area is 110 devoted to agriculture. The crops covering the larger surfaces are maize and grassland (covering 30% 111 of the agricultural surface each), followed by other cereals (25%) and vegetables (10%). The 112 catchment is an Observatory for Research on the Environment related to agricultural watersheds (SOERE-RBV Critical Zones, www6.inra.fre/ore_agrhys). The climate is oceanic, with a yearly 113 average temperature of 11°C (1994-2013) and mean yearly rainfall of 830 mm. The elevation ranges 114 between 98 and 140 m with mild slopes oriented towards the Naizin river network. Coordinates of the 115 sampling points were distributed using a systematic triangular grid to integrate the maximum of the 116 117 spatial heterogeneity of the area.

118 Most soils are well-drained Cambisol (upslope), and some poorly drained Haplic Albeluvisols are 119 located in the foot-slope farms (IUSS Working Group WRB, 2015). Parent material consists of schist 120 and alluvial deposits. The soil texture, measured by standardized 5-fraction granulometry (NF X 31– 121 107, AFNOR, 2003), is silty-loam and relatively uniform among the whole area.

122 Surface composite soil samples were collected at a 0-15 cm depth with a 5 cm wide auger. The 123 composite soil samples were made by mixing individual soil cores into one homogeneous sample, sieved in the field ($\emptyset < 5$ mm), stored at 4°C during the transportation to the lab, and split into two 124 subsamples. The first subsample was air-dried and stored at 4°C for incubation and chemical analyses. 125 The other subsample was freeze-dried and stored at -20°C until molecular biology analyses were 126 performed. Soil drainage classes and Beven's topographic wetness indexes (Beven and Kirby, 1979) 127 were derived respectively from a detailed soil map (Walter and Curmi, 1998) and from a 20-m digital 128 elevation model. In the framework of the project, farmers were surveyed regarding farming practices, 129 crop rotations, number of cropped species, the quantity of mean annual C input by crop and grassland 130 residues, C input by animal manure as well as the quantity of mineral fertilizers applied each year, on 131 an 11-year period (Viaud et al., 2018). 132

133 In this study, 100 independent samples were collected from different farms and divided in five different land-use classes considering the frequency of grassland, expressed as percentage of the 134 135 rotation time devoted to grassland, as the classification parameter. Main cropped species other than 136 grassland were Maize (silage or grain) in monoculture, or in rotation with cereals (winter wheat, winter barley and triticale), rapeseed, and vegetables. Regarding the descriptors of crop rotation, the 137 number of cropped species, considering grassland a unique specie, was the other variable added to the 138 139 variance partitioning model, further variables were adding too much complexity to the study and thus discarded. The lower and upper limits of the land-use classification were permanently cropped soils 140 (PC n=42), and permanent grassland soils (PG n=7), respectively. Three land-uses were selected using 141 the different frequency of grassland adopted by farmers on the 11-year period before soil sampling: 142 143 low-frequency (LO, from 10 to 30% of time dedicated to grassland, n=13), mid-frequency (MI, 40-60% of time under grassland, n=13), and high-frequency (HI, 70-90% of time under grassland, n=13). 144 145 All the grassland farms were mown regularly to feed animals.

After this classification, 88 independent samples respected the criteria of classification, 12 samples
collected in wetland, wasteland land and orchards were not considered because of their low
representativeness of the area.

149

150 2.2. Soil physical and chemical analyses

Each measure of soil physical and chemical parameters is a calculated mean of three replicates. The mean weight diameter (MWD) of the air-dried soil aggregates was calculated using the aggregate fractionation method ISO/FDIS 10930 (AFNOR, 2012) derived from the "slow re-wetting" and soil wet sieving procedure described by Le Bissonnais (1996). Bulk density was calculated on a 0-15 cm surface soil layer, using an 8-cm diameter manual soil core sampler. Samples were weighed and dried at 105°C for 48h to determine gravimetric water content and bulk density.

157 The total organic C and N contents (TOC and TN) and C isotopic signatures (δ^{13} C) of the soil samples 158 were measured by dry combustion (NF ISO 10694 AFNOR, 1998, and NF ISO 13878, AFNOR 159 1995)) on an elemental analyzer linked to an isotopic-ratio mass spectrometer (EA-IRMS, VarioMicro 160 Cube linked to an Isoprime, Elementar). The stable carbon isotope ratios, ${}^{13}C/{}^{12}C$ expressed as $\delta^{13}C$, 161 are calculated against Vienna Pee Dee Belemnite (VPDB) standard.

162 Moreover, soils were analyzed for the concentrations of oxalate-extractable Si, Fe, Al (NF ISO 22036,

163 (AFNOR 2009), Cu (NF X 31- 120 on EDTA extracted samples (AFNOR 2003b), extractable Olsen's

P (NF ISO 11263, (AFNOR 1995b) and pH in a 1:5 water suspension (NF ISO 10390, (AFNOR 2005)).

- 166 Pressurized hot-water-extractable organic carbon (WEOC) was obtained using a solvent extractor (ASE200, Dionex Corporation, Sunnyvale, USA) on 5 g of soil, according to the method of Guigue et 167 168 al. (2014). The C concentration in the extracts was measured in a first aliquot by thermal oxidation and NDIR detection (TOC 5000A, Shimadzu), after the addition of 2M HCl (10 µl per ml of sample) to 169 170 remove inorganic C. The results of WEOC content were expressed as WEOC contribution to TOC content of soils. The UV-light absorbance of the extracts at 254 nm (UV₂₅₄) was measured with a 171 Jenway 6715 spectrophotometer and the specific ultraviolet absorbance (SUVA in 1 mg C⁻¹ cm⁻¹) was 172 calculated as an indicator of their content in aromatic moieties (Chin et al., 1994; Weishaar et al., 173 174 2003). Another aliquot of the sample was used to measure the pH, then freeze-dried and used for the determination of δ^{13} C of WEOC by EA-IRMS. 175
- 176

177 2.3. Excitation-emission matrices of fluorescence and PARAFAC modeling

178 For fluorescence measurements, triplicates of water-extracted samples ($n=3\times88$) were analyzed using the same settings as in Guigue et al. (2014), with excitation wavelengths ranging from 220 to 500 nm, 179 and emission wavelengths from 250 to 600 nm. The whole dataset corresponding to 264 data matrices 180 (excitation wavelength \times emission wavelength \times fluorescence intensity) was processed for PARAFAC 181 182 modeling using drEEM toolbox 0.2.0 (Murphy et al., 2014) with Matlab R2016a under Linux. The 183 inner-filter effect was corrected using absorbance spectra during the preprocessing steps. Rayleigh and Raman scatter peaks were excised and substituted with interpolated data using the *smootheem* function 184 185 of the drEEM toolbox. The fluorescence signal measured at excitation wavelengths shorter than 250 nm was noisy, exerting disproportionate leverage on the model, and were thus cut for all EEMs. Each 186 EEM was normalized to its total signal, so that the effect of sample intensity was removed before the 187

188 modeling phase (Murphy et al., 2013). The normalization was reversed after the modeling so that the 189 output values presented correspond to the concentration in the original sample. Outliers identified by 190 their high leverages (n=11) were removed before fitting the final model. A 3-component model 191 explaining 99.7% of the dataset, having a CORCONDIA of 43.4% and which was successfully splitvalidated was retained as the final model. The component contribution to each sample fluorescence 192 signal were exported as Fmax in Raman Unit (RU), and then normalized to the DOC concentration in 193 194 the measured aliquot so that the reported Fmax value is expressed in RU per C unit and serves as an 195 indicator of the composition of OM in the samples. The three components of fluorescence identified by the PARAFAC model, named CP1_(fluo), CP2_(fluo) and CP3_(fluo), comprised two excitation spectral 196 197 single emission (Fig. 1). The OpenFluor online peaks and а peak database (https://openfluor.lablicate.com) was consulted to match the spectral properties of obtained 198 199 components with previously identified fluorophores (Murphy et al., 2014).

200

201 2.4. Soil microbial communities

202 A characterization of communities of soil microorganisms was carried out after the pre-incubation and 203 just before incubation, following the method proposed by Tardy et al. (2015). For each soil sample, the 204 biomass was derived from the total quantity of crude DNA extracted from 1 g of lyophilized soil using 205 the ISO 10063 method (Dequiedt et al., 2011; Plassart et al., 2012). Prior to PCR amplification, DNA 206 was purified using a MinElute gel extraction kit (Oiagem Courtaboeuf, France) at INRA Genosol Platform (Dijon, France). Bacterial and fungal diversity were resolved by 454 pyrosequencing, 207 targeting respectively the 16S rRNAV3-V4 gene fragment (about 400 bases) and an 18S rRNA gene 208 209 fragment of about 350 bases. The primer sets were respectively: F479 (5'-210 CAGCMGCYGCNGTAANAC-3')/R888 (5'-CCGYCAATTCMTTTRAGT-3') (Terrat et al., 2012); and FR1 (5'-AICCATTCAATCGGTAIT-3')/FF390 (5'-CGATAACGAACGAGACCT-3') (Plassart 211 212 et al., 2012). A second PCR was run on purified products from first PCR, adding 10 based-pair multiplex identifiers (MID) at 5' position to the primers for subsequent sample identification. Final 213 purification and pyrosequencing were carried out on a GS Roche 454 sequencing system. 214

Bioinformatics analyses were carried out using the GnS-Pipe of the Genosol Platform (Terrat et al., 215 2012). Raw reads of 16S and 18S were sorted according to MID sequences, then filtered on their 216 217 length, number of ambiguities, and primer sequences. A homogenization procedure by random 218 selection close to the lowest dataset (2,886 and 3,286 high-quality sequences for 16S and 18S rRNA genes, respectively) was carried out to avoid biased comparison of the communities. The dereplicated 219 reads were aligned using infernal alignment and clustered at a 95% similarity threshold into 220 221 operational taxonomic units (OTU) by grouping rare reads to abundant ones and not counting differences in homopolymer lengths (Terrat et al., 2012). Single Singletons constituted by unclustered 222 reads detected only once were filtered based on the quality of their taxonomic assignment. Bacterial 223 (16S) and fungal (18S) OTUs were organized into contingency tables reporting the number of reads 224 225 for each OTU in each sample. In this study, bacterial and fungal richness were expressed as number of 226 OTUs, whereas Pielou's diversity index J' assessed the evenness of the species (Pielou, 1966).

227

228 2.5. Carbon mineralization and priming effect measurements

The mineralization of SOM was measured during an 80-day incubation experiment. For each sample, six hermetically sealed 150-ml plasma flasks were filled with 30 g of dry soil and then brought to 60% of water holding capacity (WHC). To avoid the overestimation of C mineralized in the first days due to the disturbance by soil manipulation and rewetting and to allow soils to reach a more stable state, microcosms were pre-incubated for three weeks at 20°C before starting the monitored phase of the incubation.

¹³C-labeled wheat residues (7 atom% ¹³C) originated from mature wheat plants (harvested 110 days after sowing) cultivated under controlled conditions (Groupe de Recherches Appliquées en Phytotechnologie, CEA Cadarache, France). More precisely, seeds of wheat (*Triticum aestivum* cv Caphorn) were germinated at 4°C in darkness. Plantlets were grown in a mix of sand (1/4) and perlite (3/4) in an air-tight chamber which allowed accurate regulation of atmospheric gas composition and environmental parameters. The plants were watered with half-diluted Hoagland's nutrient solution and CO₂ concentration was maintained at 380µl l⁻¹. The partial pressures of both ¹³CO₂ and ¹²CO₂ in the chamber were continuously monitored by Near Infrared Spectroscopy to determine ¹³C enrichment of
the CO₂. Regulation was achieved by automatic injection of pure (>99% atom% ¹³C) ¹³CO₂
(CortecNet, Paris, France).

The microcosms were either amended with 13 C-residues of wheat (5 mg g⁻¹ of dry weight soil), which 245 had been finely ground in a bead beater to obtain a powder (wheat residues $C_{org} = 42.1\%$, C:N ratio = 246 247 77.7, and ${}^{13}C$ labelling = 7.01%), or were not amended (control). Wheat residues were used as 248 "model" C-input to assess the response of microbial activity to addition of a given quality of plant 249 material in all soils. Although they were not amended with wheat, control microcosms were mechanically disturbed as amended microcosms. The 528 microcosms (88 samples \times 2 treatments 250 $(^{13}C\text{-amended and control}) \times 3$ replicates) were incubated in the dark for 105 days under controlled 251 252 temperature (20°C) and moisture (60% WHC) conditions.

The volume of CO_2 produced, used as an index of soil respiration, was sampled after 3, 7, 14, 21, 28, 42, 63, and 80 days. Microcosms were aerated by flushing with air at each sampling date to avoid the perturbation of microbial activity mediated by increased CO_2 concentration. Therefore, the measured soil respiration corresponded to the CO_2 accumulated in the microcosm headspace between two sampling dates.

At each sampling date, the gaseous phase was sampled in 10 ml airtight flasks, to measure the CO_2 concentrations using an Agilent 7890B GC equipped with a thermal conductivity detector coupled to an automatic sampler (Agilent HS7697A Headspace), and in 12 ml airtight flasks, to measure the ¹³C-CO₂ enrichment on a Trace Gas analyzer (Micromass) linked to an isotopic-ratio mass spectrometer (Isoprime, Elementar). In this study, only the cumulative values of respired CO_2 at the end of the incubation are reported.

The multi-point calibration for CO₂ concentrations and ¹³CO₂ were both carried out using a GasMix diluter, AlyTech. For the isotopic ¹³CO₂ measurements, a variable ¹³C enrichment, ranging from natural abundance (1.08%) to 90% of ¹³C was used. The accuracy of the calibration, expressed as the difference in ¹³C percentage between the theoretical and the measured values was less than 0.3% (R²=0.99). The use of ¹³C-labeled wheat litter allowed to discriminate the contribution to total respiration (R_T expressed in µg C-CO₂ g⁻¹ soil) of the SOM- and litter-derived respiration (R_S and R_L , respectively) using mass-balance equations:

272 Eq. (1)
$$[R_T] = [R_S] + [R_L]$$
 and $[R_T] \times \delta_T = [R_S] \times \delta_S + [R_L] \times \delta_L$

273 Eq. (2)
$$[R_S] = \frac{[R_T] \times (\delta_T - \delta_L)}{\delta_S - \delta_L}$$

274 Where δ_i , i {T, S, L} is the ¹³C signature of the total, SOM- and litter-derived respired CO₂, 275 respectively. For the calculation, we made the assumption that the isotopic fractionation between the 276 solid fraction (soil or litter) and the CO₂ produced by its mineralization was negligible. Based on these 277 equations, the priming effect was calculated as following:

278 Eq. (3)
$$[R_{priming \ effect}] = [R_{T \ amended}] - [R_{T \ control}] - [R_L]$$

and expressed as mg of C-CO₂ primed per g of dry weight soil (PE-DW) and per TOC unit (PE/TOC).

For this study, values of PE are expressed as cumulative values of CO₂ emitted during the incubation.

281

282 2.6. Data analyses

The analyses were performed with R (R Core Team, http://www.r-project.org/) using "vegan", "car", 283 284 and "leaps" packages (Fox and Weisberg, 2011; Lumley, 2017; Oksanen et al., 2018). Correlation 285 heatmap was generated using "corrplot" package (Wei and Simko, 2017). A variance partitioning 286 approach was used to identify the most significant explanatory variables for the PE, following the 287 method proposed by Chemidlin Prévost-Bouré et al. (2014). Using the Spearman's correlation matrix, variables with a coefficient $\rho \ge 0.8$ were selected and only one of each pair has been included into the 288 models, to avoid collinearity. As a second step, a model selection was conducted by simultaneously 289 minimizing the Bayesian Information Criterion (BIC) and maximizing the adjusted R^2 using the 290 function regsubset ("leaps" package). On this subset of explanatory variables, a forward selection 291 procedure was applied to identify the model with the highest R^2 . The amounts of variance for PE 292 explained by each descriptor, intended as marginal and shared variance, were determined by canonical 293 variation partitioning, whereas the adjusted R^2 was determined by redundancy analysis. The variance 294

inflation factors (VIF) for each descriptor were calculated using the *vif* function ("car" package). The descriptors with a VIF \leq 3 were selected for the regression model, limiting the effect of collinearity on the estimated regression coefficients (Sheather, 2009). A total of 1000 permutations of the reduced model were computed to assess the significance of marginal effects.

Using the same procedure described above, two different models were built to further decorticate the collinearity of our data, since an increase of TOC is often correlated to an increase in microbial biomass, WEOC, and thus respiration rates. The first model (identified as "PE-DW") was run using PE and explanatory variables expressed on unit per gram of dry weight soil; the second model was run using PE and some explanatory variables (WEOC, microbial biomass, and respiration rates) normalized to TOC and identified as "PE/TOC".

305 To simplify the discussion of the manuscript, only the parameters selected as significant descriptors by 306 the two models were graphically represented and discussed within the text. Boxplots were generated using "ggplot2" (Wickham, 2009) packages in R. Data normality was checked before pairwise 307 308 comparisons among land-use classes with a Shapiro-Wilk's test. The significant differences for the 309 analyzed parameters between land-use classes were assessed by non-parametric Kruskal-Wallis tests 310 and the Dunn's multiple pairwise comparisons. This step allows to interpret how land-use affects PE descriptors and thus the susceptibility of agroecosystems to PE. Due to the non-normal distribution of 311 312 most of the values, two different levels of significance were used, p=0.05 was evidenced by lowercase 313 letters, while uppercase letters were used when significance satisfied the Bonferroni's correction for 10 hypotheses (significance level p=0.005). No letters were used in case of the lack of significance 314 315 between land-use classes.

316

317 **3. Results**

318 3.1. Soil physical and chemical parameters

The correlation heatmap in figure 2 highlights a strong and diffuse correlation network between theinvestigated variables.

The contents of TOC were significantly higher only for permanent grassland soils, whereas no significant differences were found between the other land-uses (Fig. 3A). TOC and TN were highly

323 correlated ($\rho^2 = 0.926$, p-value < 0.0001, Fig. 2) leading to the absence of significant differences 324 among land-uses for C/N ratios (10.1 to 10.2 for all treatments, Table S1). Hereafter, the variables TN 325 and TOC/TN will be omitted from the discussion, using TOC contents as the unique reference 326 parameter.

More WEOC was extracted from samples with higher frequency of grassland, the amount of extracted C normalized to TOC content showed significant differences between cropland and permanent grassland samples only, with larger amounts extracted from the latest (Fig. 3B).

The stability of soil aggregates increased with grassland frequency and the MWD was generally higher for permanent grassland, as well as for HI compared to cropland (Fig. 3D). Texture of analyzed soils evidenced how samples classified as permanent and high-frequency grassland were shifted through a slightly higher presence of clay, compensated by a relative decrease in silt content (Fig. 3E). Concomitantly, permanent grassland samples showed lower and more dispersed pH values compared to other treatments, even if no significant differences were detected (Fig. 3F). Olsen's P content was higher for cropland than for HI and permanent grassland (Fig. 3G).

337 The quantity of crop litter returned yearly to soil differed according to land-use. HI samples received338 significantly higher inputs of crop litter than MI, even if data dispersion was high (Fig. 3H).

339

340 3.2. Qualitative indicators of soil OM obtained from EEM fluorescence and PARAFAC 341 decomposition of water-extractable OM

The components identified from the EEM fluorescence and PARAFAC decomposition are shown in 342 figure 1. The CP1_(Fluo) most intense peak corresponds to fluorophores defined as "terrestrial humic-343 like" material and negatively correlated with DOM bioavailability (Lambert et al., 2017; Romero et 344 345 al., 2017), whereas the CP1_(Fluo) secondary peak is similar to fluorophores correlated to biological 346 activity in freshwater DOM (CP3 and CP5 in Lambert et al., 2017). The spectra obtained for CP2_(Fluo) 347 resemble those of CP1_(Fluo) with a shift towards longer wavelengths, previously described as "terrestrial humic-like" compounds (Stedmon and Markager, 2005) with greater aromaticity, higher 348 molecular weight, and a larger degree of conjugation. Thus, CP2_(Fluo) is interpreted as a pool of organic 349 350 compounds that have undergone more processing and transformation within the soil than CP1_(Fluo).

These characteristics and the strong correlations among $CP1_{(Fluo)}$ and $CP2_{(Fluo)}$ ($r^2 = 0.953$; n = 253) suggest their common sources, probably related to lignin breakdown during plant litter decomposition (Cory and McKnight, 2005).

354 The emission spectra of CP3_(Fluo) presented a secondary peak with a "terrestrial humic-like" profile similar to the main peaks of the other two components. The primary peak is, however, found at longer 355 wavelengths for excitation, similar to peaks found for biologically processed organic substrate (Coble, 356 357 1996; Søndergaard et al., 2003). Such spectrum is less common in the literature and suggests an even higher degree of aromaticity with extensive and conjugated chemical complexes and low $\delta^{13}C$ 358 359 signatures, indicating a scarce contribution of molecules from microbial origin (Dainard et al., 2015; 360 Osburn et al., 2012; Yamashita et al., 2011). Bernard-Jannin et al. (2018) showed that such component is present in soil pore water of a peatland and tends to concentrate during dry period and Lambert et al. 361 362 (2017) demonstrated its contribution to total fluorescence in rivers during high-water period. Therefore, these fluorescent compounds assigned to CP3_(Fluo) likely correspond to highly processed 363 organic material that tends to be preserved in soils, such as biodegraded tannins (Maie et al. 2007), or 364 365 degradation products inherited from plant litter and swine manure (Hunt and Ohno, 2007; Wheeler et al., 2017). Fluorescence signal of CP3 (Fluo) decreased with higher frequencies of grassland (p<0.05, 366 367 Fig. 3C), whereas the other fluorophores showed no significant differences among treatment.

368

369 3.3. Carbon fluxes, measured PE and soil microbial communities

Soil respiration of control microcosms, considered as the potential basal respiration of soil and 370 expressed as mg of CO₂ emitted per gram of TOC, increased with grassland frequency, leading to 371 372 significantly higher values for permanent grassland when compared with cropland (Fig. 4A). On the 373 contrary, for the respiration of amended microcosms, permanent grassland shows significantly lower CO₂ emissions compared to HI, LO, and cropland (Fig. 4B). Calculated values of PE-DW were 374 375 significantly higher (p < 0.05) for HI than for MI and cropland, and permanent grassland presented huge data dispersion (Fig. 4C). The amount of wheat litter mineralized ranged between the 29.5 and 376 the 42.9% of the total litter-¹³C added (Table 1), no significant differences between treatments were 377 378 highlighted. PE was responsible of the mineralization of a portion of TOC ranging between 1.7 and 379 3.2 mg g⁻¹ TOC (Fig. 4D) during the incubation experiment; as a comparison term, basal respiration of 380 soils mineralized between 6.3 and 10.4 mg g⁻¹ of TOC (Fig. 4B). At a p<0.05 significance level, soils 381 under LO and HI showed the highest values of PE, permanent grassland the lowest ones.

The influence of legume plants in some temporary and permanent grassland was tested by splitting grassland-only samples into two groups (with and without legumes). Soils from farms with legume crops showed slightly higher PE potential (2.40 ± 0.70 vs. 2.60 ± 0.95 mg g⁻¹ TOC) than those without legumes, but with high data dispersion (p=0.363). Running the same test excluding the samples from permanent grassland reduced the data dispersion, but differences between the two groups were still not significant (p=0.187).

The quantities of DNA extracted per g of TOC showed and increasing trend with frequency of grassland, but only HI had significantly higher values (p<0.05) when compared with Cropland (Fig. 4E). LO and MI showed significantly higher values (p<0.05) of bacterial Pielou's indexes of evenness (Eve_B) than permanent grassland (Fig. 4F).

392 3.4. Variance partitioning models rank variables related to the priming effect potentials

393 The complete statistical description of analyzed parameters is presented in table 1. Priming effect per 394 unit of dry weight soil (PE-DW) and priming effect per TOC unit (PE/TOC) were selected as dependent variables to build two separate models, and the variance explained by each one of the 395 396 independent variables was calculated with the variance partitioning models (Fig. 5). The PE-DW and 397 PE/TOC models explained 53.62 and 49.04% of the total variance, respectively, but a part of it 398 consisted of interactions, i.e. co-variance that cannot be assigned unequivocally to one descriptor. This 399 is a common issue when several independent variables present a co-linearity pattern. For the PE-DW model, the variance assigned to interactions was negative (-6.33%). This value is due to the use of 400 adjusted R^2 values as fitting parameters by the function ordiR2step, combined with a low sample to 401 402 variable ratio. The negative interactions would lead to a lower percentage of total variance explained, 403 but the value is small enough not to invalidate our analysis, and it may be approximated to zero 404 (Tabachnick and Fidell, 2007). As for positive value of interactions, this percentage of variance cannot 405 be subtracted to the descriptors selected. Among the selected descriptors for this model, frequency of 406 grassland was selected as a numeric variable, describing 5.24% (p<0.001) of the variance and with a 407 positive relationship with measured PE.

408 The significant variables explaining the PE-DW variations corresponded to water-soluble organic 409 matter chemical composition ($CP3_{(Fluo)}$), P availability, microbial biomass, soil pH and land-use 410 characteristics (grasslands frequency and crop residues). The WEOC amount and clay content were 411 also significant but explained much smaller part of variance. All the variables had a positive effect on 412 the PE-DW with the exception of soil microbial biomass and crop residues.

In the case of PE/TOC model, despite the high contribution of interactions (16.52%), a large part of PE variance (53.62%) was explained by a set of only four significant variables with the following hierarchy: MWD, grassland frequency (selected as land-use categories, not numeric), Biomass/TOC and bacterial evenness. Among the different explanatory variables, soil microbial biomass promoted PE/TOC, whereas higher aggregation and bacterial evenness impeded it. The effect of the frequency of grassland on PE/TOC was land-use dependent, indeed PE/TOC was higher at low and high frequency of grasslands in comparison to cropland and lower for samples under mid and permanent grassland.

420

421 **4.** Discussion

422 4.1. Priming effect in a mosaic of land-uses

The average CO₂ production attributed to PE for cropped soils ranged between 2 and 3 mg g^{-1} TOC, a 423 value equal to half of the basal respiration of soils for some treatments. Soil is the source for 60-90% 424 425 of the total CO_2 fluxes of an ecosystem (Longdoz et al., 2000), and Guenet et al. (2018) showed how 426 prediction on C sequestration may be halved if PE is integrated into global C models. Yet, the 427 mechanisms impacting the C balance of PE are still under debate, since they may co-occur at the same 428 time, vary from site to site, and be land-use dependent (Chen et al., 2014; Kuzyakov et al., 2000; 429 Razanamalala et al., 2018). In fact, the frequency of grassland in the rotation was selected as a 430 significant descriptor of PE by both variance partitioning models, implying that susceptibility of agroecosystems to PE is partially related to the adopted land-use. 431

Introduction of grassland into crop rotations results in changes in type and quantity of litter returned tothe soil, and potentially supports the establishment of more diverse roots-soil interactions modifying

soil physical properties and promote the stabilization of roots-derived C (Cambardella and Elliot,
1992; Panettieri et al., 2017). Most of the soils from permanent grassland received no manuring,
aboveground litter was mowed and we could not estimate the quantitative information about
rhizodeposition.

The PE-DW model selected the frequency of grassland as a positive descriptor. In the same time, we found positive correlation of TOC and WEOC amounts, as well as clay content with the frequency of grassland. Considering these results together, we hypothesized that the higher TOC contents accumulated under permanent grassland support larger C mineralization fluxes and strongly influence the PE differences between our land-use classes, as highlighted by the model. This observation justifies our approach consisting of running a second model after normalization of PE fluxes to TOC contents in the soil samples.

For PE/TOC model, the increase of PE was land-use dependent: a positive relationship was found for LO and HI, whereas the effect of MI and permanent grassland were negative. These results add key information to the results for PE-DW model, suggesting that, in our system, PE is triggered by specific frequencies in the occurrence of grassland within crop rotation cycles.

These observations suggest that alternation of cropland and grassland can create favorable conditions for PE. We observed that short-term conversion of agricultural fields into grassland (LO), or shortterm cultivation of prairies (HI), both resulted in increased values of PE/TOC. The similarity between these two situations is that the ecosystem functioning under the dominant land-use is destabilized by the land-use conversion during one to three years per ten years. The mechanisms triggering the PE, however, are likely to differ between the two situations.

If for LO we can hypothesize that frequent tillage and a C-limited environment may change the composition of microbial communities and create biogeochemical conditions prone to PE. The conversion to grassland is likely resulting in changes of SOM composition with more belowground biomass inputs (Jackson et al., 1996) compared to cropland, as well as preservation of organic compounds entering soils through root exudation or root litter (Panettieri et al., 2017). We think that these pools of OM may present a different decomposition stage, and be more easily mineralized by microorganisms than SOM under cropland causing the increased PE under LO land-use (Shahbaz et 462 al., 2018). The low C contents and high N and P availability are both favorable conditions for PE in463 these soils (Kuzyakov, 2010).

For HI samples, soils are principally devoted to grassland, with higher SOM levels and root inputs. The occasional ploughing of these soils dominated by grassland use is altering soil structure, as shown by lower MWD of aggregates and could promote PE compared to permanent grassland. Indeed, the microbial decomposers in these soils are more likely adapted to the decomposition of SOM made available by the loss of physical protection after ploughing (Kuzyakov, 2010). However, a further characterization of the agricultural practices is necessary to understand this result.

470 Interestingly, we did not find any increase in PE/TOC for MI land-use class compared to soil under 471 permanent cropland or grassland land-uses. While the processes underlying the different responses of 472 soils under various frequencies of temporary grassland use are hard to demonstrate, MI samples 473 supports the idea that the frequency of switches between land-uses alter soil susceptibility to PE in a 474 non-linear way. Increase or decrease with respect to MI frequency resulted in higher PE values. It is 475 noteworthy that MI soils showed more evenly distributed microbial communities. This result has been 476 related to a greater adaptation and resilience potential of the soil subjected to "intermediate perturbation" (Acosta-Martínez et al., 2008; Shange et al., 2012; Tardy et al., 2015). 477

478

479 4.2. Influence of land-use on the descriptors of PE-DW model

The amount of crop litter returned to soil was selected by PE-DW model, showing that a soil with lower input of labile C represents an energy-limited environment prone to PE when labile C is occasionally added (Hamer and Marschner, 2005; Razanamalala et al., 2018). Among the microbiological variables measured, only the quantity of microbial biomass was selected as a predictor of PE-DW, with a negative relation. Higher content of biomass is normally found in soils of "high quality" and microbial interactions are more devoted to mineralization of fresh substrates rather than degradation of physically or chemically protected SOM (Blagodatskaya and Kuzyakov, 2008).

Soil pH was selected as a positive descriptor of PE-DW, indicating that among the studied soils, those
with slightly higher pH were more prone to PE. In general, soils with slightly higher pH have higher
biomass, higher respiration rates, and faster degradation of SOM. In a single soil type with a similar

490 pH gradient of our study, Aciego Pietri and Brookes (2009) found that pH and substrate availability affected the structure of microbial community during the decomposition of wheat straw via the 491 492 succession of litter degradation activities assigned to specific microbial groups. For their study, the 493 increase in soil respiration after straw addition was positively correlated with pH. Taking into account our results, we can therefore assume that a part of the respiration measured by Aciego Pietri and 494 495 Brookes (2009) at higher pH was originated by priming effect. No differences in mean pH were found 496 between land-use classes (Fig. 3F), suggesting that the pH effect on PE is independent from our classification. 497

Further interesting results are the positive relations of WEOC, CP3(fluo) and clay with PE-DW. For this 498 499 study, WEOC per TOC unit increased with frequency of grassland, and is a common early indicator of 500 changes in SOM chemical and biochemical composition (Bongiorno et al., 2019; Panettieri et al., 501 2014) that increases with the amount of microbial biomass and root exudation (Kalbitz et al., 2000). 502 On one hand, microbial biomass is larger for HI compared to cropland (p < 0.05, Fig. 4C), and a trend 503 towards higher values of WEOC at higher frequencies of grassland was detected. On the other hand, 504 grassland has higher root density than the cereal crops of the study area, especially for surface soils 505 (Jackson et al., 1996), resulting in larger inputs by rhizodeposition and higher microbial activity (van 506 Eekeren et al., 2008).

507 The component of WEOC fluorescence CP3_(fluo) is the variable explaining the largest part of variance 508 in the PE/DW model. It corresponds to a highly processed form of SOM with a high degree of 509 aromaticity, therefore not an ideal source of C for microbes (Coble, 1996; Søndergaard et al., 2003). 510 The relative contribution of $CP3_{(fluo)}$ decreased with the increase of grassland frequency, indicating 511 differences in WEOC composition between land-uses. This result may be explained by larger inputs of 512 fresh and labile organic matter, larger microbial biomass content, and different composition of 513 microbial communities under grassland, while cropland soils generally receive larger quantities of 514 exogenous OM, especially swine manure in the study area. Hunt and Ohno (2007) identified CP3(Fluo) 515 as being conserved during biodegradation of swine manure. It should be taken into account that the measured CP3_(fluo) represents a group of fluorescent compounds referenced as resistant to microbial 516 517 degradation and co-varying with PE-DW. Each fluorophore identified in WEOC and dissolved organic 518 matter is likely accompanied with a consortium of non-fluorescent soluble organic compounds 519 (Stubbins et al., 2014). Deciphering the high molecular complexity of WEOC, e.g. using high-520 resolution mass spectrometry (Guigue et al., 2016; Roth et al., 2014), could allow to investigate 521 organic molecules (e.g. easily degradable ones, N-rich molecules, or organic acids) directly or indirectly related to PE fluxes. The positive relation of CP3(fluo) with PE-DW may be due to: (i) the 522 dynamics of co-metabolism of the compounds forming the consortium with CP3(fluo) in presence of 523 524 fresh litter (Chen et al., 2014), or (ii) to the temporary stabilization of the consortium with CP3_(fluo) via 525 adsorption on clay surfaces and short-range order minerals investigated in recent works (Finley et al., 526 2018; Keiluweit et al., 2015), most probably a combination of both. This adsorption of highly aromatic 527 compounds can be reversed by input of organic acids from root exudates, manure (Hamer and Marschner, 2005), or produced during the degradation of new litter inputs (Blagodatskaya and 528 529 Kuzyakov, 2008).

The texture of some of the permanent grassland and, to a minor extent, HI samples shifted through a more clayey texture (Fig. 3E). Since a lower water drainage is often associated with higher clay contents, rotations with higher frequency of grassland may have been established in higher wetness areas, to avoid crop losses due to flooding. The positive effect of clay content on PE could be attributed to an increase of carbon content in these soils.

535 Olsen P explained the second highest percentage of variance of PE-DW model. High frequencies of 536 grassland were characterized by lower amounts of organic amendment per year and lower available P 537 contents. In the study area, differences in soil available P contents are attributable mostly to the 538 spreading of swine manure derived from the widely developed husbandry farms in the area of study (Matos-Moreira et al. 2017). However, the absence of correlation between Olsen P and PE-DW ($\rho^2 =$ 539 540 0.035, p-value 0.7) highlight that the variance explained may be due to intragroup variability and/or by 541 different P speciation, not investigated in this study. The fact that P is mainly derived from swine manure and that CP3, the descriptor explaining the higher percentage of variance, was also found in a 542 swine manure degradation experiment (Hunt and Ohno, 2007) points towards the importance of 543 manure degradation in PE dynamics, even if further experiments are needed to corroborate these 544 findings. 545

546

547 4.3. Influence of land-use on the descriptors of PE/TOC model

548 The selected descriptors for PE/TOC model were MWD, microbial C normalized to TOC and bacterial 549 evenness. MWD, a proxy of aggregation, represents a physical protection of SOM from degradation. Thus, the negative relation with PE is easily understandable by the limited access to the protected 550 551 SOM (Six et al. 1999). The MWD of the water-stable aggregates increased with frequency of 552 grassland, thus lower PE is expected at higher frequencies of grassland. Consistent with literature, 553 temporary and permanent grassland are reported to provide a better protection of aggregates due to 554 two main factors: (i) a lower number of tillage operations which physically disrupt soil aggregates (Six 555 et al. 1999; Panettieri et al. 2015) and (ii) a higher contribution of aggregation factors, such as root 556 exudates and extracellular polymeric substances (Redmile-Gordon et al. 2014) proceeding from a 557 more extended and dense root network (Tisdall and Oades 1982; Jackson et al. 1996; Six et al. 1998; Panettieri et al. 2017). However, the higher PE/TOC measured for HI (Fig. 4D) contrasts this 558 559 interpretation, suggesting that PE regulation is dependent of other co-occurring factors.

560 The other two descriptors are related with microbial communities. A higher value of microbial biomass per TOC unit represents the paradigm of a higher microbial growth with higher respiration, 561 but also a more energy limited environment, in which more microorganisms compete for the SOM 562 563 substrate. The PE/TOC model selected this descriptor as a trigger for PE, and the bacterial evenness as 564 a descriptor with a negative relationship with PE. In a substrate limited environment, competition and 565 predation may increase, inducing a shift of microbial communities through a less even distribution of species (Banerjee et al., 2016; Fontaine et al., 2003). This reduce the resilience of the communities and 566 567 the capacity of degradation of SOM, limiting the colonization of specific niches and, consequently, 568 making the soil more prone to PE when fresh energy sources are added during incubation (Chen et al., 569 2014; Razanamalala et al., 2018. Bacterial evenness had a positive correlation with two descriptors of 570 SOM quality (CP3_(fluo) and TOC/TN ratio). SOM with a more complex and diverse chemical structure 571 promote the development of a higher bacterial evenness and increase the capacity of microbial communities to degrade substrates (Goldfarb et al., 2011), resulting in a more efficient cycling of the 572 whole pool of allochthonous fresh-C molecules, rather than triggering PE from autochthonous pools of 573

SOM. The trend to higher microbial biomass per TOC unit at higher frequency of grassland (Fig. 4C)
is consistent with literature for soils under grassland (Baumann et al., 2013; Potthoff et al., 2005),
since a higher microbial activity is found in rhizosphere and grasslands have a denser and
interconnected root architectures, especially for upper soil layers (Jackson et al., 1996; Panettieri et al.,
2017; Rasse et al., 2005; Sparling, 1992).

579 Regarding the composition of soil microbial communities, Le Guillou et al. (2018) conducted a study 580 on the same area using the theory of intermediate perturbation, or hump-backed curve (Acosta-Martínez et al., 2008; Shange et al., 2012; Tardy et al., 2015) to relate their metagenomics results with 581 582 land-use classes. Maximum bacterial diversity occurs at an optimum of soil perturbation, in which 583 more diverse species can proliferate. Considering the classification adopted for the present study, 584 higher values for bacterial richness (Table S1) and evenness (Fig 4E), found for LO and MI 585 confirming that slight modifications in crop rotation led to slightly but significant modification of 586 bacterial richness and evenness. This explanation is in line with the fact that MI soils, corresponding to 587 the ecosystems with the most equal distribution between time under cropland and under grassland, 588 tend to have a higher evenness J' index and a lower potential for PE than LO and HI. It suggests that 589 this frequency of disturbances (tillage and changes in land-use) stimulate the adaptation of microbial 590 communities that become more resilient and are thus able to sustain ecosystem functions better than in 591 agroecosystems where lower frequencies of disturbances had a more negative impact regarding SOM 592 decomposition and PE, i.e. in LO and HI.

593

594 Conclusions

595 Unveiling the dynamics of priming effect (PE) at landscape scale and on soil samples from real farms 596 is an extremely complex operation, especially for such study area (5 km²) with little heterogeneities in 597 climate, soil types and agricultural management. Chemical and physical interactions with soil mineral 598 phases are fundamental parts of the environmental context in which living organisms are developing. 599 PE represents an indicator of SOM turnover and nutrient cycling, but particular attention must be paid 500 to high values of PE per TOC unit (PE/TOC), a clear indication that C losses may occur. Targeted 501 land-uses may help to mitigate PE/TOC or at least reduce the susceptibility of soils to PE/TOC. For 602 the studied area, SOM contents with lower PE potentials are attained for temporary grassland at an 603 optimum of 40-60% of time devoted to grassland, corresponding to the intermediate frequency in 604 grassland use (MI), while a change in the frequency of grassland seems to create conditions favorable to over-mineralization of soil C through PE. The MI frequency also represents a microbiological 605 optimum with higher values of bacterial richness and evenness. Higher frequencies of grassland (HI) 606 607 may tend to increase SOM contents, whereas lower frequency may be uninfluential for this purpose, 608 when compared with permanent cropland. However, HI samples were also intensively prone to PE, 609 and the newly stored SOM may be unstable in the long-term. Interestingly, a pool of specific water soluble organic compounds identified by fluorescence analyses was selected among the descriptors for 610 the susceptibility of soils to PE, highlighting that the composition of soil organic matter in both solid 611 and dissolved phases are potential indicators of PE dynamics, thus offering new possibilities for future 612 research aiming to refine prediction models of C cycle. 613

614

615 Acknowledgments

- 616 This study has been funded by the French National Research Agency (ANR) through the project
- 617 MOSAIC (ANR-12-AGRO-005). The authors acknowledge the support of CNRS and INRA for the
- 618 Observatory ORE-AgrHys, and the support of AllEnvi for the Observatory SOERE-RBV.

619

620 References

- Aciego Pietri, J.C., Brookes, P.C., 2009. Substrate inputs and pH as factors controlling microbial
 biomass, activity and community structure in an arable soil. Soil Biol. Biochem. 41, 1396–1405.
 https://doi.org/10.1016/J.SOILBIO.2009.03.017
- Acosta-Martínez, V., Dowd, S., Sun, Y., Allen, V., 2008. Tag-encoded pyrosequencing analysis of
 bacterial diversity in a single soil type as affected by management and land use. Soil Biol.
 Biochem. 40, 2762–2770. https://doi.org/10.1016/J.SOILBIO.2008.07.022
- AFNOR, 2012. ISO 10930. Soil quality Measurement of the stability of soil aggregates subjected to
 the action of water -.
- AFNOR, 2003. NF X31-107. Soil Quality Particle Size Determination by Sedimentation Pipette
 Method.
- AFNOR, 1998. NF ISO 13878. Soil Quality Determination of Total Nitrogen Content by Dry
 Combustion ("Elemental Analysis").

- AFNOR, 1995. NF ISO 10694. Soil Quality Determination of Organic and Total Carbon after Dry
 Combustion (Elementary Analysis).
- Banerjee, S., Kirkby, C.A., Schmutter, D., Bissett, A., Kirkegaard, J.A., Richardson, A.E., 2016.
 Network analysis reveals functional redundancy and keystone taxa amongst bacterial and fungal
 communities during organic matter decomposition in an arable soil. Soil Biol. Biochem. 97, 188–
 198. https://doi.org/10.1016/j.soilbio.2016.03.017
- 639 Bastida, F., García, C., Fierer, N., Eldridge, D.J., Bowker, M.A., Abades, S., Alfaro, F.D., Asefaw 640 Berhe, A., Cutler, N.A., Gallardo, A., García-Velázquez, L., Hart, S.C., Haves, P.E., Hernández, 641 T., Hseu, Z.Y., Jehmlich, N., Kirchmair, M., Lambers, H., Neuhauser, S., Peña-Ramírez, V.M., Pérez, C.A., Reed, S.C., Santos, F., Siebe, C., Sullivan, B.W., Trivedi, P., Vera, A., Williams, 642 M.A., Luis Moreno, J., Delgado-Baquerizo, M., 2019. Global ecological predictors of the soil 643 644 priming effect. Nat. Commun. 10. https://doi.org/10.1038/s41467-019-11472-7Baumann, K., 645 Sanaullah, M., Chabbi, A., Dignac, M.F., Bardoux, G., Steffens, M., Kögel-Knabner, I., Rumpel, C., 2013. Changes in litter chemistry and soil lignin signature during decomposition and 646 647 stabilisation of 13C labelled wheat roots in three subsoil horizons. Soil Biol. Biochem. 67, 55-648 61. https://doi.org/10.1016/j.soilbio.2013.07.012
- Bernard-Jannin, L., Binet, S., Gogo, S., Leroy, F., Défarge, C., Jozja, N., Zocatelli, R., Perdereau, L.,
 Laggoun-Défarge, F., 2018. Hydrological control of dissolved organic carbon dynamics in a
 rehabilitated Sphagnum-dominated peatland: a water-table based modelling approach. Hydrol.
 Earth Syst. Sci. 22, 4907–4920. https://doi.org/10.5194/hess-22-4907-2018
- Beven, K.J., Kirby, M.J., 1979. A physically based, variable contributing area model of basin
 hydrology / Un modèle à base physique de zone d'appel variable de l'hydrologie du bassin
 versant. Hydrol. Sci. Bull. 24, 43–69. https://doi.org/10.1080/026266667909491834
- Bingeman, C.W., Varner, J.E., Martin, W.P., 1953. The Effect of the Addition of Organic Materials on
 the Decomposition of an Organic Soil1. Soil Sci. Soc. Am. J. 17, 34.
 https://doi.org/10.2136/sssaj1953.03615995001700010008x
- Blagodatskaya, E., Kuzyakov, Y., 2008. Mechanisms of real and apparent priming effects and their
 dependence on soil microbial biomass and community structure: Critical review. Biol. Fertil.
 Soils 45, 115–131.
- Bongiorno, G., Bünemann, E.K., Oguejiofor, C.U., Meier, J., Gort, G., Comans, R., Mäder, P.,
 Brussaard, L., de Goede, R., 2019. Sensitivity of labile carbon fractions to tillage and organic
 matter management and their potential as comprehensive soil quality indicators across
 pedoclimatic conditions in Europe. Ecol. Indic. 99, 38–50.
 https://doi.org/10.1016/J.ECOLIND.2018.12.008
- Cambardella, C.A., Elliot, E.T., 1992. Particulate soil organic-matter changes across a grassland
 cultivation sequence. Soil Sci. Soc. Am. J. 56, 777–783.
- Chemidlin Prévost-Bouré, N., Dequiedt, S., Thioulouse, J., Lelièvre, M., Saby, N.P.A., Jolivet, C.,
 Arrouays, D., Plassart, P., Lemanceau, P., Ranjard, L., 2014. Similar Processes but Different
 Environmental Filters for Soil Bacterial and Fungal Community Composition Turnover on a
 Broad Spatial Scale. PLoS One 9, e111667. https://doi.org/10.1371/journal.pone.0111667
- 673 Chen, R., Senbayram, M., Blagodatsky, S., Myachina, O., Dittert, K., Lin, X., Blagodatskaya, E.,
 674 Kuzyakov, Y., 2014. Soil C and N availability determine the priming effect: microbial N mining
 675 and stoichiometric decomposition theories. Glob. Chang. Biol. 20, 2356–2367.
 676 https://doi.org/10.1111/gcb.12475
- Chin, Y.-P., Aiken, G., O'Loughlin, E., 1994. Molecular Weight, Polydispersity, and Spectroscopic
 Properties of Aquatic Humic Substances. Environ. Sci. Technol. 28, 1853–1858.
 https://doi.org/10.1021/es00060a015

- Coble, P.G., 1996. Characterization of marine and terrestrial DOM in seawater using excitationemission matrix spectroscopy. Mar. Chem. 51, 325–346. https://doi.org/10.1016/03044203(95)00062-3
- Cory, R.M., McKnight, D., 2005. Fluorescence spectroscopy reveals ubiquitous presence of oxidized
 and reduced quinones in dissolved organic matter. Environ. Sci. Technol. 39, 8142–8149.
- Craig, M.E., Pearson, S.M., Fraterrigo, J.M., 2015. Grass invasion effects on forest soil carbon depend
 on landscape-level land use patterns. Ecology 96, 2265–2279. https://doi.org/10.1890/14-1770.1
- Dainard, P.G., Guéguen, C., McDonald, N., Williams, W.J., 2015. Photobleaching of fluorescent
 dissolved organic matter in Beaufort Sea and North Atlantic Subtropical Gyre. Mar. Chem. 177,
 630–637.
- de Graaff, M.-A., Jastrow, J.D., Gillette, S., Johns, A., Wullschleger, S.D., 2014. Differential priming
 of soil carbon driven by soil depth and root impacts on carbon availability. Soil Biol. Biochem.
 692 69, 147–156. https://doi.org/10.1016/J.SOILBIO.2013.10.047
- bequiedt, S., Saby, N.P.A., Lelievre, M., Jolivet, C., Thioulouse, J., Toutain, B., Arrouays, D., Bispo,
 A., Lemanceau, P., Ranjard, L., 2011. Biogeographical patterns of soil molecular microbial
 biomass as influenced by soil characteristics and management. Glob. Ecol. Biogeogr. 20, 641–
 652. https://doi.org/10.1111/j.1466-8238.2010.00628.x
- 697 Dignac, M.-F., Derrien, D., Barré, P., Barot, S., Cécillon, L., Chenu, C., Chevallier, T., Freschet, G.T.,
 698 Garnier, P., Guenet, B., Hedde, M., Klumpp, K., Lashermes, G., Maron, P.-A., Nunan, N.,
 699 Roumet, C., Basile-Doelsch, I., 2017. Increasing soil carbon storage: mechanisms, effects of
 700 agricultural practices and proxies. A review. Agron. Sustain. Dev. 37, 14.
 701 https://doi.org/10.1007/s13593-017-0421-2
- Finley, B.K., Dijkstra, P., Rasmussen, C., Schwartz, E., Mau, R.L., Liu, X.-J.A., van Gestel, N.,
 Hungate, B.A., 2018. Soil mineral assemblage and substrate quality effects on microbial priming.
 Geoderma 322, 38–47. https://doi.org/10.1016/J.GEODERMA.2018.01.039
- Fontaine, S., Mariotti, A., Abbadie, L., 2003. The priming effect of organic matter: a question of
 microbial competition? Soil Biol. Biochem. 35, 837–843. https://doi.org/10.1016/S00380717(03)00123-8
- Fox, J., Weisberg, S., 2011. An {R} Companion to Applied Regression, Second. ed. Sage, Thousand
 Oaks {CA}.
- Frank, A.B., Liebig, M.A., Hanson, J.D., 2002. Soil carbon dioxide fluxes in northern semiarid
 grasslands. Soil Biol. Biochem. 34, 1235–1241.
- Glover, J.D., Culman, S.W., DuPont, S.T., Broussard, W., Young, L., Mangan, M.E., Mai, J.G.,
 Crews, T.E., DeHaan, L.R., Buckley, D.H., Ferris, H., Turner, R.E., Reynolds, H.L., Wyse, D.L.,
 2010. Harvested perennial grasslands provide ecological benchmarks for agricultural
 sustainability. Agric. Ecosyst. Environ. 137, 3–12. https://doi.org/10.1016/j.agee.2009.11.001
- Goldfarb, K.C., Karaoz, U., Hanson, C.A., Santee, C.A., Bradford, M.A., Treseder, K.K., Wallenstein,
 M.D., Brodie, E.L., 2011. Differential Growth Responses of Soil Bacterial Taxa to Carbon
 Substrates of Varying Chemical Recalcitrance. Front. Microbiol. 2.
 https://doi.org/10.3389/fmicb.2011.00094
- Guenet, B., Camino-Serrano, M., Ciais, P., Tifafi, M., Maignan, F., Soong, J.L., Janssens, I.A., 2018.
 Impact of priming on global soil carbon stocks. Glob. Chang. Biol.
 https://doi.org/10.1111/gcb.14069
- Guenet, B., Leloup, J., Raynaud, X., Bardoux, G., Abbadie, L., 2010. Negative priming effect on
 mineralization in a soil free of vegetation for 80 years. Eur. J. Soil Sci. 61, 384–391.
 https://doi.org/10.1111/j.1365-2389.2010.01234.x

- Guigue, J., Harir, M., Mathieu, O., Lucio, M., Ranjard, L., Lévêque, J., Schmitt-Kopplin, P., 2016.
 Ultrahigh-resolution FT-ICR mass spectrometry for molecular characterisation of pressurised hot water-extractable organic matter in soils. Biogeochemistry 128, 307–326.
 https://doi.org/10.1007/s10533-016-0209-5
- Guigue, J., Mathieu, O., Lévêque, J., Mounier, S., Laffont, R., Maron, P.A., Navarro, N., Chateau, C.,
 Amiotte-Suchet, P., Lucas, Y., 2014. A comparison of extraction procedures for waterextractable organic matter in soils. Eur. J. Soil Sci. 65, 520–530.
 https://doi.org/10.1111/ejss.12156
- Haines-Young, R., Potschin, M., 2010. The links between biodiversity, ecosystem services and human
 well-being. Ecosyst. Ecol. a new Synth. 1, 110–139.
- Hamer, U., Marschner, B., 2005. Priming effects in different soil types induced by fructose, alanine,
 oxalic acid and catechol additions. Soil Biol. Biochem. 37, 445–454.
 https://doi.org/10.1016/J.SOILBIO.2004.07.037
- Helgason, B.L., Konschuh, H.J., Bedard-Haughn, A., VandenBygaart, A.J., 2014. Microbial
 distribution in an eroded landscape: Buried A horizons support abundant and unique
 communities. Agric. Ecosyst. Environ. 196, 94–102. https://doi.org/10.1016/j.agee.2014.06.029
- Hunt, J.F., Ohno, T., 2007. Characterization of fresh and decomposed dissolved organic matter using
 excitation-emission matrix fluorescence spectroscopy and multiway analysis. J. Agric. Food
 Chem. 55, 2121–2128. https://doi.org/10.1021/jf063336m
- 745 IUSS Working Group WRB, 2015. World Reference Base for Soil Resources 2014, update 2015
 746 International soil classification system for naming soils and creating legends for soil maps.,
 747 World Soil. ed. FAO, Rome.
- Jackson, R.B., Canadell, J., Ehleringer, J.R., Mooney, H.A., Sala, O.E., Schulze, E.D., 1996. A global
 analysis of root distributions for terrestrial biomes. Oecologia 108, 389–411.
 https://doi.org/10.1007/bf00333714
- Kalbitz, K., Solinger, S., Park, J.-H., Michalzik, B., Matzner, E., 2000. Controls on the dynamics of
 dissolved organic matter in soils: a review. Soil Sci. 165, 277–304.
- Keiluweit, M., Bougoure, J.J., Nico, P.S., Pett-Ridge, J., Weber, P.K., Kleber, M., 2015. Mineral
 protection of soil carbon counteracted by root exudates. Nat. Clim. Chang. 5, 588–595.
 https://doi.org/10.1038/nclimate2580
- Kirschbaum, M.U.F., 1995. The temperature dependence of soil organic matter decomposition, and
 the effect of global warming on soil organic C storage. Soil Biol. Biochem. 27, 753–760.
 https://doi.org/10.1016/0038-0717(94)00242-S
- Kuzyakov, Y., 2010. Priming effects: Interactions between living and dead organic matter. Soil Biol.
 Biochem. 42, 1363–1371. https://doi.org/10.1016/J.SOILBIO.2010.04.003
- Kuzyakov, Y., Blagodatskaya, E., 2015. Microbial hotspots and hot moments in soil: Concept & review. Soil Biol. Biochem. 83, 184–199. https://doi.org/10.1016/J.SOILBIO.2015.01.025
- Kuzyakov, Y., Friedel, J.K., Stahr, K., 2000. Review of mechanisms and quantification of priming
 effects. Soil Biol. Biochem. 32, 1485–1498.
- Lal, R., Stewart, B.A., 1990. Advances in Soil Science : Soil Degradation. Springer New York.
- Lambert, T., Bouillon, S., Darchambeau, F., Morana, C., Roland, F.A.E., Descy, J.-P., Borges, A. V.,
 2017. Effects of human land use on the terrestrial and aquatic sources of fluvial organic matter in
 a temperate river basin (The Meuse River, Belgium). Biogeochemistry 136, 191–211.
 https://doi.org/10.1007/s10533-017-0387-9
- TTO Le Bissonnais, Y., 1996. Aggregate stability and assessment of soil crustability and erodibility: I.

- Theory and methodology Stabilité structurale et évaluation de la sensibilité des sols à la
 battance et à l'érosion: I: Théorie et méthologie. Eur. J. Soil Sci. 47, 425–437.
 https://doi.org/10.1111/j.1365-2389.1996.tb01843.x
- Le Guillou, C., Chemidlin Prévost-Bouré, N., Karimi, B., Akkal-Corfini, N., Dequiedt, S., Nowak, V.,
 Terrat, S., Menasseri-Aubry, S., Viaud, V., Maron, P.-A., Ranjard, L., 2018. Tillage intensity and
 pasture in rotation effectively shape soil microbial communities at a landscape scale.
 Microbiologyopen e00676. https://doi.org/10.1002/mbo3.676
- Lemaire, G., Gastal, F., Franzluebbers, A., Chabbi, A., 2015. Grassland–Cropping Rotations: An
 Avenue for Agricultural Diversification to Reconcile High Production with Environmental
 Quality. Environ. Manage. https://doi.org/10.1007/s00267-015-0561-6
- Longdoz, B., Yernaux, M., Aubinet, M., 2000. Soil CO₂ efflux measurements in a mixed forest:
 impact of chamber disturbances, spatial variability and seasonal evolution. Glob. Chang. Biol. 6,
 907–917. https://doi.org/10.1046/j.1365-2486.2000.00369.x
- Louis, B.P., Maron, P.-A., Menasseri-Aubry, S., Sarr, A., Lévêque, J., Mathieu, O., Jolivet, C.,
 Leterme, P., Viaud, V., 2016. Microbial Diversity Indexes Can Explain Soil Carbon Dynamics as
 a Function of Carbon Source. PLoS One 11, e0161251.
 https://doi.org/10.1371/journal.pone.0161251
- 788 Lumley, T., 2017. leaps: Regression Subset Selection. Based on Fortran code by Alan Miller.
- Maron, P.A., Sarr, A., Kaisermann, A., Lévêque, J., Mathieu, O., Guigue, J., Karimi, B., Bernard, L.,
 Dequiedt, S., Terrat, S., Chabbi, A., Ranjard, L., 2018. High microbial diversity promotes soil
 ecosystem functioning. Appl. Environ. Microbiol. 84. https://doi.org/10.1128/AEM.02738-17
- Minasny, B., Malone, B.P., McBratney, A.B., Angers, D.A., Arrouays, D., Chambers, A., Chaplot, V.,
 Chen, Z.-S., Cheng, K., Das, B.S., Field, D.J., Gimona, A., Hedley, C.B., Hong, S.Y., Mandal,
 B., Marchant, B.P., Martin, M., McConkey, B.G., Mulder, V.L., O'Rourke, S., Richer-de-Forges,
 A.C., Odeh, I., Padarian, J., Paustian, K., Pan, G., Poggio, L., Savin, I., Stolbovoy, V.,
 Stockmann, U., Sulaeman, Y., Tsui, C.-C., Vågen, T.-G., van Wesemael, B., Winowiecki, L.,
 2017. Soil carbon 4 per mille. Geoderma 292, 59–86.
- 798 https://doi.org/10.1016/J.GEODERMA.2017.01.002
- Murphy, K.R., Stedmon, C.A., Graeber, D., Bro, R., 2013. Fluorescence spectroscopy and multi-way
 techniques. PARAFAC. Anal. Methods 5, 6557. https://doi.org/10.1039/c3ay41160e
- Murphy, K.R., Stedmon, C.A., Wenig, P., Bro, R., 2014. OpenFluor– an online spectral library of
 auto-fluorescence by organic compounds in the environment. Anal. Methods 6, 658–661.
 https://doi.org/10.1039/C3AY41935E
- 804 Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P.R.,
 805 O'Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., Wagner, H., 2018.
 806 vegan: Community Ecology Package.
- 807 Osburn, C.L., Handsel, L.T., Mikan, M.P., Paerl, H.W., Montgomery, M.T., 2012. Fluorescence
 808 Tracking of Dissolved and Particulate Organic Matter Quality in a River-Dominated Estuary.
 809 Environ. Sci. Technol. 46, 8628–8636. https://doi.org/10.1021/es3007723
- Panettieri, M., Knicker, H., Murillo, J.M., Madejón, E., Hatcher, P.G., 2014. Soil organic matter
 degradation in an agricultural chronosequence under different tillage regimes evaluated by
 organic matter pools, enzymatic activities and CPMAS 13C NMR. Soil Biol. Biochem. 78, 170–
 181. https://doi.org/10.1016/j.soilbio.2014.07.021
- Panettieri, M., Rumpel, C., Dignac, M.-F., Chabbi, A., 2017. Does grassland introduction into
 cropping cycles affect carbon dynamics through changes of allocation of soil organic matter
 within aggregate fractions? Sci. Total Environ. 576.
- 817 https://doi.org/10.1016/j.scitotenv.2016.10.073

- Pielou, E.C., 1966. The measurement of diversity in different types of biological collections. J. Theor.
 Biol. 13, 131–144. https://doi.org/10.1016/0022-5193(66)90013-0
- Plassart, P., Terrat, S., Thomson, B., Griffiths, R., Dequiedt, S., Lelievre, M., Regnier, T., Nowak, V.,
 Bailey, M., Lemanceau, P., Bispo, A., Chabbi, A., Maron, P.-A., Mougel, C., Ranjard, L., 2012.
 Evaluation of the ISO Standard 11063 DNA Extraction Procedure for Assessing Soil Microbial
 Abundance and Community Structure. PLoS One 7, e44279.
- 824 https://doi.org/10.1371/journal.pone.0044279
- Potthoff, M., Jackson, L.E., Steenwerth, K.L., Ramirez, I., Stromberg, M.R., Rolston, D.E., 2005. Soil
 biological and chemical properties in restored perennial grassland in California. Restor. Ecol. 13,
 61–73.
- Powlson, D.S., Gregory, P.J., Whalley, W.R., Quinton, J.N., Hopkins, D.W., Whitmore, A.P., Hirsch,
 P.R., Goulding, K.W.T., 2011. Soil management in relation to sustainable agriculture and
 ecosystem services. Food Policy 36, S72–S87. https://doi.org/10.1016/J.FOODPOL.2010.11.025
- Rasse, D.P., Rumpel, C., Dignac, M.F., 2005. Is soil carbon mostly root carbon? Mechanisms for a
 specific stabilisation. Plant Soil 269, 341–356. https://doi.org/10.1007/s11104-004-0907-y
- Razanamalala, K., Razafimbelo, T., Maron, P.-A., Ranjard, L., Chemidlin, N., Lelièvre, M., Dequiedt,
 S., Ramaroson, V.H., Marsden, C., Becquer, T., Trap, J., Blanchart, E., Bernard, L., 2018. Soil
 microbial diversity drives the priming effect along climate gradients: a case study in Madagascar.
 ISME J. 12, 451–462. https://doi.org/10.1038/ismej.2017.178
- Romero, C.M., Engel, R.E., D'Andrilli, J., Chen, C., Zabinski, C., Miller, P.R., Wallander, R., 2017.
 Bulk optical characterization of dissolved organic matter from semiarid wheat-based cropping
 systems. Geoderma 306, 40–49. https://doi.org/10.1016/j.geoderma.2017.06.029
- Roth, V.-N., Dittmar, T., Gaupp, R., Gleixner, G., 2014. Ecosystem-Specific Composition of
 Dissolved Organic Matter. Vadose Zo. J. 13, 0. https://doi.org/10.2136/vzj2013.09.0162
- Shahbaz, M., Kumar, A., Kuzyakov, Y., Börjesson, G., Blagodatskaya, E., 2018. Interactive priming
 effect of labile carbon and crop residues on SOM depends on residue decomposition stage:
 Three-source partitioning to evaluate mechanisms. Soil Biol. Biochem. 126, 179–190.
 https://doi.org/10.1016/J.SOILBIO.2018.08.023
- Shange, R.S., Ankumah, R.O., Ibekwe, A.M., Zabawa, R., Dowd, S.E., 2012. Distinct Soil Bacterial
 Communities Revealed under a Diversely Managed Agroecosystem. PLoS One 7, e40338.
 https://doi.org/10.1371/journal.pone.0040338
- Sheather, S., 2009. A Modern Approach to Regression with R, Springer Texts in Statistics. Springer
 New York, New York, NY. https://doi.org/10.1007/978-0-387-09608-7
- Six, J., Elliott, E.T., Paustian, K., Doran, J.W., 1998. Aggregation and soil organic matter
 accumulation in cultivated and native grassland soils. Soil Sci. Soc. Am. J. 62, 1367–1377.
- 853 Smith, P., 2014. Do grasslands act as a perpetual sink for carbon? Glob. Chang. Biol.
 854 https://doi.org/10.1111/gcb.12561
- Solomon, D., Lehman, J., Kinyangi, J., Amelung, W., Lobe, I., Pell, A., Riha, S., Ngoze, S., Verchot,
 L., Mbugua, D., Skjemstad, J., Schafer, T., 2007. Long-term impacts of anthropogenic
 perturbations on dynamics and speciation of organic carbon in tropical forest and subtropical
 grassland ecosystems. Glob. Chang. Biol. 13, 511–530. https://doi.org/10.1111/j.13652486.2006.01304.x
- Søndergaard, M., Stedmon, C.A., Borch, N.H., 2003. Fate of terrigenous dissolved organic matter
 (DOM) in estuaries: Aggregation and bioavailability. Ophelia 57, 161–176.
 https://doi.org/10.1080/00785236.2003.10409512

- Sparling, G.P., 1992. Ratio of microbial biomass carbon to soil organic carbon as a sensitive indicator
 of changes in soil organic matter. Aust. J. Soil Res. 30, 195–207.
- Stedmon, C.A., Markager, S., 2005. Resolving the variability in dissolved organic matter fluorescence
 in a temperate estuary and its catchment using PARAFAC analysis. Limnol. Oceanogr. 50, 686–
 697. https://doi.org/10.4319/lo.2005.50.2.0686
- Stubbins, A., Lapierre, J.-F., Berggren, M., Prairie, Y.T., Dittmar, T., del Giorgio, P.A., 2014. What's
 in an EEM? Molecular Signatures Associated with Dissolved Organic Fluorescence in Boreal
 Canada. Environ. Sci. Technol. 48, 10598–10606. https://doi.org/10.1021/es502086e
- 871 Swinton, S.M., Lupi, F., Robertson, G.P., Hamilton, S.K., 2007. Ecosystem services and agriculture:
- 872 Cultivating agricultural ecosystems for diverse benefits. Ecol. Econ.
- 873 https://doi.org/10.1016/j.ecolecon.2007.09.020
- Tabachnick, B.G., Fidell, L.S., 2007. Using multivariate statistics, 5th ed., Using multivariate
 statistics, 5th ed. Allyn & Bacon/Pearson Education, Boston, MA.
- Tardy, V., Spor, A., Mathieu, O., Lévèque, J., Terrat, S., Plassart, P., Regnier, T., Bardgett, R.D., van
 der Putten, W.H., Roggero, P.P., Seddaiu, G., Bagella, S., Lemanceau, P., Ranjard, L., Maron,
- P.-A., 2015. Shifts in microbial diversity through land use intensity as drivers of carbon
- 879 mineralization in soil. Soil Biol. Biochem. 90, 204–213.
- 880 https://doi.org/10.1016/J.SOILBIO.2015.08.010
- Terrat, S., Christen, R., Dequiedt, S., Lelièvre, M., Nowak, V., Regnier, T., Bachar, D., Plassart, P.,
 Wincker, P., Jolivet, C., Bispo, A., Lemanceau, P., Maron, P.-A., Mougel, C., Ranjard, L., 2012.
 Molecular biomass and MetaTaxogenomic assessment of soil microbial communities as
 influenced by soil DNA extraction procedure. Microb. Biotechnol. 5, 135–141.
 https://doi.org/10.1111/j.1751-7915.2011.00307.x
- Tiessen, H., Cuevas, E., Chacon, P., 1994. The role of soil organic matter in sustaining soil fertility.
 Nature 371, 783–785. https://doi.org/10.1038/371783a0
- van Eekeren, N., Bommelé, L., Bloem, J., Schouten, T., Rutgers, M., de Goede, R., Reheul, D.,
 Brussaard, L., 2008. Soil biological quality after 36 years of ley-arable cropping, permanent
 grassland and permanent arable cropping. Appl. Soil Ecol. 40, 432–446.
 https://doi.org/10.1016/J.APSOIL.2008.06.010
- Viaud, V., Santillàn-Carvantes, P., Akkal-Corfini, N., Le Guillou, C., Prévost-Bouré, N.C., Ranjard,
 L., Menasseri-Aubry, S., 2018. Landscape-scale analysis of cropping system effects on soil
 quality in a context of crop-livestock farming. Agric. Ecosyst. Environ. 265, 166–177.
 https://doi.org/10.1016/J.AGEE.2018.06.018
- Walter, C., Curmi, P., 1998. Les sols du bassin-versant du Coët Dan: organisation, variabilité spatiale
 et cartographie, in: Agriculture Intensive et Qualité Des Eaux. INRA Editions, Paris, France, pp.
 898 85–108.
- Walz, J., Knoblauch, C., Böhme, L., Pfeiffer, E.M., 2017. Regulation of soil organic matter
 decomposition in permafrost-affected Siberian tundra soils Impact of oxygen availability,
 freezing and thawing, temperature, and labile organic matter. Soil Biol. Biochem. 110, 34–43.
 https://doi.org/10.1016/j.soilbio.2017.03.001
- 903 Wei, T., Simko, V., 2017. R package "corrplot": Visualization of a Correlation Matrix.
- Weishaar, J.L., Aiken, G.R., Bergamaschi, B.A., Fram, M.S., Fujii, R., Mopper, K., 2003. Evaluation
 of Specific Ultraviolet Absorbance as an Indicator of the Chemical Composition and Reactivity
 of Dissolved Organic Carbon. https://doi.org/10.1021/ES030360X
- Wheeler, K.I., Levia, D.F., Hudson, J.E., 2017. Tracking senescence-induced patterns in leaf litter
 leachate using parallel factor analysis (PARAFAC) modeling and self-organizing maps. J.

- 909 Geophys. Res. Biogeosciences 122, 2233–2250. https://doi.org/10.1002/2016JG003677
- 910 Wickham, H., 2009. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.
- 911 Yamashita, Y., Kloeppel, B.D., Knoepp, J., Zausen, G.L., Jaffé, R., 2011. Effects of Watershed
- 912 History on Dissolved Organic Matter Characteristics in Headwater Streams. Ecosystems 14,
- 913 1110–1122. https://doi.org/10.1007/s10021-011-9469-z

914

Table 1 Descriptive statistic of environmental parameters for the 88 soil samples from MOSAIC project.

Variable	Units	Minimum	1 st Quartile	Median	Mean	3 rd Quartile	Maximum
Priming Effect DW soil ^a	µg C g ⁻¹ DW soil	7.97	48.0	56.5	59.6	66.8	147.5
Priming Effect/TOC ^a	µg C g ⁻¹ DW soil mg C g ⁻¹ TOC	0.24	1.98	2.48	2.50	2.96	4.96
Control Soil CO ₂ ^a	μg C-CO ₂ g ⁻¹ DW soil	86	128	154	182	191	726
Amended Soil CO_2^a	$\mu g C - CO_2 g^{-1} DW soll$ $\mu g C - CO_2 g^{-1} DW soll$	808	915	969	995	1048	1522
Control Soil CO ₂ Norm TOC ^a	$mg C-CO_2 g^{-1} TOC$	4.0	5.6	6.7	7.1	8.2	15.7
Amended Soil CO ₂ Norm TOC ^a	$mg C-CO_2 g^{-1} TOC$	21.9	35.0	41.3	41.8	45.7	78.1
Mineralized litter C _{org}	% of litter C_{org} added	29.5	33.6	34.9	35.5	37.4	42.9
m o a		10.4	21 0	22.0	25.2	2.5.0	<i></i>
TOC	$mg C g^{-1} DW soil$	13.4	21.0	23.8	25.2	26.0	69.6
TN	mg N g ⁻¹ DW soil	1.6	2.1	2.3	2.5	2.5	6.4
TOC/TN	0/	8.64	9.61	10.1	10.2	10.8	11.6
Soil δ ¹³ C signature	%0	-29.5	-27.0	-26.1	-26.3	-25.3	-24.2
WEOC DW soil	mg WEOC g ⁻¹ DW soil	0.25	0.41	0.48	0.52	0.53	1.60
WEOC/TOC	$mg WEOC g^{-1} TOC$	8.1	18.3	20.4	20.9	22.1	36.9
WEOC δ^{13} C signature	%	-29.9	-26.0	-25.2	-25.2	-23.8	-22.7
SUVA	$l mg C^{-1} m^{-1}$	0.013	0.021	0.023	0.023	0.025	0.045
Fluo CP1	RU mg ⁻¹ WEOC	0.93	1.62	1.84	1.91	2.18	3.22
Fluo CP2	$\frac{RU}{RU} \frac{mg^{-1}}{mg^{-1}} \frac{WEOC}{WEOC}$	0.73	1.22	1.42	1.50	1.77	2.68
Fluo CP3	RU mg ⁻¹ WEOC	0.26	0.51	0.58	0.57	0.64	0.91
Microbial biomass DW soil	mg DNA g ⁻¹ DW Soil	13.3	35.7	51.4	58.1	71.4	251.7
Microbial biomass/TOC	$mg DNA g^{-1} TOC$	0.56	1.46	2.21	2.32	2.83	6.055
Richness_b	Number of OTUs	573	730	754	755	786	853
Evenness_b	J' Index	0.80	0.86	0.86	0.86	0.88	0.89
Richness_f	Number of OTUs	235	419	462	473	527	742
Evenness_f	J' Index	0.55	0.64	0.67	0.66	0.69	0.74
MWD	mm	0.51	1.20	1.73	1.83	2.33	3.37
Bulk Density	$g \ cm^{-3}$	0.52	1.05	1.14	1.148	1.26	1.53
Clay	$mg g^{-1} soil$	135	165	172	178	181	356
Silt	mg g ⁻¹ soil mg g ⁻¹ soil	511 77	643 128	673 148	668 154	702 173	744 255
Sand Altitude	mg g sou M	80.7	113.0	123.4	121.1	131.5	137.6
Annuae Hydromorphy	171	0	0	125.4	0.24	0	137.6
Beven's Index		2.10	3.20	3.80	4.10	4.30	1.00
Soil pH		4.80	5.82	6.04	6.02	6.25	7.23
Olsen P	mg g ⁻¹ soil	0.02	0.16	0.14	0.02	0.195	0.33
Cu	$mg kg^{-1}$ soil	1.15	3.51	4.92	5.48	7.33	14.30
Si	$mg kg^{-1} soil$	0.03	0.04	0.05	0.05	0.06	0.09
Al	mg kg ⁻¹ soil	0.16	0.25	0.30	0.31	0.37	0.49
Fe	mg kg ⁻¹ soil mg kg ⁻¹ soil	0.34	0.47	0.50	0.54	0.58	1.31
Consultant I Francis		0	0	0.10	0.20	0.00	1.00
Grassland Frequency		0	0	0.10	0.28	0.60	1.00
No. cropped species ^b	M. Chll	1.00	3.00	3.00	3.61	4.25	8.00
Crop Litter	$Mg C ha^{-1} y^{-1}$	0	2.71	3.16	3.15	4.18	4.83
Manure	$Mg \ C \ ha^{-1} \ y^{-1}$	0	0.34	0.61	0.63	0.88	1.78

^a Cumulative values obtained after 80 days of soil incubation in microcosms

^b For permanent grassland, number of cropped species was assumed to be 1, not taking into account species

diversity within prairie.

923 Figure Captions

Figure 1 Representation of WEOM fluorescence PARAFAC components in excitation-emission
matrices. All components were previously described as humic-like components in the literature and
identified in soil DOM, as well as in natural waters. (see the text for details)

927

Figure 2 Correlation heatmap of Spearman's correlation of all measured parameters. Size and color of the circles are proportional to the r_s coefficient of correlation

930

Figure 3 Box and whisker plots of the chemical and edaphological parameters of sampled soils. Cropland (n=42); LO: low frequency grassland (n=13); MI: mid-frequency grassland (n=13); HI: high-frequency grassland (n=13); Permanent Grassland (n=7). Treatments with identified by different letters are significantly different. Capital letters are used to highlight significance levels satisfying Bonferroni's correction (p<0.005). Lowercase letters indicate significance levels at p<0.05. Circles represent outliers

937

Figure 4 Box and whisker plots of the priming effect and biological parameters of sampled soils. Cropland (n=42); LO: low frequency grassland (n=13); MI: mid-frequency grassland (n=13); HI: high-frequency grassland (n=13); Permanent Grassland (n=7). Treatments with identified by different letters are significantly different. Capital letters are used to highlight significance levels satisfying Bonferroni's correction (p<0.005). Lowercase letters indicate significance levels at p<0.05. Circles represent outliers

944

Figure 5 Outputs of the two variance partitioning models for (i) priming effect per g of soil (PE-DW)
and (ii) priming effect per g of TOC (PE/TOC). HI: high-frequency grassland; MI: mid-frequency
grassland; LO: low-frequency grassland. Significance levels: ** p<0.001, * p<0.05, ° p<0.1. (+)

and (-) represent positive or negative coefficients of relation between modeled variables anddescriptors.

- 950 df: degrees of freedom.
- 951 ‡ :variance assigned to interactions for Priming Effect/DW soil model had a negative value of -6.33%,
- 952 see text for more details

3rd 1^{st} Units Median Variable Minimum Mean Maximum Quartile Quartile $\mu g C g^{-1} DW soil$ mg C g⁻¹ TOC Priming Effect DW soil ^a 7.97 48.0 56.5 59.6 66.8 147.5 Priming Effect/TOC * 2.50 0.24 1.98 2.484.96 2.96 $\mu g C-CO_2 g^{-1} DW soil$ $\mu g C-CO_2 g^{-1} DW soil$ Control Soil CO₂^a 86 128 154 182 191 726 Amended Soil CO₂^a 915 969 995 808 1048 1522 $mg C-CO_2 g^{-1} TOC$ Control Soil CO₂ Norm TOC^a 4.0 5.6 6.7 7.1 8.2 15.7 $mg C-CO_2 g^{-1} TOC$ Amended Soil CO2 Norm TOC^a 21.9 35.0 41.3 41.8 45.7 78.1 Mineralized litter C_o % of litter C_{org} added 29.5 34.9 35.5 37.4 33.6 42.9 $mg C g^{-1} DW soil$ TOC 13.4 21.0 23.8 25.2 26.069.6 $mg N g^{-1} DW soil$ TN 1.6 2.12.3 2.5 2.5 6.4 TOC/TN 8.64 9.61 10.1 10.2 10.8 11.6 Soil δ¹³C signature ‰ -29.5 -27.0 -26.1 -26.3 -25.3 -24.2 mg WEOC g⁻¹ DW soil mg WEOC g⁻¹ TOC WEOC DW soil 0.25 0.41 0.48 0.52 0.53 1.60 WEOC/TOC 18.3 20.4 20.9 22.1 36.9 8.1 WEOC δ¹³C signature -29.9 -26.0 -25.2 -23.8 -22.7 ‰ -25.2 $l mg C^{-1} m^{-1}$ **SUVA** 0.013 0.021 0.023 0.023 0.025 0.045 RU mg⁻¹ WEOC 0.93 1.84 1.91 Fluo CP1 1.62 2.18 3.22 RU mg⁻¹ WEOC Fluo CP2 0.73 1.22 1.42 1.50 1.77 2.68 Fluo CP3 RU mg⁻¹ WEOC 0.51 0.58 0.64 0.91 0.26 0.57 $mg DNA g^{-1} DW Soil$ 35.7 51.4 71.4 251.7 Microbial biomass DW soil 13.3 58.1 $mg DNA g^{-1} TOC$ Microbial biomass/TOC 0.56 2.21 2.32 2.83 6.055 1.46 **Richness** b Number of OTUs 573 730 754 755 786 853 Evenness_b J' Index 0.800.86 0.86 0.86 0.88 0.89 Number of OTUs 527 742 Richness_f 235 419 462 473 0.55 0.67 0.69 0.74 J' Index 0.64 0.66 Evenness_f MWD 2.33 0.51 1.20 1.73 1.83 3.37 тт g cm⁻³ **Bulk Density** 0.52 1.05 1.14 1.148 1.26 1.53 mg g⁻¹ soil 135 165 172 178 181 356 Clay $mg g^{-1} soil$ $mg g^{-1} soil$ $mg g^{-1} soil$ 702 744 Silt 511 643 673 668 Sand 77 128 148 154 173 255 Altitude М 80.7 113.0 123.4 121.1 131.5 137.6 0 0 1.00 Hydromorphy 0 0 0.24 **Beven's Index** 2.10 3.20 3.80 4.10 4.30 17.0 Soil pH 4.80 6.04 6.25 7.23 5.82 6.02 $mg g^{-1} soil$ 0.195 Olsen P 0.02 0.16 0.14 0.15 0.33 $mg kg^{-1} soil$ 4.92 5.48 7.33 14.30 Cu 1.15 3.51 mg kg⁻¹ soil mg kg⁻¹ soil mg kg⁻¹ soil mg kg⁻¹ soil 0.05 Si 0.03 0.04 0.05 0.06 0.09 Al 0.16 0.25 0.30 0.31 0.37 0.49 0.47 0.50 0.54 0.58 0.34 1.31 Fe 0 0 Grassland Frequency 0.10 0.28 0.60 1.001.00 3.00 3.00 4.25 8.00 3.61 No. cropped species $Mg \ C \ ha^{-1} \ y^{-1}$ **Crop Litter** 0 2.71 3.16 3.15 4.18 4.83 $Mg C ha^{-1} y^{-1}$ 0 0.34 0.61 0.63 0.88 1.78 Manure

 Table 1 Descriptive statistic of environmental parameters for the 88 soil samples from MOSAIC

 project

^a Cumulative values obtained after 80 days of soil incubation in microcosms

^b For permanent grassland, number of cropped species was assumed to be 1, not taking into account species diversity within prairie.

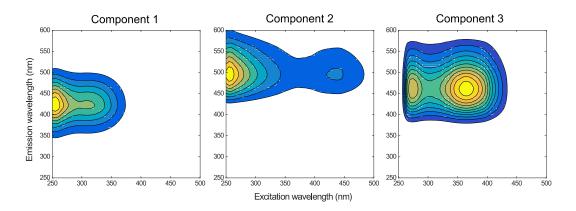
		Cropland			LO		MI		HI		Permanent Grassland	
			Std.		Std.		Std.		Std.		Std.	
Variables	Units	Mean	deviation	Mean	deviation	Mean	deviation	Mean	deviation	Mean	deviation	
	N J DW 1	2 220	0.070	0.115	0.001	0.000	0.170	0.405	0.555	4 600 1	1 1 2 0	
TN	mg N g ⁻¹ DW soil	2.238 a	0.272	2.115 a	0.321	2.369 a	0.170	2.485 a	0.555	4.600 b	1.130	
TOC/TN	0/	10.2	0.7	10.2	0.8	10.1	0.6	10.1	0.5	10.2	0.8	
Soil δ^{13} C signature	%	-25.68	0.78	-25.44	0.60	-26.71	0.70	-27.42	0.68	-28.78	0.63	
SUVA	$l mg C^{-1} m^{-1}$	2.269	0.242	2.631	0.750	2.285	0.316	2.262	0.491	2.129	0.718	
WEOC δ^{13} C signature	%	-24.44	1.01	-24.04	0.84	-25.70	0.91	-26.76	0.73	-27.65	2.08	
CP1 _(fluo)	$RU mg^{-1} WEOC$	1.868	0.328	2.106	0.498	1.832	0.359	2.024	0.592	1.683	0.590	
CP2 _(fluo)	RU mg ⁻¹ WEOC	1.480	0.295	1.632	0.447	1.420	0.343	1.574	0.565	1.374	0.485	
Richness_b	Number of OTUs	757 A	38	773 AB	25	770 AB	36	742 AB	48	697 A	68	
Richness f	Number of OTUs	451 a	90	457 ab	63	508 b	91	494 ab	79	527 ab	107	
Evenness_f	J' Index	0.654 A	0.035	0.648 A	0.025	0.670 AB	0.030	0.697 B	0.023	0.693 B	0.025	
Silt	g kg ⁻¹ soil	679.0 B	32.1	667.3 B	56.8	676.1 B	14.7	669.7 B	60.5	583.7 A	37.8	
Sand	g kg ⁻¹ soil	148.9	29.1	164.4	46.3	152.2	13.8	144.4	32.2	181.6	66.4	
Bulk Density	g cm ⁻³	1.122	0.179	1.212	0.127	1.108	0.165	1.234	0.161	1.101	0.197	
Si	mg kg ⁻¹ soil	0.502	0.100	0.538	0.171	0.438	0.087	0.431	0.085	0.443	0.113	
41	mg kg ⁻¹ soil	3.29	0.75	3.20	0.69	2.95	0.74	2.66	0.62	2.71	0.69	
Fe	mg kg ⁻¹ soil	5.29	1.03	5.28	0.70	5.05	1.05	5.16	0.59	7.37	3.42	
Cu	mg kg ⁻¹ soil	5.88	3.06	5.34	1.69	5.45	2.64	5.14	3.02	4.02	2.90	
Altitude	m	122.1 AB	9.9	116.9 AB	9.8	125.0 AB	8.0	127.4 B	8.3	103.8 A	21.1	
Beven's index		4.21	2.36	4.01	0.73	4.11	1.44	3.62	0.78	4.86	2.94	
Manure	$Mg C ha^{-1} y^{-1}$	0.725 B	0.330	0.513 AB	0.286	0.785 B	0.387	0.585 AB	0.176	0.136 A	0.269	
No. cropped species*		3.86 B	1.07	4.46 B	1.85	3.92 B	0.49	2.92 AB	0.64	$1.00 \mathbf{A}$		

Table S1 Means and standard deviations of the measured parameters not selected as PE descriptors for each land-use

Significant differences at p < 0.05 are marked with different lowercase letters. Significant differences at p < 0.005 are marked with different capital letters. LO: low-frequency grassland; MI: mid-frequency grassland; HI: high-frequency grassland;

*For permanent grassland, number of cropped species was assumed to be 1, not taking into account species diversity within prairie.







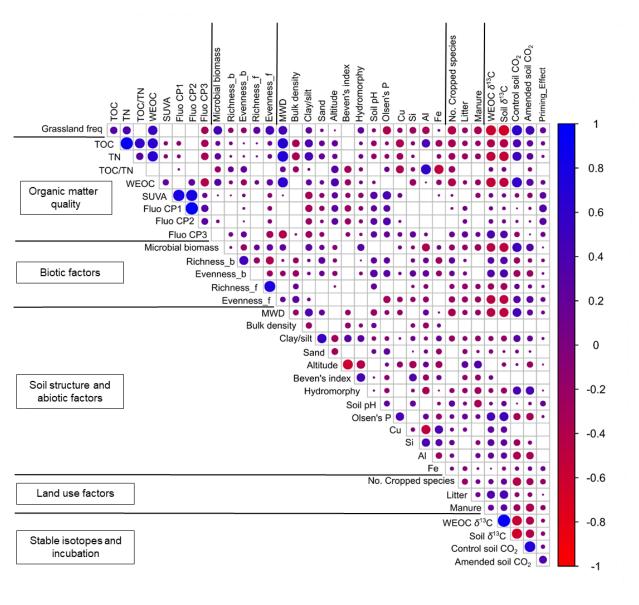


Figure 3 Click here to download Figure : Figure 3REV_d.pdf

