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1 Nitrogen benefits of ten legume pre-crops for wheat assessed by

2 field measurements and modelling

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10 Highlights

- N pre-crop effects of spring and summer legumes were assessed in two biennial field trials
- The STICS model accurately predicted soil N and humidity and was used to calculate N
 mineralisation and leaching
- Legumes had a positive effect on wheat grain and shoot N yields compared to cereals
- Part of soil N was not retrieved by the following wheat and was leached
- Half of wheat shoot N variability was explained by soil nitrogen availability

17 Abstract

- 18 The positive effect of grain legume pre-crops on the yield of the subsequent crop has been studied
- 19 widely, whereas less information is available on the nitrogen (N) processes related to this positive
- 20 effect, especially for a wide range of grain legume species.
- 21 The objective was to quantify and understand the effect of grain legume compared to cereal pre-
- 22 crops (sown in 2014 and 2016) on grain and shoot N yields and shoot N concentration of wheat
- 23 (Triticum aestivum) grown the following year (2015 and 2017). Spring legumes (faba bean (Vicia
- 24 faba), fenugreek (*Trigolia foenum-graecum*), common vetch (*Vicia sativa*), lentil (*Lens culinaris*), lupin

(Lupinus albus), and pea (Pisium sativum)) were compared to barley (Hordeum vulgare). Summer legumes (chickpea (Cicer arietinum), common bean (Phaseolus vulgaris), soybean (Glycine max) and Narbonne vetch (Vicia narbonensis) were compared to sorghum (Sorghum bicolor). Inorganic N remaining in the soil at pre-crop harvest (N sparing) was measured. The STICS model which accurately predicted soil humidity and soil inorganic N in the pedoclimatic conditions of the field experiments was used to calculate N mineralisation from pre-crop residues and N leaching between pre-crop harvest and wheat harvest. Grain and shoot N yields of unfertilised N wheat were respectively 27 and 25% higher after faba bean and lentil compared to barley pre-crops, and 66 and 51% higher after summer legumes compared to sorghum pre-crop. In the second experiment, N fertilisation reduced the positive effect of fenugreek, and lentil on wheat yield compared to barley while it did not modify the pre-crop effect of the other legumes. A positive relationship between N mineralisation from pre-crop residues and wheat shoot N yield was established, with higher amounts of N available for the subsequent wheat after legume pre-crops. In 2014, N sparing was higher after spring pre-crops compared to summer pre-crops inducing higher N leaching after spring legume pre-crops, especially in the first experiment which was characterised by heavy rain in summer and autumn. Estimating N availability by taking into account N sparing, N mineralisation from soil and pre-crop residues and N leaching explained 49% of wheat shoot N yield variability. Unquantified N processes and non-N processes might also have contributed to the positive effects of legumes on the subsequent wheat.

Key words

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45 Pre-crop effect; Grain legumes; Crop residues; Mineralisation; N leaching

1. Introduction

- 47 Cereal crops generally have higher yields when cultivated after unrelated species (Kirkegaard et al.,
- 48 2008) and especially after legume crops (Angus et al., 2015; Preissel et al., 2015). Some studies also
- 49 highlighted higher grain N concentrations in cereals grown after legumes compared to cereals grown

after cereals (Biederbeck et al., 1996; Gan et al., 2003). Part of the positive effect of legumes on the following cereals are due to N benefits. Indeed, a substantial amount of N can be left by legume crop residues. Moreover, during crop residue decomposition by soil microbes, organic N is converted into inorganic forms that can be taken up by the subsequent crop. Much of the inorganic N released is rapidly immobilised by soil microbial biomass. Inorganic N accumulation in the soil occurs when the amounts of N released exceeds the microbial N requirements (net N mineralisation). Conversely, when inorganic N released from the mineralisation of crop residues is not sufficient, micro-organisms will also assimilate soil inorganic N to meet their N requirements, which results in net N immobilisation (Nicolardot et al., 2001). Net N mineralisation or smaller amounts of net N immobilisation are expected for legume residues compared to cereal residues, since legume residues are generally characterised by higher N concentrations and lower C:N ratios. Furthermore, part of the additional soil inorganic N after legume crops can be attributed to N "sparing" during legume growth (Evans et al., 1987; Herridge et al., 1995). Legume superficial root systems compared to those of cereals can lead to higher amounts of inorganic N remaining in the soil, especially in deeper layers (Hauggaard-Nielsen et al., 2001). A positive relationship between wheat yields and the amount of inorganic N left in the soil after several pre-crops was established by Angus et al. (2015) for dry land field experiments, and legumes systematically presented the highest post-harvest amounts of inorganic N. However, the effects of legume crops on the following cereals are highly variable (Kirkegaard et al., 2008; Preissel et al., 2015) and they are difficult to predict. Part of this variability can be attributed to the diversity of legume species, environmental conditions and agronomic practices. Legumes species may differ according to the C:N ratio of crop residues (Palm et al., 2001) or their ability to take up soil inorganic N during their growth cycle (Dayoub et al., 2017; Guinet et al., 2018). The amounts of inorganic N available for the following cereals, derived from N mineralisation of crop residues and/or N "sparing" may therefore depend on legume pre-crop species.

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For a given pre-crop species, the effect on the following wheat can vary considerably with pedoclimatic conditions (Cernay et al., 2017). Hence, N fluxes occurring after pre-crop harvest that determine the amount of inorganic N available for the following cereal are highly dependent on pedoclimatic conditions. On the one hand, N mineralisation from crop residues is positively stimulated by high soil moisture and temperature (Abera et al., 2012). On the other hand, soil inorganic N can be lost by N leaching if heavy rainfall occurs when high amounts of inorganic N are present in the soil. In European climatic conditions, N leaching mainly occurs during the autumnwinter period, when the soil is bare after pre-crop harvest and/or because the following cereal crop requires only very small amounts of inorganic N during its early development. Higher amounts of N losses by leaching are often measured after legume crops compared to cereals because of higher N sparing and the higher N mineralisation from legume residues (Plaza-Bonilla et al., 2015). Moreover, N mineralisation from crop residues and N loss dynamics may also vary according to legume growth cycle. Early harvested legumes (i.e. spring legumes) induce a longer fallow period before the sowing of the following crop, compared to legumes harvested later in the season (i.e. summer legumes). Hence, legume growth cycles could affect N leaching by modulating the synchronisation between N supply by legume pre-crops and N requirements of the following crop. However, N fluxes occurring between pre-crop harvest and the subsequent crop harvest are particularly difficult to quantify in field conditions. Therefore, their estimation often requires agronomic dynamic models that take into account the impact of both crop residues characteristics (i.e. C:N ratio) and pedoclimatic conditions on N fluxes after pre-crop harvest, thus allowing the calculation of N budgets over time. Cereal performance after legumes are also affected by agricultural practices. Several studies showed that N fertilisation on the following cereal crops reduces the yield gap between a cereal cultivated after legumes and a cereal cultivated after cereals (Cernay et al., 2017; Jensen et al., 2004). Hence, in situations where high levels of N fertilisation are applied, the effect of legumes on the subsequent cereal performance is likely to be negligible.

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Although it is widely acknowledged that legumes have a positive effect on subsequent crop performance (Angus et al., 2015; Cernay et al., 2017; Espinoza et al., 2012; O'Donovan et al., 2014; Preissel et al., 2015; Walley et al, 2007; Williams et al., 2014), it is unclear to which extent N processes explain legume positive pre-crop effect on the following crop, because of the difficulty to measure some N fluxes occurring after legume harvest. Moreover, little is known about the effect of legume growth cycle on the dynamic of N fluxes occurring after legume harvest and on the performance of the following crop. Hence, the aims of our study were the following: i) to compare the effect of ten legume and two cereal pre-crops on the subsequent performance of wheat; ii) to quantify N fluxes occurring between the pre-crop harvest and the subsequent wheat harvest through field measurements and modelling, and evaluate how they were affected by pre-crop (spring and summer) and climatic conditions; and iii) to evaluate whether the subsequent wheat performance could be explained by those N fluxes. To achieve these aims, a two-year field experiment was repeated twice between 2014 and 2017; i.e. the first year, legume and cereal pre-crops were cultivated in the same field and followed by wheat the second year, after the incorporation of pre-crop residues in the soil. Simulations with STICS model were performed to estimate soil N fluxes occurring during the period spanning from pre-crop harvest to wheat harvest: N mineralisation of pre-crop residues and soil organic-matter, and N leaching.

2. Materials and Methods

2.1. Field experiments

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2.1.1. Site and climatic conditions

Two field experiments, each lasting two years were carried out at the INRA Dijon experimental site of
Bretenière (Eastern France; 47.241 N, 5.115 E, altitude = 206 m) in 2014-2015 (Experiment I) and 2016-2017 (Experiment II). The soil is classified as Cambisol (Eutric) (WRB, 2006) with a clayey surface
layer (depth = $0.65 \pm 0.15 \text{ m}$) developed on an alluvial coarse layer. Before the beginning of both

characteristics (Table 1).

The site is subject to a semi-continental climate but the climatic conditions were contrasted between Experiments I and II. Cumulative rainfall and mean temperature between the beginning of pre-crop harvest and wheat harvest were 850 mm and 11.8°C in Experiment I and 600 mm and 11.3 °C in

experiments (2014 and 2016), soil samples were collected at 30 cm depth to determine soil

Experiment II (Fig. 1). Rainfall were unusually high between July and November 2014 (541 mm) compared to 2016 (303 mm) and to the mean rainfall over the 20 past years during the same period (362 mm). Rainfall between January and June were similar in 2015 (252 mm) and 2017 (293 mm), but

they were lower than the rainfall averaged over the 20 past years during the same period (340 mm).

2.1.2. Experimental design

Pre-crop treatments: Each experiment consisted of a two-year crop succession. Legume and cereal pre-crop treatments were sown in 2014 and followed by winter wheat that was harvested in 2015 (Experiment I). In 2016, the same pre-crops were grown in a field nearby the first experiment, and followed by winter wheat harvested in 2017 (Experiment II). This resulted in two separate data sets that allowed us to evaluate the effect of pre-crops on the following wheat performance.

In Experiment I and II, nine legume pre-crops were included: faba bean (*Vicia faba*, cv. Expresso), chickpea (*Cicer arietinum*, cv. Twist in 2014 and Vulcano in 2016), common bean (*Phaseolus vulgaris*, cv. Flavert), common vetch (*Vicia sativa*, cv. Candy), lentil (*Lens culinaris*, cv. Anicia), lupin (*Lupinus albus*, cv. Feodora), pea (*Pisium sativum*, cv. Kayanne), soybean (*Glycine max*, cv. Sultana(000)) and Narbonne vetch (*Vicia narbonensis*, cv. Clara). In Experiment II, fenugreek (*Trigolia foenum-graecum*, cv. Fenu-fix) was also cultivated (Table 2). Pre-crops were either sown in March or in May, depending on the physiology of each species (Table 2) and were referred as spring pre-crops and summer precrops, respectively, thereafter. Legume seeds were inoculated at sowing with species-specific strains of N₂-fixing bacteria to ensure symbiotic N₂ fixation, as described in Guinet et al. (2018). No N

fertilisation was applied to any of the legume pre-crops in either experiment.

Two N fertilised cereal pre-crops were also grown in Experiments I and II: barley (Hordeum vulqare, cv. Irina) and sorghum (Sorghum bicolor, cv. Quebec); they were sown in March and May, respectively. Nitrogen fertilisation was applied in form of NH₄NO₃ at rates of 60 and 75 kg N ha⁻¹ in 2014 (3rd and 28th April, respectively) for barley. In 2016, due to the extremely wet conditions in April and May, only 70 kg N ha⁻¹ were applied on barley on the 19th of April to reduce the risk of N leaching. N fertiliser was applied twice during sorghum growth at rates of 50 kg N ha⁻¹ each in 2014 (9th May and 13th June) and 2016 (7th June and 7th July)Pre-crops were sown on plots 1.5 m wide and 12 m long with four replicates per pre-crop in Experiment I and eight replicates in Experiment II. Sowing densities are summarised in Table 2. Pre-crop management: In order to reduce water stress, irrigation was supplied on pre-crop treatments. In Experiment I, spring pre-crops were irrigated with 35, 40 and 50 mm on March 26th, May 16th and June 18th 2014, respectively. Summer pre-crops were irrigated with 40, 40 and 50 mm on May 16th, May 27th and June 14th 2014, respectively. In Experiment II, 50 mm was supplied to summer pre-crops on July 11th 2016. Weeds were controlled by hand weeding and one insecticide (Karate Zeon) was applied on April 25th 2014 and April 21th 2016 in Experiment I and II, respectively. Wheat following all pre-crop treatments: Pre-crop grains were harvested with a harvesting machine. No grains were harvested for chickpea in 2014 and Narbonne vetch in 2014 and 2016, due to climatic conditions unsuitable for grain production. In these cases, the whole plant was considered as crop residues. Two to seven weeks after pre-crop grain harvest, crop residues were chopped and incorporated into the soil using a rototiller (see Table 2 for pre-crop incorporation dates). Thereafter, winter wheat (Triticum aestivum, cv. Rubisko) was sown on the same plots on October 20th 2014 (300 seeds m⁻²) in Experiment I and October 24th 2016 (350 seeds m⁻²) in Experiment II. In order to control weeds and pathogens, an herbicide (February 26th 2015, Arbalète) and a fungicide (April 30th 2015, Adexar + Voxan) were applied on wheat in Experiment I. In Experiment II, irrigation was applied on wheat on April 5th and 19th 2017 (27 mm and 30 mm, respectively) (Fig. 1B).

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176 Wheat N treatments: In order to express pre-crop maximum effect on wheat, no N fertiliser was 177 applied in Experiments I and II, corresponding hereafter to the ON treatment. 178 In Experiment II, an additional N fertilisation treatment on wheat was studied. At the end of wheat tillering (April 4th 2017), 40 kg N ha⁻¹ in the form of NH₄NO₃ was applied on half of the trial (with four 179 180 replicates per pre-crop treatment), corresponding to the 40N treatment. 181 2.1.3. Sampling and measurements Pre-crop samplings: The above-ground biomass of legume and cereals pre-crops was sampled at 182 183 physiological maturity (Table 2) on subplots of 1 m². Straw was separated from grains and considered 184 as crop residues left on the soil at maturity (except for chickpea in 2014 and Narbonne vetch in 2014 185 and 2016 where the whole plant was considered as crop residues). 186 Wheat samplings: At physiological maturity, wheat was manually harvested on subplots of 1 m² and 187 straws were separated from grains. In order to obtain a better estimation of the wheat grain yield, 188 grains were also harvested on 6.5 m² using a harvesting machine. 189 Measurements: Pre-crop residues and wheat straw and grain samples were oven-dried for 48 h at 190 80°C before dry matter was determined. Samples were milled and N and C concentrations were 191 measured using an elemental analyser (ANCA-GLS, Sercon Ldt., Crewe, UK). 192 2.1.4. Calculations 193 The amount of N in pre-crop residues was calculated by multiplying dry matter of residues by N 194 concentration of residues (Table S1). 195 Total wheat shoot dry matter was calculated by multiplying wheat grain yield measured on the 6.5 196 m² surface by total wheat dry matter/wheat grain dry matter ratio measured on the 1 m² subplot. 197 The total amount of wheat shoot N (i.e. shoot N yield) was then calculated by multiplying wheat 198 shoot dry matter by N concentration of wheat shoot.

2.2. Estimation of N fluxes using STICS model

The STICS model (Simulateur mulTI disciplinaire pour les Cultures Standards) was developed at INRA (France) to simulate crop growth, soil water, carbon and nitrogen balances as influenced by climatic conditions, over either one crop cycle or several cultural successions at a daily time-step (Brisson et al. (2003). The model STICS v8.50 was used to simulate N fluxes, such as N leaching, N mineralisation from pre-crop residues and soil organic matter, between the pre-crop harvest and the winter wheat harvest in Experiments I and II.

To run the simulations, a number of input variables are required, such as soil characteristics, initial

2.2.1. Model inputs

soil and pre-crop residue states, crop characteristics, crop management and climatic conditions during the simulated period.

Soil Parameters: Permanent soil characteristics (clay, organic N, calcium, pH, etc.) were defined and fixed using measurements carried out at the beginning of Experiments I and II (Table 1). Soil moisture content at field capacity was estimated as the median value of water content measured on soil core samples in mid-winter. Soil moisture content at wilting point and bulk density was estimated according to Ugarte Nano et al. (2016), who measured these soil characteristics in a neighbouring field experiment with standard reference farming practices as in the field prior to our experiment. Maximal rooting depth was set at 60 cm because of the presence of an alluvial coarse layer, greatly limiting root exploration below this layer. The thickness of the biological active layer for N mineralisation from soil and pre-crop residues was set at ploughing depth.

Soil initialisation: soil inorganic N and soil moisture measurements from soil core samplings performed shortly before or after pre-crop harvest were used to initiate the soil profile. In Experiment I, soil core samplings were performed on August 5th 2014 for spring pre-crops and on September 3rd 2014 for summer pre-crops. In Experiment II, soil core samplings were performed on

August 26th 2016 for spring pre-crops and on September 23rd 2016 for summer pre-crops.

224 <u>Crop residue inputs</u>: Pre-crop residue dry matter and its physicochemical characteristics (C:N ratio, C
 225 concentrations, water content) at pre-crop harvest were used to initiate the simulations.

<u>Wheat management</u>: Winter wheat is one of several crops parametrised in STICS, using physiological and morphological traits. Crop management specified dates and methods of soil tillage, wheat sowing dates, sowing depth, sowing densities, row spacing, and date and amounts of water applied by irrigation.

<u>Climatic inputs</u>: Climatic conditions were measured by a weather station located at the INRA experimental site of Bretenière. Minimum and maximum temperatures, rainfall, global radiation and Penman evapotranspiration were specified in the model at a daily time-step.

2.2.2. Model simulations and evaluation

A simulation was performed for each pre-crop treatment and for Experiments I and II focusing on the ON wheat treatment, leading to a total of 23 simulations. The model was evaluated for its ability to predict the daily amount of inorganic N and water content in the soil profile. Soil core samplings in the O-60 cm soil layer were performed monthly on wheat plots for all pre-crop treatments, between January and July 2015 in Experiment I, and between October 2016 and July 2017 in Experiment II, with a 5 cm diameter auger.

The amounts of soil inorganic N and soil moisture measured from the soil samples were expressed in kg N ha⁻¹ and mm, respectively and compared with the simulated data on the same date and for the same pre-crop treatment.

Three statistical criteria were used (Wallach et al., 2013) based on the comparison between model predictions and observed values: RRMSE, Bias and maximal error (ME):

$$RRMSE = \frac{\sqrt{\frac{1}{n}\sum_{i=1}^{n}(yi-\hat{y}i)^{2}}}{\overline{y}}$$
 (1)

Bias =
$$\frac{1}{n}\sum_{i=1}^{n}(\mathbb{I}_{i}-\hat{y}_{i})$$
 (2)

$$ME = \max | \mathbb{Z}i - \hat{y}i|_{i=1}^{n}$$
 (3)

where yi is the observed value; ŷi the simulated value; ② the mean observed values and n the number of observations (sampling dates x pre-crop treatments).

From the simulations, the cumulative amounts of N leached under the soil profile, and N mineralisation from pre-crop residues and soil organic matter were calculated.

In order to understand the effect of pre-crop treatments on the N fluxes and their link with wheat performance, wheat shoot N yield was plotted as a function of the cumulative amount of N mineralisation from pre-crop residues, which was calculated with the STICS model.

The amount of inorganic N actually available for wheat during wheat growth cycle was calculated as follows (eq. 4):

Wheat shoot N yield measured in the field experiments was then plotted as a function of N available for wheat.

2.3. Data analysis

Statistical analyses were performed with R software version 3.4.2. (R Core Team, 2016). Two-way analysis of variance (ANOVA) were performed to test for year effects (2015 and 2017), pre-crop effects and year x pre-crop interaction effects on wheat grain yield, shoot N concentration and shoot N yield, using data collected for unfertilized N wheat. ANOVAs were also performed to test for N rate effects (0N and 40N), pre-crop effects and N rate x pre-crop interaction effects on the same three response variables, using data collected in 2017 for fertilised and unfertilised N wheat. In both cases spring and summer pre-crops were analysed separately. When no pre-crop by year or N rate interactions were detected, pre-crop values were averaged over year or N rate. If the effects were significant, contrasts among pre-crops were adjusted using the package {emmeans}.

The effect of year, legume growth cycle (spring and summer) and their interaction on Nleaching, was also analysed. The homoscedasticity and normality of the residuals of the response variables were tested using a Bartlett and Shapiro-Wilk test, respectively.

For all data analysis, significant differences among treatments were identified at the 0.05 probability level of significance.

A multivariate analysis was performed to determine the relation between pre-crop characteristics, N fluxes, environmental conditions and wheat performance. Pre-crop treatments were ordered through a Principal Component Analysis (PCA) using the Canoco 5.10 program (ter Braak and Šmilaueur, 2012). The variables were standardised to obtain equal weights in the PCA.

3. Results

For both Experiments I and II, once the grains were harvested, the crop residues were incorporated in the soil at two different dates for spring and summer pre-crops (Table 2). Because of the timespan between the two incorporation dates, crop residues from spring and summer pre-crops were subjected to different environmental conditions that may have affected their N mineralisation and therefore the performance of the following wheat. Hence, in the following section, the performance of wheat cultivated after spring and summer pre-crops are presented separately.

3.1. Effect of pre-crop treatments and year on unfertilised wheat performance

For spring pre-crops, species had a significant effect on grain and shoot N yields of unfertilised wheat, but the interaction between year and pre-crop species was not significant (Table 3). On average for both years (2015 and 2017), only faba bean and lentil increased significantly wheat grain yield (+1.1 and +9 t ha⁻¹, respectively) and wheat shoot N yield (+ 18 and + 13 kg N ha⁻¹, respectively) compared to barley. Conversely, wheat shoot N concentration presented no differences among spring pre-crop treatments.

For summer pre-crops, all legume pre-crops induced higher wheat grain yields compared to wheat cultivated after sorghum with an average increase of 1.6 t ha⁻¹. The interaction between year and

pre-crop was significant for wheat shoot N yield. Except for soybean in 2017, all legume pre-crops induced higher wheat shoot N yields compared to wheat cultivated after sorghum with an average increase of 24 kg N ha⁻¹ in 2015 and 26 kg N ha⁻¹ in 2017. For both experiments, Narbonne vetch pre-crop treatment always induced the highest wheat grain yield and shoot N yields. Conversely, for both years, wheat cultivated after sorghum, presented a slightly higher shoot N concentration compared to wheat after legume summer pre-crop treatments.

Compared to sorghum, summer legume pre-crops induced higher wheat grain yield and shoot N yield benefits than spring legume pre-crops compared to barley. This was explained by the choice of reference cereal pre-crop. Indeed, wheat after sorghum always presented lower grain yields than wheat after barley for both years (2015 and 2017) (Table 3).

3.2. Effect of N fertilisation on wheat performance

In 2017, N fertilisation had a significant effect on grain yield, shoot N concentration and shoot N yield of wheat cultivated after all spring and summer pre-crops (Table 4). On average, for wheat cultivated after legume and cereal spring pre-crops, N fertilisation increased grain yield, shoot N concentration and shoot N yield by 1.2 t ha⁻¹, 0.10% and 27 kg N ha⁻¹, respectively. Considering all summer pre-crops, wheat N fertilisation increased wheat grain yield by 1.6 t ha⁻¹, shoot N concentration by 0.04% and shoot N yield by 29 kg N ha⁻¹. For spring pre-crops, the interaction between N rate and species was significant for wheat grain and shoot N yields. While grain yield of unfertilised wheat was greater after lentil and fenugreek compared to barley, it was no longer the case when wheat was N fertilised (40N treatment). Likewise, wheat shoot N yield after lentil was not significantly different from wheat after barley when N fertilisation was applied. For faba bean and common vetch and for all summer pre-crops, grain and shoot N yields of unfertilised (0N) and fertilised (40N) wheat were always higher after all legume pre-crops compared to cereal pre-crops. There were no longer significant differences of wheat shoot N concentration between legumes and sorghum pre-crops when wheat was N fertilised.

3.3. Estimation of N fluxes between pre-crop harvest and wheat harvest

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324 To evaluate the model's predictive performance in our pedoclimatic conditions, simulated data were 325 compared with measured soil water content and soil inorganic N, which are known to be strong 326 drivers of NO₃- leaching, N mineralisation from pre-crop residues and soil organic matter (Fig. 2). The 327 STICS model was efficient for predicting soil water content, leading to a small RRMSE (0.092) (Fig. 328 2A). The RRMSE (0.462) was higher for soil inorganic N (Fig. 2B). 329 There was a slight tendency to over-estimate both variables, as we obtained an overall negative bias 330 but with small absolute values: -8.9 mm for soil water (Fig. 2A) and -3.9 kg N ha-1 for soil inorganic-N 331 (Fig. 2B). Soil inorganic-N was systematically under-estimated in Experiment I (bias = +12.2 kg N ha⁻¹) while it was always over-estimated in Experiment II (bias = -14.2 kg N ha⁻¹). However, the 332 333 classification between pre-crop species according to soil inorganic N was correctly simulated in both 334 experiments. Therefore, for a given experiment, simulations can be used to compare pre-crop 335 treatments but absolute values need to be considered with caution when comparing experiments. 336 Nitrogen mineralisation from crop residues was simulated for all pre-crops and both experiments 337 (Fig. 3A: Experiment I and 3B: Experiment II). Except for Narbonne vetch, rapid N immobilisation 338 occurred just after the incorporation of crop residues in the soil, followed by net N mineralisation a 339 few weeks later. Because summer pre-crop residues were incorporated several weeks after spring 340 pre-crop residues, we observed a delay in the dynamics of N mineralisation from pre-crop residues 341 between spring (Fig. 3: dark colours) and summer (Fig. 3: light colours) pre-crop residues. At the end 342 of the simulations, the cumulative amounts of N mineralisation from pre-crop residues ranged from -343 24 kg N ha⁻¹ (barley) to +32 kg N ha⁻¹ (Narbonne vetch) in Experiment I and from -33 kg N ha⁻¹ (sorghum) to +35 kg N ha⁻¹ (Narbonne vetch) in Experiment II. 344 345 At the beginning of the simulation, the total amount of soil inorganic N was higher after spring pre-346 crops compared to summer pre-crops in Experiment I (Fig 3C). Consequently, soil inorganic N was higher during the whole simulation period for spring pre-crop treatments compared to summer pre-347

crops, but with smaller differences near the end of Experiment I. In Experiment II (Fig 3D) the mean amount of soil inorganic N at the beginning of the simulation was on average 53 kg N ha⁻¹.

The simulated amounts of soil water (Fig. 3E: Experiment I and 3F: Experiment II), were also higher at

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Experiment I.

simulation initiation for spring pre-crop treatments compared to summer pre-crops in Experiment I and for barley in Experiment II. Once rainfall had filled the soil profile up to full water capacity, the amount of soil water was identical for all pre-crop treatments. However, in Experiment I, soil reached field water capacity almost immediately after initiating the simulations for spring pre-crop treatments while it was reached two months later for summer pre-crops (October 12th 2014). In Experiment II, field soil water capacity was reached even later for all pre-crop treatments (between November 4th 2016 for barley pre-crop treatment and November 21th 2016 for chickpea, common bean and sorghum). In both experiments, most of the N leaching occurred between wheat sowing and the end of winter (Fig. 4) as the soil remained at maximum water capacity almost throughout the period (Fig 3E and F). The soil profile was regularly filled by rainfall. During this period, N leaching in Experiment I was on average 77 kg N ha⁻¹ for spring pre-crop treatments, with values ranging from 63 kg N ha⁻¹ (pea) to 93 kg N ha⁻¹ (common vetch) (Fig. 4C). For summer pre-crop treatments the average value of N leaching during the same period was 52 kg N ha⁻¹ (Fig 4D). Based on all pre-crop treatments, N leaching was higher in Experiment I compared to Experiment II (F(1,19)=55.51, p < 0.001), because of lower rainfall in Experiment II, especially in November (Fig. 1). Considering both experiments, N leaching was higher after spring pre-crops compared to summer pre-crops (F(1,19)=14.68 p < 0.01), especially in

3.4. Relationships between wheat performance, pre-crop characteristics, and pedoclimatic conditions The first and second axes of the PCA accounted for 42.5 and 32.7%, respectively, of the total variance (Fig. 5). The cumulative amounts of $N-NO_3^-$ leached between pre-crop harvest and wheat harvest (19%), cumulative rainfall during the simulation period (14%), water content in the soil profile (11%) and soil inorganic N content (11%) at the beginning of the simulation accounted the most for the first

axis. These four variables were positively correlated but all four were correlated negatively with wheat shoot N concentration. C:N ratio of pre-crop residues (21%) and the cumulative amounts of N mineralised from pre-crop residues (20%) contributed the most to the second axis and were correlated negatively. Conversely, the cumulative amounts of N mineralised from pre-crop residues was positively correlated with wheat grain yield and wheat shoot N yield. A clear distinction between pre-crop treatments could be made on the first axis of the PCA. Firstly, according to the year of the experiment and, secondly, between spring and summer pre-crop treatments, especially in Experiment I (Fig. 5). The distinction between both experiments was mainly explained by contrasting rainfall conditions leading to different amounts of leached N-NO₃. Experiment I was characterised by higher rainfall, especially between August and November 2014, compared to Experiment II. The second axis of the PCA separated pre-crops species according to their effect on wheat grain yield and shoot N yield. Pre-crops for which the whole plant was considered as crop residues since no grains were harvested (chickpea in Experiment I and Narbonne vetch in Experiment I and II) led to the highest amounts of N mineralised from pre-crop residues (Nres_min, Fig. 5) and to the highest wheat grain yield (Wheat_Y) and shoot N yield (Wheat_N). For harvested grain pre-crops, the highest amounts of N mineralised from pre-crop residues was measured for faba bean, lentil and common vetch, and associated with high wheat grain yield and shoot N yield. Conversely, cereal pre-crops always had low N mineralisation rates of crop residues and were associated with the lowest wheat grain and shoot N yields. Figure 6A shows the positive linear relationship between wheat shoot N yield and cumulative amounts of N mineralised from pre-crop residues throughout the simulation period. However, for a given amount of N mineralised from pre-crop residues (for example 0 kg N ha-1), wheat shoot N yield still varied widely (between 60 and 77 kg N ha-1 in our example) according to the year of the experiment and between summer and spring pre-crops. Hence, climatic conditions affected the N losses by leaching, and may also have influenced the mineralisation soil organic N (as it depends on

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soil temperature and moisture). Moreover, at pre-crop harvest, the amounts of soil inorganic N left in the soil (N sparing) were variable (Fig 3C and D). The amount of N actually available during the wheat growth cycle was therefore calculated and was plotted versus wheat shoot N yield (Fig. 6B). A unique positive linear relationship was found between both variables for Experiments I and II and for spring and summer pre-crops. Generally, spring pre-crops and unharvested legume pre-crops (chickpea in Experiment I and Narbonne vetch in Experiment I and II) led to the highest amounts of N available for the following wheat and were associated with the highest wheat shoot N yield. Conversely, the rest of summer pre-crops led to the lowest amounts of N available for the following wheat and were associated with the lowest wheat shoot N yield. However, the relationship did not entirely explain the variability of wheat shoot N yield.

4. Discussion

4.1. Response of wheat performance to pre-crops and N fertilisation

Our study confirmed the positive effect of a wide variety of grain legumes on the yield of the following unfertilised cereal, with several legume species relatively poorly studied in European conditions. The increases of wheat grain yields after lentil and faba bean compared to barley (on average 27 %) were similar to the average yield response of cereals cultivated after legumes reported by Angus et al. (2015) and Cernay et al. (2017) (on average 33% and 29 % increases, respectively). Conversely, increases of wheat grain yields after summer legume pre-crops compared to sorghum (on average 61 %) were close to values reported by Preissel et al. (2015) (27 – 110% improvement) and Espinoza et al. (2012) (23 to 152% increases). Higher increases of wheat grain yields after summer legume pre-crops vs sorghum compared to spring legume pre-crop vs barley were mainly due to the choice of the reference cereal pre-crop (i.e. barley or sorghum). Indeed, wheat after sorghum always presented lower grain yields than wheat after barley.

However, benefits to wheat grain yields showed large variations among spring and summer pre-

crops, respectively, reflecting differences in legume pre-crops. For spring pre-crops, wheat grain

yields were the highest after faba bean and lentil as highlighted by Williams et al. (2014), Wright (1990), O'Donovan et al. (2014) and St Luce et al. (2016). Surprisingly, wheat grain yields after pea were slightly higher but not significantly different from those of wheat after barley, while several studies measured increases in cereal grain yields after pea in comparison to wheat, triticale and oat (Kirkegaard and Ryan, 2014; Miller et al., 2003; O'Donovan et al., 2014, St Luce et al., 2016, Strong et al., 1986). For summer legume pre crops, Narbonne vetch led to the highest increases in wheat grain yields. For a given year, only wheat after sorghum led to a higher shoot N concentration compared to wheat after summer pre-crops. These results can certainly be explained by higher N availability per unit of grain biomass, since wheat after sorghum always presented the lowest grain yields. Consequently, for spring legume pre-crops the increases in wheat shoot N yield (on average 25%) were within the same range as the increases in wheat grain yields (on average 27%). For summer legume pre-crops wheat shoot N yield increases (on average 51%) were slightly lower than the increases in grain yield (on average 66%). Although wheat was supplied with small amounts of N fertilisation (40 kg N ha⁻¹), grain yield, shoot N concentration and shoot N yield increased for all pre-crop treatments (legumes and cereals). It reflected the limitation of N for all the pre-crop treatments when wheat was unfertilised. While Preissel et al. (2015) and Cernay et al. (2017) showed that the positive effect of legume pre-crops decreased when N fertilisation was applied on the following cereal, our study showed that it was only the case for fenugreek and lentil compared to barley pre-crop. An average N fertilisation threshold of 153 kg N ha-1 was calculated in the meta-analysis of Cernay et al. (2017) above which legume precrops no longer had a positive effect on the subsequent cereal grain yield compared to cereal precrops. As the amount of N fertiliser applied in our experiment was far below this threshold, most legume pre-crops still induced an increase in wheat grain yields compared to cereal pre-crops, because N was still a limiting resource for wheat growth.

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4.2. N fluxes simulated by the STICS model as affected by pre-crops and climatic conditions

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The STICS model was used to quantify N fluxes difficult to measure in field conditions and to assess the effect of pre-crops and climatic conditions on these N fluxes. Maximum differences between the lowest and the highest amounts of N mineralised from pre-crop residues were 56 kg N ha⁻¹ in Experiment I and 68 kg N ha⁻¹ in Experiment II. The STICS module of N mineralisation is based on the C:N ratio of crop residues (Justes et al., 2009; Nicolardot et al., 2001). Hence, a single negative logarithmic relationship between the cumulative amount of N mineralised from pre-crop residues at the end of the simulation period and the C:N ratio of pre-crop residues was established for our two experiments $(y=-32.77ln(x)+113.4; R^2=0.94)$ (Fig. S1). The STICS model allowed to distinguish precrops species according the N mineralisation of their crop residues. Most pre-crop residues induced soil inorganic N immobilisation during their decomposition, but the highest amounts of N immobilised were measured for barley, sorghum soybean and lupin pre-crop residues, which had the highest C:N ratios. Conversely, the highest amounts of N mineralised from pre-crop residues were found for legume pre-crops for which the whole plant was considered as crop residues (chickpea in Experiment I and Narbonne vetch in Experiment II) and for which C:N ratios were the lowest. Part of the differences in soil inorganic-N and soil water content between spring and summer precrops may probably have been due to the growth cycle delay between spring and summer pre-crops. Once spring crops have reached physiological maturity, water and nitrogen (originating from rainfall and soil mineralisation, respectively) accumulate in the soil. Meanwhile, summer pre-crops continued to take up these resources to fulfil their requirements for growth. Hence, soil water and inorganic N contents were not only affected by pre-crop species and climatic conditions, but also by the growth period of spring and summer pre-crops and the positioning of their water and N requirements for growth. A part of soil inorganic-N was lost by leaching during the autumn-winter period because of a desynchronisation between N uptake by the subsequent wheat and soil inorganic-N dynamics. During this period, on the one hand, wheat development was very low, thus inducing low inorganic-N

and water uptake, whereas on the other hand, there were large amounts of soil inorganic-N and high water contents remaining in the soil after the pre-crop harvest combined with rainfall during autumn.

The amounts of N leached were certainly slightly under-estimated in Experiment I and over-

estimated in Experiment II since the amounts of soil inorganic N were on average under-estimated by 12 kg N ha⁻¹ and over-estimated by 14 kg N ha⁻¹ by the STICS model, respectively. Several studies (Beaudoin et al., 2008; Constantin et al., 2012; Coucheney et al., 2015) also highlighted the difficulty of accurately predicting soil inorganic N with the STICS model and Coucheney et al. (2015) found an RRMSE close to our own value (0.52 vs 0.46). Soil N dynamics result from multiple and simultaneous processes in interaction with the plant and water fluxes, often making prediction more difficult compared to soil water dynamics (De Willigen, 1991). In the case of a maximum under-estimation of 12 kg N ha⁻¹ in Experiment I and a maximum over-estimation of N leaching by +14 kg N ha⁻¹ in Experiment II, it supports the fact that in Experiment II N leaching was much lower than in

4.3. N fluxes induced by pre-crops partially explained wheat shoot N yield variability

Experiment I, especially because of lower rainfall between August and November.

One of our objectives was to better understand increases in wheat shoot N yield due to legume precrops N benefits. A positive relationship was established between the amount of N available for the subsequent crop and wheat shoot N yield. This positive relationship highlights a N limitation for wheat growth. Hence, in all situations, wheat shoot N yield could have been further increased by higher amounts of inorganic N availability if N losses had been reduced during the autumn-winter period. Our study points out the necessity of synchronisation between soil inorganic N availability and the N demand of the subsequent crop to maximise its growth and fulfil its N requirements.

Considering our relationship, the amount of N available for the subsequent wheat accounted for 49%

of wheat shoot N yield variability. Several N fluxes were not taken into account in our calculation and

could have varied among pre-crop species. Nitrogen mineralisation from pre-crop residues only took

into account aboveground crop residues while ignoring below-ground residues and N rhizodeposition. Nor were nitrogen losses by nitrous oxide emissions taken into account. Furthermore, improvements of wheat shoot N yield after legume pre-crops may not be solely attributable to N benefits (Peoples et al., 2009). Introducing legumes in a cereal based rotation can significantly reduce cereal soil borne diseases (Angus et al., 2015; Kirkegaard et al., 2008), help control weeds at the rotation scale and enhance microbial diversity and activity (McDaniel et al., 2014; Tiemann et al., 2015). These latter factors could vary between legumes due to properties inherent to species (e.g. competitiveness against weed) or because of the positioning of their growth cycle in a crop rotation (e.g. spring and summer species vs winter species). Yet, these factors were not measured in the present study but may have contributed to improved wheat grain and shoot N yields after legume pre-crops compared to cereal pre-crops. These factors might have been especially important to understand wheat yield limitation after sorghum especially in Experiment I, as N availability did not satisfactorily explain the very small grain yields of wheat following sorghum.

5. Conclusion

This study confirmed the relevance of reintroducing grain legumes in cereal- based cropping systems in Europe, due to the positive effect of a wide range of legume pre-crops on grain and shoot N yield of the subsequent wheat. Using the STIC model allowed to accurately predict N processes particularly difficult to measure directly in the field conditions. Differences between legume pre-crop species were highlighted according to: i) the amount of N mineralised from their residues, ii) the amount of inorganic N left in the soil at their harvest, and iii) the amount of N lost after legume harvest. These differences between pre-crop legumes need to be taken into account to increase the efficiency of subsequent crop use of N provided by legumes while limiting N losses. Higher amounts of inorganic N derived from the mineralisation of crop residues were found for species with low crop residues C:N ratios and were positively correlated to the amounts of N in the subsequent wheat. Legume growth cycle positioning in the year relative to soil inorganic N dynamics also modulated the amount of N left in the soil at harvest and the risks of N leaching when high precipitation occurred during the

autumn-winter period. A longer fallow period, especially after spring legumes with earlier harvest compared to summer legumes, increased the risk of N leaching due to inorganic N accumulation in the soil and the more frequent rainfall during this period. Adapted management practices, such as the establishment of cover-crop during the fallow period (Constantin et al., 2012; Tribouillois et al., 2015), especially for long fallow period, are crucial to reduce N losses by stocking soil nitrogen before the establishment of the subsequent crop. Finally, choosing a subsequent crop either with an early sowing date and a high capacity to retrieve soil inorganic N during the early stages (for example, oilseed rape) or with a deep rooting depth to access N leached in the deeper soil layers, might also be sensible choices for reducing N losses after legume harvest (Dejoux et al., 2003).

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Author's contribution

BN and A-SV designed the study. BN, and A-SV funded the experiment and provided funding for MG's work. BN, A-SV and MG collected the data. MG analyzed the data. All authors were involved in the interpretation of the results and contributed to writing the original version of the manuscript and improving the subsequent ones.

Conflict of interest

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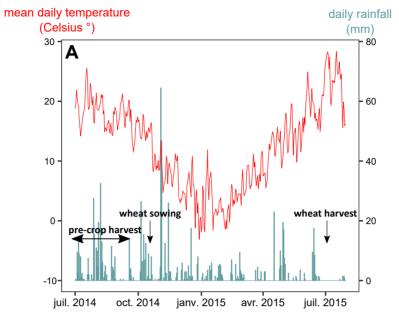
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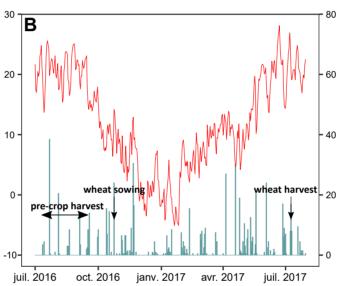
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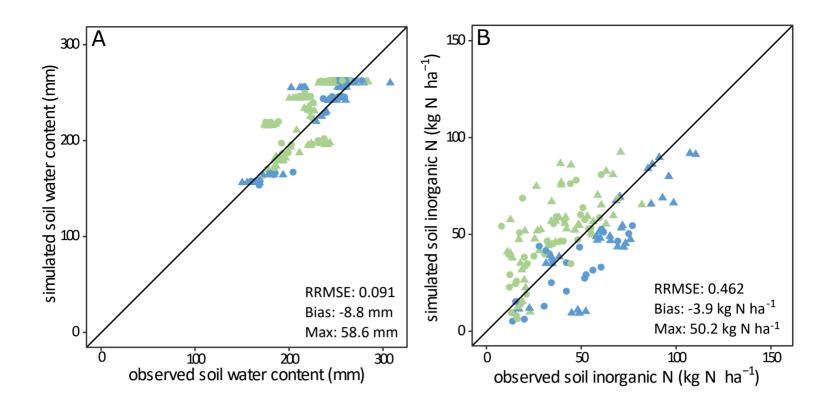
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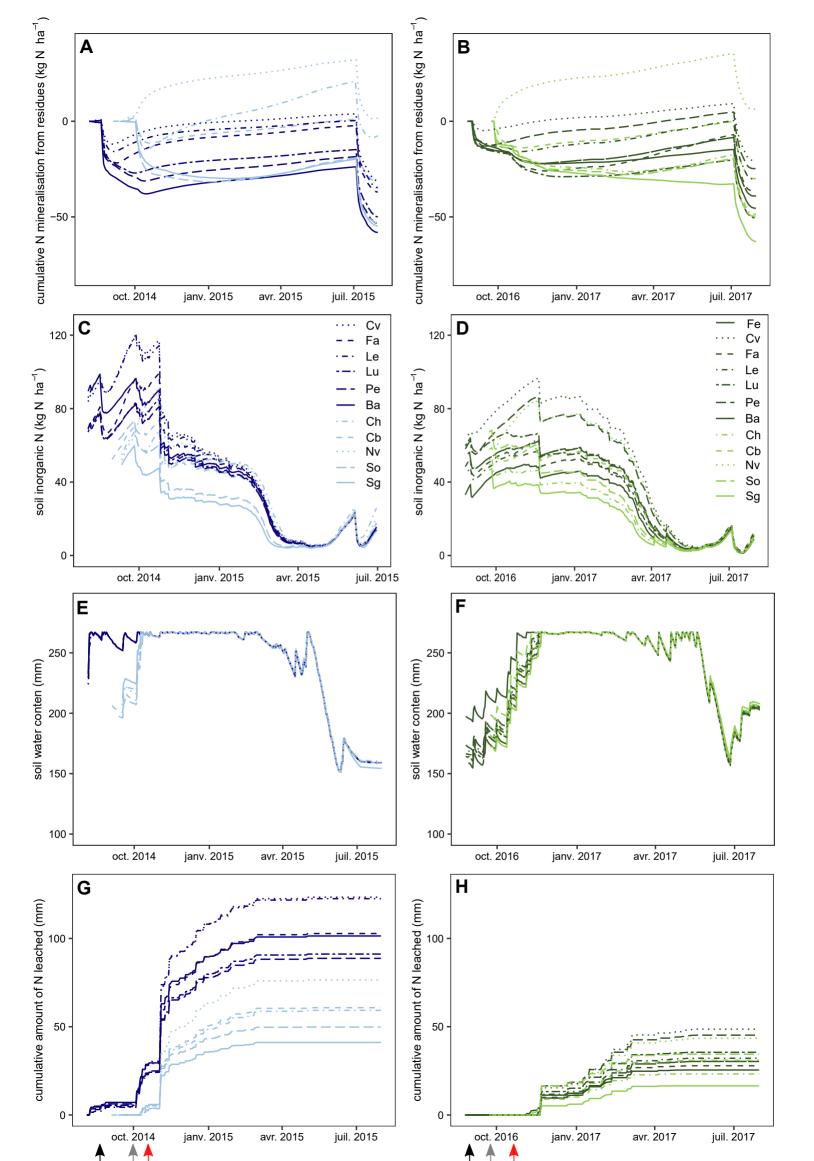
- Fig. 1 Daily rainfall, irrigation and mean temperature between pre-crop harvest and wheat harvest in Experiment I (A) and Experiment II (B). I: Irrigation; R: Daily rainfall; T: mean daily temperature. I, R and T were measured from a weather station located in the INRA experimental site of Bretenière. Horizontal dotted arrows indicate the pre-crop harvest period and vertical arrows indicate wheat sowing and harvest dates.
- Fig. 2 Simulated vs. observed soil water content (0 60cm) (A) and soil inorganic N (0 60 cm) (B) in Experiment I (blue) and Experiment II (green) between pre-crop harvest and wheat harvest. Wheat was cultivated after spring pre-crops (▲) and summer pre-crops (●). The solid line corresponds to the 1:1 line.
- 692 Fig. 3 Cumulative amounts of N mineralised from pre-crop residues (A, B), soil inorganic N (C, D), soil 693 water content (E, F) and cumulative amounts of N-NO₃- leached (G, H) simulated with STICS for 694 Experiment I (A, C, E, G) and Experiment II (B, D, F, H). Simulations were performed for the period 695 between pre-crop harvest and winter wheat harvest. Dark blue and dark green curves are the 696 simulations performed for the spring pre-crop treatments and light blue and light green curves are 697 the simulations performed for summer pre-crops treatments. Black and grey arrows indicate the 698 incorporation dates of spring and summer pre-crop residues, respectively. The red arrows indicate 699 the sowing dates of wheat. Pre-crop ID are specified in Table 2.
 - Fig. 4 STICS model simulations obtained for the cumulative N-NO₃⁻ leached between pre-crop harvest and wheat sowing (**A**, **B**) and between wheat sowing and the end of winter (1st March) (**C**, **D**) in Experiment I (blue bars) and II (green bars). Simulations were performed for spring pre-crops (**A**, **C**) and summer pre-crops (**B**, **D**). Legume pre-crops: solid bars; cereal pre-crops: hatch bars. Bars with a heavy dark border indicate legumes for which the whole plant was considered as crop residues since there was no grain production.
 - **Fig. 5** Biplot PCA showing the distribution of pre-crop species according to environmental conditions, proprieties of crop residues and wheat performance. Amounts of N in pre-crop shoot residues (Res_N); C:N ratio of pre-crop residues (Res_C:N); soil inorganic N at the start of simulation (SoilN_i); water content in the soil profile at the start of simulation (SoilW_i); cumulative rainfall during the simulation period (Rain); cumulative amounts of N mineralised from pre-crop residues during the simulation period (Nres_min); cumulative amounts of N-NO₃- leached during the simulation period

- 712 (Nleached); wheat grain yield (Wheat_Y); wheat shoot N yield (Wheat_N); wheat shoot N 713 concentration (Wheat_%N). Pre-crop ID are specified in Table 2.
- Fig. 6 Relationships between wheat shoot N yield and the amount of N mineralised from pre-crop residues estimated by the STICS model (A) and the amounts of N available for wheat (soil inorganic N at pre-crop harvest + N mineralisation from pre-crop residues + N mineralisation from soil organic matter amount of N leached) (B). The significance of the linear regression is shown as ** p < 0.01 and *** p < 0.001. Pre-crop ID are specified in Table 2.

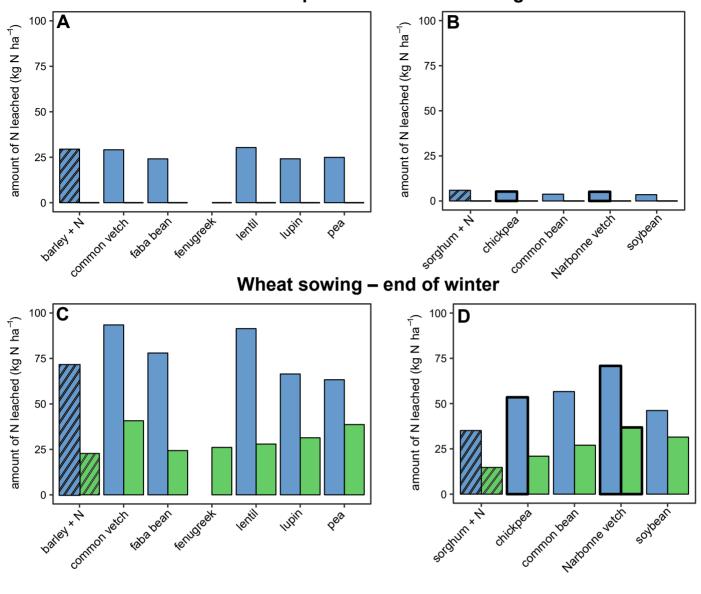


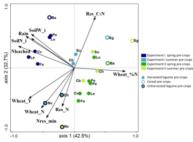


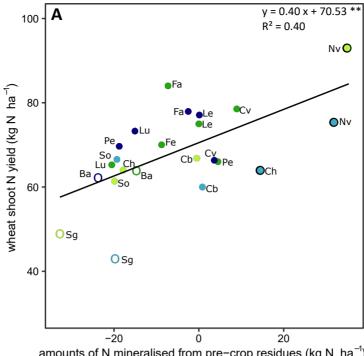




Pre-crop harvest - wheat sowing







amounts of N mineralised from pre-crop residues (kg N ha⁻¹)

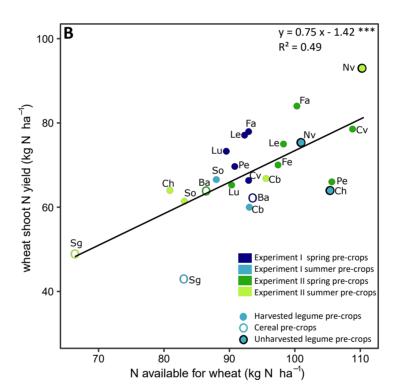


Table 1 Soil characteristics in Experiments I and II

	Experiment I (2014-2015)	Experiment II (2016-2017)
Clay (g kg ⁻¹)	425	512
Silt (g kg ⁻¹)	515	428
Sand (g kg ⁻¹)	60	60
Organic C (g kg ⁻¹)	19.3	19.6
Organic N (g kg ⁻¹)	1.69	1.81
C:N ratio	11.4	10.8
рН	7.31	7.8
CaCO ₃ (g kg ⁻¹)	< 1	4

 Table 2 Pre-crops and wheat characteristics and management in Experiments I and II.

Pre-crop treatments	ID	Pre-crop sowing period	Sowing density (seeds m ⁻²)	Pre-crop harvest date	Residue C:N ratio	Incorporation of pre-crop residues	Wheat sowing date	End of winter	Wheat harvest date
						in the soil			
				Experiment I	(2014-2015)				
Common vetch	Cv		100	11 July 2014	28.6				
Faba bean	Fa		45	25 July 2014	34.2				
Lentil	Le	18 March 2014	250	11 July 2014	31.9	21 August 2014	20 October 2014	1 March 2015	6 July 2015
Lupin	Lu	(spring)	70	1 August 2014	69.7				6 July 2015
Pea	Pe		80	1 July 2014	64.6				
Barley + N	Ва		350	15 July 2014	73.2				
Chickpea	Ch		70	16 September 2014	23.4				
Common bean	Cb	E NA 2044	30	4 September 2014	28.8		20 October 2014	1 March 2015	6 July 2015
Narbonne vetch	Νv	5 May 2014	35	5 September 2014	13.6	3 October 2014			
Soybean	So	(summer)	70	5 September 2014	47.8				
Sorghum + N	Sg		30	5 September 2014	50.0				
				Experiment II	(2016-2017)				
Common vetch	Cv	15 March 2016	100	27 July 2016	22.3				
Faba bean	Fa	1 March 2016	45	2 August 2016	34.2				
Fenugreek	Fe		180	2 August 2016	36.9	2 September	24 October		
Lentil	Le	15 March 2016	250	27 July 2016	29.5	2 September 2016		1 March 2017	6 July 2017
Lupin	Lu		70	12 August 2016	65.6	2010	2016		
Pea	Pe	(spring)	80	13 July 2015	26.7				
Barley + N	Ba		350	20 July 2016	56.6				
Chickpea	Ch		55	12 September 2016	45.0				
Common bean	Cb	17 14 2016	30	12 September 2016	29.5	07.0	24 October 2016	1 March 2017	6 July 2017
Narbonne vetch	Νv	17 May 2016	35	1 September 2016	12.6	27 September			
Soybean	So	(summer)	70	12 September 2016	55.8	2016			
Sorghum + N	Sg		30	14 September 2016	82.7				

Table 3 Grain yields, shoot N concentration and shoot N yield of unfertilised (0N) wheat as affected by spring and summer pre-crop treatments and years (2015 and 2017). When no pre-crop by year interactions were detected, pre-crop values were averaged over year. Values are mean (n=4 or 8) \pm sd (standard deviation). Lowercase letters refer to contrasts between pre-crops. Mean value sharing the same letter ('a' and / or 'b') are not different according to Tukey's test (ns: non-significant, * p < 0.05, ** p < 0.01, *** p < 0.001).

	SPRING	PRE-CROPS		SUMMER PRE-CROPS						
	Wheat yield 0% humidity (t ha ⁻¹)	Wheat shoot N concentration (%N)	Wheat shoot N yield (kg N ha ⁻¹)		Wheat yield 0% humidity (t ha ⁻¹)	Wheat shoot N concentration (%N)	Wheat shoot N yield (kg N ha ⁻¹)			
	2015 and 2017 pooled	2015 and 2017 pooled	2015 and 2017 pooled		2015 and 2017 pooled	2015 and 2017 pooled	2015	2017		
Pea	4.0 (0.5) bc	0.79 (0.06)	68 (6) bc	Common bean	3.7 (0.4) b	0.75 (0.02) b	60 (6) b	67 (5) b		
Lupin	4.2 (0.4) abc	0.76 (0.05)	69 (6) abc	Chickpea	3.8 (0.2) b	0.77 (0.05) b	64 (4) ab	64 (2) b		
Common vetch	4.2 (0.4) abc	0.79 (0.06)	73 (9) abc	Soybean	3.7 (0.8) b	0.78 (0.02) b	67 (13) ab	62 (10) cb		
Lentil	4.5 (0.4) ab	0.78 (0.05)	76 (5) ab	Narbonne vetch	4.9 (0.3) a	0.76 (0.05) b	76 (2) a	93 (3) a		
Faba bean	4.7 (0.9) a	0.76 (0.02)	81 (16) a	Sorghum +N	2.5 (0.3) c	0.87 (0.03) a	43 (3) c	49 (5) c		
Barley +N	3.6 (0.4) c	0.79 (0.06)	63 (4) c							
year	**	***	ns	year	ns	**		*		
species	***	ns	**	species	***	***	k	***		
year*species	ns	ns	ns	year*species	ns	ns		*		

Table 4 Grain yields, shoot N concentration and shoot N yield of wheat in 2017 as affected by spring and summer pre-crop treatments and N fertilisation rates (0N and 40N). When no pre-crop by N rate interactions were detected, pre-crop values were averaged over N rate. Values are mean (n=4 or 8) \pm sd (standard deviation). Lowercase letters refer to contrasts between pre-crops. Mean value sharing the same letter ('a' and / or 'b') are not different according to Tukey's test (ns: non-significant, * p < 0.05, ** p < 0.01, *** p < 0.001).

		SPRING PRI	E-CROPS			SUMMER PRE-CROPS					
	Wheat yield 0% humidity (t ha ⁻¹)		Wheat shoot N Wheat shoot N yield concentration (%N) (kg N ha ⁻¹)			Wheat yield 0% humidity	Wheat shoot N concentration (%N)		Wheat shoot N yield		
	ON	40N	0N and 40N pooled	ON (Kg I	40N		(t ha ⁻¹) ON and 40N pooled	ON	40N	(kg N ha ⁻¹) ON and 40N pooled	
Fenugreek	4.0 (0.5) ab	4.7 (0.1) c	0.84 (0.08)	70 (8) bcd	86 (4) c	Common bean	4.9 (1.0) b	0.75 (0.03) b	0.82 (0.01)	84 (19) a	
Pea	3.6 (0.4) bc	5.3 (0.3) abc	0.92 (0.11)	66 (8) cd	105 (6) ab	Chickpea	4.5 (0.8) b	0.79 (0.07) b	0.84 (0.04)	78 (16) b	
Lupin	3.8 (0.1) bc	5.1 (0.3) bc	0.85 (0.08)	65 (3) d	99 (5) abc	Soybean	4.4 (1.0) b	0.79 (0.01) b	0.84 (0.05)	77 (17) b	
Common vetch	4.2 (0.1) ab	5.7 (0.2) ab	0.88 (0.05)	79 (5) ab	109 (8) ab	Narbonne vetch	5.9 (0.7) a	0.80 (0.04) ab	0.90 (0.07)	108 (17) b	
Lentil	4.2 (0.2) ab	4.9 (0.6) c	0.86 (0.08)	75 (3) abc	93 (15) bc	Sorghum +N	3.4 (0.9) c	0.89 (0.04) a	0.82 (0.01)	60 (13) c	
Faba bean	4.7 (0.5) a	5.8 (0.3) a	0.82 (0.08)	84 (9) a	114 (9) a						
Barley +N	3.3 (0.1) c	4.9 (0.1) c	0.84 (0.04)	64 (3) d	88 (5) c						
N rate	*	**	***	*	**	N rate	***	**		***	
species	*	:**	**	:	**	species	***	*		***	
N rate*species		**	ns		*	N rate*species	ns	*		ns	