

# Comparison of molecularly imprinted polymers (MIP) and sol—gel molecularly imprinted silica (MIS) for fungicide in a hydro alcoholic solution

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- 1 Comparison of molecularly imprinted polymers (MIP) and sol-gel
- 2 molecularly imprinted silica (MIS) for fungicide in a hydro alcoholic
- 3 **solution.**

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#### 13 **Abstract**

- 14 The aim of this work was (i) to design the best molecularly imprinted silica (MIS) specific for
- iprodione, a fungicide polluting water, fruits, wine and (ii) to compare the performance of the
- optimal MIS with its analogue molecularly imprinted polymer (MIP), with the objective to
- 17 obtained the highest imprinting factor possible. Iprodione was widely used in viticulture. It was
- 18 recently banned by the European community because of its carcinogenic potential and its
- 19 presence in or on foods.
- 20 The binding capacity performance of these polymers was investigated using batch binding
- studies in hydro alcoholic solutions (10 and 50 %). Langmuir, Freundlich and Temkin models
- 22 have been used to fit adsorption isotherms. The optimal MIS was obtained using APTMS as
- functional monomer, low solvent volume and high cross linker quantity during its synthesis.

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- 24 Increasing the quantity of crosslinker and decreasing the solvent volume during the MIS
- 25 synthesis enhanced the imprinting factor and the binding capacity.
- 26 The optimal MIS had a higher imprinting factor but a lower binding capacity than the optimal
- 27 MIP specific to iprodione. The imprinting factor of the optimal MIS was 135 times higher than
- 28 the optimal MIP. Unlike MIP, MIS kept its molecular recognition nature even under extremely
- 29 polar media such as the aqueous sample.

#### Keywords

Molecularly imprinted silica, molecularly imprinted polymer, fungicide, adsorption isotherm

#### 1. Introduction

The worldwide increase in the usage of pesticides during the last decades has resulted in the accumulation of their residues in the environment. These residues can cause serious detrimental effects on human health and on the ecosystem <sup>1</sup>. Therefore, the maximum residue limits (MRL) were established by the Codex Alimentarius Commission and the European Union to regulate the use of pesticides. Thus, the monitoring of these compounds is extremely important to ensure that only permitted levels of pesticide are used. Several techniques have been developed for pesticide detection: conventional analytical methods (gas chromatography, high performance liquid chromatography, immunoassay, capillary electrophoresis) and advanced detection methods such as biosensor (electrochemical, optical, piezoelectric, *etc...*) <sup>1</sup>.

Although very effective, these techniques are expensive, time-consuming and difficult to miniaturize for detection in the field. In this context, molecularly imprinted polymers (MIP) specific to pesticides can overcome some of these difficulties, both for its extractions and detections: significant reduction in extraction time and improvement in the concentration of the target molecule for subsequent analysis. That is why since 2000, molecular imprinting science

and its applications of molecularly imprinted polymers have been attracting the interest of the scientific community, hence the broad increase of literature in the field. Over 1237 references within the field of molecular imprinting science and technology for the period 1931 – 2018 have been collected. However, numerous developments still need to be made, particularly in terms of the synthesis method of molecularly imprinted silica.

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For this project, we focused on the iprodione (other name glycophen, PubChem CID 37517). Iprodione is a dicarboximide fungicide widely used in viticulture in most agricultural countries, but also in other crops activities like fruits production, green lawns etc.. <sup>2-4</sup>. It was banned recently by the European Community because of its carcinogenic and endocrine disrupting characters <sup>5</sup>. Though this interdiction, iprodione must be detected because of possible residues in soil. It is a very stable molecule and its degradation in the soil takes a long time <sup>6</sup>. In fact, in a soil treated for the first time, the time to 50% loss of iprodione was greater than 35 days. Iprodione has also to be easily detected for the monitoring of possible non-authorized uses like the elimination of stocks etc... Various molecular imprinted polymers (MIP) have been recently developed and applied for the selective determination of residual pesticides <sup>7</sup>. A brief overview of this research area shows that the majority of the studied imprinted polymers are acrylate based molecularly imprinted polymers (MIP) (Table 1). The most used functional monomer is the methacrylic acid (MAA). It is commonly chosen by default as monomer as soon as the target molecules are neutral or basic. In fact, MAA has the capacity of being an H donor and acceptor with strong abilities to create dipole-dipole interactions and can develop hydrogen and ionic interactions with basic molecules 8. MIP synthesis is relatively cheap and easy. However, a severe limitation of this type of imprinting is the requirement for an organic solvent in which all compounds are soluble.

Pesticides are generally poorly soluble in water. This chemical property is favorable to MIP synthesis protocols that use organic solvents. Initially, the selection of low water soluble molecules was done to reduce lixiviation in groundwater and water pollution <sup>9</sup>. For nebulization in the fields, pesticides are dispersed in water using a surfactant <sup>10</sup>. These poorly soluble in water molecules should nevertheless not be too liposoluble in order to avoid bioaccumulation. Like per example pentachlorophenolas who is known to contaminate underground water <sup>11</sup> due to its high solubility in water (20 mg.L<sup>-1</sup>) when, on the other hand, chlordane and DDT are very hydrophobic (water solubility 0.056 and 0.024 mg.L<sup>-1</sup> respectively), stable and toxic, and therefore, bioaccumulate. Iprodione is relatively hydrophobic and slightly soluble in water (13.9 mg·L<sup>-1</sup>). It is more soluble in organic solvents: in ethanol (25 g·L<sup>-1</sup>), which explains the high risk of contamination of wine, in acetone (300 g·L<sup>-1</sup>), in dimethylformaide (500 g·L<sup>-1</sup>), or in dichloromethane (500 g·L<sup>-1</sup>). A convenient alternative to MIP is sol – gel molecularly imprinted silicas (MIS), even if pesticides are not very water-soluble <sup>12</sup>. The use of a co-solvent (water miscible solvent such as ethanol or acetone) can favor the solubilization of pesticide without perturbating the polymerization of the silane. In fact, the synthesis of MIS is relatively easy: it can be done at room temperature and atmospheric pressure. Their advantage remains in their synthesis in a water / ethanol mixture, which respects one of the principles of green chemistry by using safe solvents contrary to those used in MIP synthesis. Moreover, MIS are more chemically stable and specific towards the target species than acrylate based MIP 13, also, they allow faster diffusion of analytes <sup>14</sup>. The interest in these materials is growing and their applications are broad: phenobarbital <sup>13</sup>, caffeine <sup>14</sup>, nitroarmatic explosives <sup>15</sup>, pesticides in almond oil <sup>16</sup>, nitro musk compounds in water <sup>17</sup>. However, the studies about MIS using fungicide as template are very scare (Table 1).

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All these studies proved the efficiency of MIP and MIS but, to the best of our knowledge, this study is the first that compares the molecular imprinting of iprodione between these two types of polymers. The aim of this work was to design the best MIS specific for ipriodione fungicide and to compare its performance with its MIP analogue. The binging capacity performance of these polymers was investigated in a hydroalcoholic solution towards iprodione.

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#### 2. Materials and methods

#### 2.1. Chemicals

105 Iprodione (97%, CAS number 36734-19-7, Hazard class carcinogenicity category 2 (CMR 2), H351, H400, H410), (3-Aminopropyl-)trimethoxysilane (APTMS 97%, CAS number 13822-106 107 56-5, Hazard class H314, H315, H318, H319), 1-[3-(Trimethoxysilyl)propyl]urea (urea, 97%, 108 **CAS** number 23843-64-3, Hazard class H315. H318, H335), N-[3-109 (Trimethoxysilyl)propyl]aniline (aniline, 97%, CAS number 3068-76-6, Hazard class H302, 110 H314, H315), tetraethoxysilane (TEOS, ≥ 99%, CAS number 78-10-4, Hazard class H226, 111 H319, H332, H335), ammonium hydroxide (NH<sub>4</sub>OH 28 – 30%, CAS Number 1336-21-6, 112 Hazard class H314, H400), ethanol (≥ 99.8%, CAS Number: 64-17-5, Hazard class H225), 113 allylamine (98%, CAS 107-11-9, Hazard class H225, H301, H311, H331, H411), ethylene 114 glycol dimethacrylate (EGDMA,  $\geq$  99%, CAS number 97-90-5, Hazard class H317, H335), 2,2-115 dimethoxy-2-phenylacetophenone (DMPAP, > 99%, CAS number 24650-42-8, Hazard class 116 H302, H373, H400, H410, H412), toluene (≥ 99.8%, CAS Number: 108-88-3, Hazard class 117 reproductive toxicity category 2 (CMR 2), H225, H304, H315, H336, H361d, H373) and 118 acetonitrile (≥ 99.9%, CAS Number: 75-05-8, Hazard class H225, H302, H312, H319, H332) 119 were purchased from Sigma Aldrich, France. Water used in all experiments was deionised and 120 obtained from an Elga Ionic system PURELAB Option

# 2.2. Synthesis of the sol-gel molecularly imprinted silica (MIS) and non-imprinted solgel polymers silica (NIS)

The synthesis protocol used for this study and the different chemical reactions that might be involved in the MIS synthesis and the interactions between the MIS and iprodione have been presented previously  $^{12}$ . MIS were prepared at  $40^{\circ}$ C in a thermostatic water bath under magnetic stirring. The template molecule, iprodione, was first solubilized in ethanol. Then, water was added, followed by the functional monomer and the crosslinker TEOS (tetraethoxysilane). Finally, aqueous solution of ammonium hydroxide (NH<sub>4</sub>OH, 100 mmol) was introduced. The reaction mixture was left under stirring for 20 hours. Then the mixture was sonicated for 10 minutes. The polymers were separated from the liquid phase by centrifugation at 10~000~g for 10~minutes at room temperature. In order to eliminate iprodione and free the specific cavities, the polymers were washed several times with pure ethanol until iprodione was no longer detectable by reversed phase high performance liquid chromatography (HPLC). After washing, the polymers were dried for 6~h at 60~°C. In parallel, NIS (Non Imprinted Silica) was synthesized under the same synthesis conditions as those of MIS, but without using the template molecule. NIS served as a control polymer.

Several factors, including the nature of functional monomer, molar ratios of crosslinker / functional monomer / solvent were studied to optimize the synthesis of MIS. Table 2 summarizes the different MIS synthesized depending on the experimental conditions and the structure of functional monomers tested for the synthesis of molecularly imprinted silica (MIS) and for the molecularly imprinted polymers (MIP) are presented in Figure 1.

# 2.3. Synthesis of the molecularly imprinted polymers (MIP) and non-imprinted polymers (NIP)

The synthesis of MIP (non-covalent synthesis method) was done as previously described <sup>8</sup>. MIP were prepared with 1 mmol of allylamine as functional monomer, 4 mmol of crosslinker (EGDMA) and 0.1 mmol of template (iprodione) and the polymerization was carried out in 1 mL of toluene using 0.05 mmol of DMPAP initiator. In parallel, non-imprinted polymers (NIP) were prepared with the same protocol but without the template. NIP served as a control polymer.

#### 2.4. Binding properties of imprinted polymer (MIS) and non-imprinted polymer (NIS)

#### towards iprodione

The binding capacity was estimated by batch adsorption. 10 mg of imprinted polymer (MIS 1, MIS 4, MIS 5, MIS 6, MIS 7, MIS 8) or non-imprinted polymer (NIS 1, NIS 4, NIS 5, NIS 6, NIS 7, NIS 8) were suspended in 20 mL of iprodione solution at  $10^{-5}$  M (10 %, v/v, ethanol/water). The mixture was stirred for two hours at 25 °C. After, the supernatant was filtered using a 0.22  $\mu$ m filter and analysed by HPLC for the determination of iprodione. For each analysis, three replicates were done.

#### 2.5. Kinetic adsorption study of iprodione to MIS

The kinetic profile of iprodione binding to MIS was performed in the same experimental conditions than those previously described, except for the concentration of iprodione solution. An iprodione solution at  $5.10^{-4}$  M (10 %, v/v, ethanol/water) was used. At different times of stirring at 25 °C, the supernatant was filtered using a 0.22  $\mu$ m filter and analysed by HPLC for the determination of iprodione. Three replicates were done by sample.

#### 2.6. Isotherm study

10 mg of imprinted polymer (MIS 4, MIS 5', MIS 8 or MIP) or non-imprinted polymer (NIS 4, NIS 5', NIS 8 or NIP) were suspended in 20 mL iprodione solutions at different concentration  $10^{-5}$ ,  $5.10^{-4}$ ,  $10^{-4}$ ,  $5.10^{-4}$ ,  $10^{-3}$  M with ethanol/water mixture (50 %, v/v) as solvent. After one hour of stirring at 25 °C, the supernatant was filtered using a 0.22 µm filter and analysed by HPLC for the determination of iprodione. The results presented are the means of three replicates. The sorption isotherms obtained were tested with the Langmuir, Freundlich, and Temkin isotherm equations. The model equations are shown in table \$1. The Langmuir isotherm model assume that the adsorption of molecules occurs at specific homogeneous sites of the adsorbent and, once a molecule occupies a site, no additional adsorption can occur there (monolayer coverage). The Freundlich isotherm model assumes the molecules undergoes adsorption onto the heterogeneous surface of adsorbent. This model describe a multilayer adsorption with interaction between adsorbed molecules. The Temkin isotherm model assumes that the adsorption heat of all molecules decreases linearly with the increase in coverage of the adsorbent surface, and that adsorption is characterized by a uniform distribution of binding energies, up to a maximum binding energy<sup>18,19</sup>.

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#### 2.7. High-performance liquid chromatography

For HPLC iprodione analysis, a Shimadzu HPLC (LC-20AT) pump equipped with a UV-Visible detector (SPD-20A), a Restek column (Pinacle II, C18 5  $\mu$ m, 150 x 4.6 mm) and a precolumn (C18, 5  $\mu$ m, 10 x 4 mm) were used. Separations were conducted at room temperature. Isocratic acetonitrile / water (60/40, v/v) was used as mobile phase and the flow rate was 1mL/min. Iprodione detection was done at 220 nm. A calibration curve was constructed over the range 5.10<sup>-4</sup> and 10<sup>-8</sup> M of iprodione (r<sup>2</sup> = 0.9993). The amount of iprodione retained by the

polymer was calculated by the difference between the initial concentration of iprodione and the free concentration of iprodione in the supernatant.

#### 2.8. Scanning electron microscope (SEM)

The morphological characterization of the samples was performed by a scanning electron microscope. The samples were suspended in ethanol. Then a drop was placed on a silicon grid and examined using a Hitachi VPSU1510 scanning electron microscope. More than four views were recorded by sample. Representative images were presented with the accelerating voltage and the magnification used.

#### 2.9. Statistical analysis

The Statistica V8 software was used for Anova. Significance was established at p < 0.05. Fisher t-test was used to determine significant differences between values.

#### 3. Results and discussion

#### 3.1. Synthesis of sol – gel molecularly imprinted silica (MIS) optimization

An optimal MIS must satisfy the following two criteria: present a strong absorption of iprodione and have a better specificity compared to NIS. Several factors, including the nature of functional monomer, molar ratios of crosslinker / functional monomer / solvent were studied to optimize the synthesis of MIS. Table 2 summarizes the different MIS synthesized depending on the experimental conditions.

#### **3.1.1.** Effect of functional monomer nature

The choice of the functional monomer depends largely on the imprinted molecule. Indeed, it is interesting to match the functionality of the functional monomer to that of the imprinted

- molecule (template), in order to maximize the interactions between them and to create highly specific cavities designed for the template molecule. The monomer must be stable and soluble in the chosen solvent. In our study, three different monomers were chosen considering their ability to interact with iprodrione (Figure 1):
- (3-Aminopropyl-)trimethoxysilane (labelled APTMS). This momoner has a hydrogen bond acceptor group.
- 1-[3-(Trimethoxysilyl)propyl]urea (labelled urea) which has a donor group and a hydrogen bond acceptor group.
- N-[3-(Trimethoxysilyl)propyl]aniline (labelled aniline) which has a hydrogen bond acceptor
   group and a benzene ring which can establish π-π interactions.

In a typical MIS synthesis, according to the literature  $^{14,15}$ , the following molar ratio (1 / 4/20) for (template / functional monomer / crosslinker) were commonly used. Following this literature review, the three selected functional monomers (APTMS, urea, aniline) have been tested depending on this molar ratio (1 / 4/20) and the following MIS labelled MIS 1, MIS 6, MIS 7 were obtained. The binding properties results obtained are presented in Figure 2. MIS 6 (synthesized with aniline residue) and MIS 1 (synthetized with APTMS) showed the same quantity of adsorbed iprodione (around 1 mg / g of polymer). For these two functional monomers, the same type of functional monomer - iprodione interactions, hydrogen bonds, were involved. On the other side, MIS 7 (synthesized with aniline residue) had an average adsorption of 5.86 mg / g of polymer, five times larger than these counterparts. The binding capacity of the polymer is a critical factor. In the case of aniline, the types of functional monomer - iprodione interactions involved were hydrogen bonds and  $\pi$ - $\pi$  interactions. This higher retention can be explained by the  $\pi$ - $\pi$  interactions involved between the phenyl ring of aniline residue and of iprodione. This expected  $\pi$ - $\pi$  interactions were more favorable than

hydrogen bonding (APTMS - iprodione and urea moieties - iprodione) for the formation of the pre-polymerization complex and thus for molecular printing.

No NIS were obtained with urea function (NIS 6) and aniline function (NIS 7) used as functional monomers with the molar ratios used. To confirm these results, the experiment was done in duplicate. We also noticed that the polymerization between the samples was different and consequently the MIS physical forms were different. Our experimental conditions gave very small granules for MIS 1 while soft polymer monoliths were obtained for MIS 6 and MIS 7. Consequently, before analysis, MIS 6 and MIS 7 were crushed and ground into a powder with a mortar and pestle.

As the involved interactions were different and the obtained binding capacity interesting, the functional monomers aniline (MIS 7) and APTMS (MIS 1) were kept for the further studies. Therefore, we can suppose that the change of the (template / functional monomer / crossliker) molar ratio and solvent volume could be a solution to obtain NIS 7.

#### 3.1.2. Crosslinker quantity and volume solvent effect

The volume of the solvent is also a key element in the MIS synthesis. To study it, two different volumes of solvents were used for the synthesis. Figure 3 shows that the decrease of solvent volume during the synthesis step with APTMS as functional monomer induced an increase of the quantity of adsorbed iprodione (MIS 1 and MIS 4). This observation could be explained by the dilution effect. The higher the solvent volume, the lower the reactants meeting probability, the fewer the cavities specific to iprodione. Therefore, there is a great interest in decreasing the solvent volume during the MIS synthesis.

Another important reagent for the synthesis of molecularly imprinted silica is the amount of crosslinker. The monomer and the crosslinker react mainly in aqueous solution to form silanol (Si-OH) groups through hydrolysis, then siloxane bonds (Si-O-Si) are formed by condensation reaction with silanol groups around the template molecule favouring the creation of a strong pre-complex template/monomer and an important reticulation of the sorbent to create more-well designed cavities. In other words, the crosslinker controls the morphology of the polymer: gel, micro or macroporous. It also provides a mechanical stability to silicate with the template molecule (rigidity), as well as a stability of the sites of interactions between the functional monomer and the template molecule. Liu et al. <sup>20</sup> showed that an excess of crosslinker can change the physicochemical interactions between monomers and template molecules. In fact, if the newtwork is too flexible, the template may not be retained. On the other hand, if the network is too reticulated, the template diffusion may be reduced.

On this basis, MIS 5 was synthesized with a small volume of solvent but with an increase of the crosslinker amount. MIS 5 had the highest iprodione binding capacity, three times more

On the contrary, with aniline as functional monomer, the trend was completely different (Figure 3). The increase of the crosslinker quantity and the decrease of the solvent volume implied a decrease in the amount of adsorbed iprodione (MIS 7 and MIS 8). It should be noted that the amount of crosslinker was greater for MIS 8 (1 / 4 / 59) than MIS 5 (1 / 4 / 20). However, contrary to the previous results, a NIS was obtained (NIS 8). MIS 8 and NIS 8 were this time in powder form.

than the other counterparts MIS 1 and MIS 4 (around 6 mg/g polymer).

To evaluate the specific binding property of MIS and NIS towards iprodione and to highlight the non-specific interactions that result from the iprodione adsorption, the imprinting factor (IF) has been calculated using the following equation <sup>21</sup>:

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$$IF = \frac{K(MIS)}{K(NIS)} = \frac{B(MIS) \times F(NIS)}{F(MIS) \times B(NIS)}$$

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- F: the free concentration of iprodione (mol/L) in the supernatant
- B: the amount of iprodione bound (adsorbed) by the polymer (mg of iprodione/g of polymer) calculated by difference between the initial concentration of iprodione (mol/L) and F.

The higher the imprinting factor, the higher the specific interactions. Therefore, the imprinted

300 polymers exhibit a good performance with the template molecule. The imprinting factors were 301 approximately around 1 for MIS1, MIS 4 and MIS 5 (Table 3) meaning the absence of specific 302 cavities designed for iprodione with no significant differences (p = 0.66). Table 3 and Figure 3 303 show that MIS 5 had the best imprinting factor and the best amount of adsorbed iprodione. This 304 observation could be explained by the increase of the specific interactions between iprodione 305 and the polymer with this molar ratio of reagent used. Therefore, with APTMS as functional 306 monomer, there is a great interest to increase the crosslinker quantity and to reduce the solvent 307 volume during the MIS synthesis. In fact, an increase of crosslinker quantity and a reduction of 308 solvent volume would normally increase the reticulation and reduce the porosity of the polymer. 309 The imprinting factor of MIS 8 was not significantly different than those obtained with APTMS 310 as a functional monomer (Table 3). MIS 8 was less selective than MIS 5 but better than MIS 4 311 (Figure 3). To try to understand this difference, the morphology of each MIS has been studied 312 by microscopy. SEM images (Figure 4) show the morphology of MIS with different 313 magnifications. MIS 5 and MIS 1 seem to be aggregates. A very fine homogeneous population 314 of about 400 nm in diameter characterized them. At the opposite, MIS 7 and MIS 8 were 315 characterized by a heterogeneous population with irregular shapes and greatly variable 316 dimensions ranging respectively from 99 µm to 952 µm for MIS 7 (with a majority of 200 µm)

and from 578 nm to 861 µm for MIS 8. The heterogeneity of MIS 7 populations explained by the gridding done. Due to the heterogeneity of the MIS 8 population and its larger size, this specific surface area may be smaller than MIS 5 and MIS 1 and could explain the difference of binding capacity observed previously by comparison.

Whatever the studied MIS, no differences were observed on the amount of adsorbed iprodione between MIS and NIS (Figure 3). In order to be able to observe a clear difference between MIS and NIS in the amount of adsorption iprodione and to correctly perform the isotherm studies of each polymer, the concentration of iprodione solution used have to be increased. The problem encountered is the low solubility of iprodione using 10% ethanol / water, (v/v) as solvent. In this condition, the solubility limit of iprodione is 2.10<sup>-4</sup> M. The iprodione solubility increases with the increase of ethanol proportion. That is the reason why we decided to work with a solution containing 50% ethanol / water (v/v) where the solubility limit of iprodione is 1.2 10<sup>-3</sup> M. To confirm the results obtained previously, the isotherms were performed on MIS 4, MIS 5 and MIS 8.

#### 3.1.3. Isotherm study

Isotherm studies were performed on both imprinted and non-imprinted polymers. Previously, equilibrium binding experiments were performed to determine the time necessary to reach this equilibrium (Figure 5). It is observed that the initial binding rate was very high with a very fast uptake profile. In fact, the system reached an equilibrium after 20 minutes. This means that the porosity of the silica matrix had a favorable effect on the diffusion time of the target molecules into the recognition sites. The fact that MIS showed fast kinetics for iprodione adsorption is advantageous. Based on this characterization, all furthers experiments, whatever the studied MIS, were determined within 60 minutes.

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In order to compare MIS 8 with its APTMS analogue, a new MIS, labelled MIS 5', was synthesized with the same (template / functional monomer / crosslincker) ratio molar 1 / 4 / 59 used for MIS 8 (Table 2). Figure 6 shows the isotherms for MIS 4, MIS 5' and MIS 8 and their respective control polymer NIS as a function of iprodione solution concentrations. The results show that all the MIS can bind iprodione in larger amounts than the analogous NIS and binding capacity for MIS increase with the rise of iprodione concentration solution. The results obtained in a 50% ethanol solution (v/v) confirm those obtained in a 10% ethanol solution (v/v): MIS 5' had the highest adsorption capacity of iprodione, followed by MIS 8 and MIS 4 in decreasing order of adsorption capacity. In addition to having the best iprodione binding property (around 25 mg of iprodione bound per g of MIS), MIS 5' had the best imprinting factor (about 271, Table 3) meaning the presence of specific interactions between iprodione and the functional monomer. This result indicates that in this case, the increase of crosslinker content enhanced the specific interaction between MIS and iprodione and so show the selectivity of MIS 5'. In order to get a better view on the adsorption of iprodione by MIS, the Langmuir, Freundlich and Temkin isotherm models were applied to fit the experimental data. Linear regression is frequently used to determine the best-fitting isotherm. The applicability and suitability of the isotherm equation to the equilibrium data were compared by evaluating the values of the correlation coefficients. The results are presented in Table 4 and Figure 7. The regression coefficients r<sup>2</sup> ranged from 0.83 to 0.97 (r from 0.91 to 0.98) for Freundlich model, from 0.76 to 0.89 (r from 0.87 to 0.94) for Temkin model and from 0.30 to 0.80 (r from 0.55 to 0.89) for Langmiur model. With the Pearson's table (critical values of correlation coefficient), using a value of level of significance for a two-tailed test of 0.05, and with 5 points (freedom degree = 3), the critical r value is 0.878. These values of regression coefficients reflect a good fit of the Freundlich model and Temkin model but it can be noticed that the Langmuir model is not suitable for fitting the present data. Therefore the Freundlich model is most suitable and that applicability follows the order: Freundlich > Temkin model. Freundlich model is the best suited model in our study suggesting that the adsorption occurs in multilayers and in heterogeneous sites of this material with a non-uniform distribution of heat of adsorption over the surface <sup>22,23</sup>. Moreover as the experimental data fitted well to Tempkin isotherm, we can added that iprodione adsorption occurred because the heat of adsorption of iprodione in the layer decreases linearly because of increased surface coverage.

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- When the Freundlich equation has been determined to be a suitable binding isotherm model for
- 376 MIS, two additional binding parameters can be calculated incorporating the parameters of the
- 377 Freundlich: the number of binding sites,  $N_{K1-K2}$  (Eq. 1) and the weighted average affinity,
- 378 K<sub>K1-K2</sub> (Eq. 2). K<sub>1</sub> of K<sub>2</sub> values are within the boundaries K<sub>min</sub> and K<sub>max</sub> as defined by Eq. 3
- 379 <sup>24,25</sup>.

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$$N_{K_{min-K_{max}}} = a(1 - n^2)(K_{min}^{-n} - K_{max}^{-n})$$
 Equation 1

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$$K_{Kmin-Kmax} = \left(\frac{n}{n-1}\right) \left(\frac{K_{min}^{1-n} - K_{max}^{1-n}}{K_{min}^{-n} - K_{max}^{-n}}\right)$$
 Equation 2

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$$K_{max} = \frac{1}{C_{e(min)}}$$
 and  $K_{min} = \frac{1}{C_{e(max)}}$  Equation 3.

with C<sub>e</sub> the free concentration of iprodione (mM) in the solution.

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- Using Freundlich isotherm model, no significant n values were observed between MIS 4, MIS
- 5' and MIS 8 (Table 4). A more homogeneous system will have an n value approaching unity
- and a more heterogeneous system will have an *n* value approaching zero. As *n* values were
- 388 close to the value 1, MIS 4, MIS 5' and MIS 8 also show high degree of binding site
- 389 homogeneity.

On the other hand, important variations were observed for the parameter  $K_F$  (Table 4). The  $K_F$ values show that the MIS 5' had significantly the highest binding capacity following by MIS 8 and then MIS 4. The comparison of the average number of binding sites N confirms that MIS 5' displayed significantly much higher binding capacities (31, 8 µmol/g) than MIS 8 and MIS 4 (p value = 0,0157). Moreover, the average association constant  $K_{Kmin-Kmax}$  is around two times greater for the MIS 5' than for MIS 8 and MIS 4 (p value = 0, 0046). No significant differences were observed for MIS 8 and MIS 4 about the value of N and K<sub>Kmin-Kmax</sub>. This higher capacity for MIS 5' should be attributed to a gain in the specific sites in line with this higher imprinting factor. The experimental data fitted well to Tempkin isotherm indicating that iprodione adsorption occurred because the heat of adsorption of iprodione in the layer decreases linearly because of increased surface coverage <sup>18,19</sup>. No significant K<sub>T</sub> values were observed between MIS 4, MIS 5' and MIS 8 (Table 4, p value = 0, 418) meaning a constant maximum bonding energy whatever the MIS. The calculated values of sorption heat, B value and b value, were significantly different for MIS 4 as opposed to MIS 5' and MIS 8, which had similar values. Very low values of b obtained in the present study suggesting that the uptake of iprodione onto MIS was by physisorption. In fact, the physisorption processes are reported to have adsorption energies less than -40 kJ/mol and characterized by very weak forces, such as Van der Waals forces and hydrogen bonding<sup>26,27</sup>. On the basis of these results, MIS 5' was chosen as the best MIS and these performances were

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#### 3.2. Comparison of MIS MIP performances

compared to that of an analogue MIP.

A MIP was synthesized using a functional monomer analogous to APTMS. Allylamine was chosen (Figure 1). Previous studies were performed to determine the optimal (template /

functional monomer / crosslinker) molar ratio. Isotherm studies were performed on MIS 5' and optimized MIP (Figure 8). The results show that MIS 5' and optimized MIP can bind iprodione and the binding capacity increased with the rise of iprodione solution concentration. The quantity of iprododione adsorbed was very different between MIS 5' (around 25 mg/g polymer) and MIP (around 120 mg/g polymer). It was 5 times lower for MIS than for MIP. However, the difference in the adsorption of iprodione between MIS and NIS was greater than the difference between NIP and MIP. In fact, the imprinting factor was of 271 for MIS and only 2 for MIP (Table 3). MIS 5' presented a high specific recognition of iprodione. The imprinting factor of MIP is in agreement with the previous study 8, where a value of 2.40 was determined. The imprinting factor of MIS is considerably higher than ones reported in the literature for molecularly imprinted sorbent materials, ranging from 0.8 to 39.85 <sup>12,14,28</sup>. The large imprinting factor value of this MIS is an indication of the great analytical potential of silica-based molecularly imprinted materials. This high selectivity is promising for further development in this area. Our hypothesis to explain this difference of imprinting factors between MIP and MIS is the homogeneity of the silica polymer. MIS 5' show high degree of binding site homogeneity (n value 0.86). MIP had an n value of 0.47 meaning a binding site heterogeneity (Table 4). At the opposite, for the binding affinity (K<sub>F</sub>), which refers to the strength of binding interaction, MIP had a higher binding capacity (K<sub>F</sub> of 490) than the MIS 5' (K<sub>F</sub> of 202). We can supposed that this higher binding capacity could be due to non-specific interactions. The imprinting factor takes into account the non-specific interactions which are due to the interaction between the template and the crosslinker. The average number of binding sites N confirms this assumption: MIS 5' displayed significantly much lower binding capacities (31,8 µmol/g) than MIP (229 µmol/g). Moreover, the average association constant K<sub>Kmin-Kmax</sub> is around five times greater for the MIP than for MIS 5'.

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The values obtained from the Temkin isotherm were:  $K_T = 75$  g/ $\mu$ mol and 312 g/ $\mu$ mol for respectively MIS 5' and MIP and b values = 104 J/mol and 40 J/mol for respectively MIS 5' and MIP, indicating that the adsorption of iprodione onto MIS or MIP occurred via physisorption involving low energies interactions between iprodione - MIS and between iprodione - MIP.

Scarce are the papers describing the comparison between MIS and MIP. A direct comparison between MIS and MIP was performed for the analysis of propranolol <sup>29</sup>. The findings were that the MIP exhibited higher uptake towards propranolol, but this was accompanied by a high degree of nonspecific binding. The MIS had lower uptake, but remarkably lower nonspecific binding (<10%). Another comparative study of the potential of MIP and MIS for molecular imprinting of 2-aminopyridine showed that the MIP retained approximately 50% of the template in rebinding studies in chloroform compared to 100% for the MIS. However, this higher affinity for the MIS was accompanied by a higher degree of non-specific binding. While the MIP performed poorly in acetonitrile, the MIS maintained a high degree of discrimination <sup>30</sup>.

#### 4. Conclusion

The optimal MIS was obtained with APTMS as functional monomer, low solvent volume and high crosslinker quantity during its synthesis. It showed a very fast uptake profile for iprodione. The data modeling revealed that the adsorption process follows the Freundlich model. The optimal MIS had a higher imprinting factor but a lower binding capacity than the optimal MIP specific to iprodione. The imprinting factor of MIS was 135 times higher than MIP. Unlike MIP, MIS kept its molecular recognition nature even under extremely polar media such as the aqueous sample. This work emphasizes the high potential of MIS as molecularly imprinted

materials for analytical applications and provides promising insights into the future development of MIS. Practically, MIS is as simple as MIP to prepare. MIS technology seems to allow more specific binding than MIP but less sensitivity. To compensate the lower sensitivity compared to MIP, a bigger column for SPE and more MIS quantity plus a bigger volume of sample during the loading phase would be necessary to identify traces in a beverage (wine for ex.) or in washing water of fruits. The next step to the study is to assess the selectivity of MIP and MIS towards pesticides.

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#### **Conflict of interest statement**

- The authors declare that there is no conflict of interest. No conflicts, informed consent, human
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### 543 Figure captions 544 545 Figure 1: Structure of functional monomers tested for the synthesis of molecularly imprinted 546 silica (MIS) (a, b, c) and for the molecularly imprinted polymers (MIP) (d) and structure of 547 iprodione (template, e) 548 (a) (3-Aminopropyl)trimethoxysilane 549 **(b)** 1-[3-(Trimethoxysilyl)propyl]urea 550 (c) N-[3-(Trimethoxysilyl)propyl]aniline 551 (d) allylamine 552 (e) iprodione 553 554 Figure 2: Binding capacity of polymer toward iprodione for MIS 1, MIS 6 and MIS 7 per 10 mg of polymer in a solution of iprodione at 10<sup>-5</sup> M (10% ethanol / water, v/v). Bars represent 555 556 standard deviation. 557 558 Figure 3: Binding capacity of polymer towards iprodione for MIS 1, MIS 4, MIS 5, MIS 7 and 559 MIS 8 and their control polymer NIS respective per 10 mg of polymer in a solution of iprodione at 10<sup>-5</sup> M (10% ethanol / water, v/v). Bars represent standard deviation. 560 561 562 Figure 4: SEM images of (a) MIS 1, (b) MIS 5, (c) MIS 8 and (d) MIS 7. Scale bar and 563 magnifications are mentioned on the SEM images. 564 Figure 5: Kinetic adsorption of MIS to iprodione solution (50% ethanol / water, v/v). Bars 565 represent standard deviation.

Figure 6: Binding isotherme of MIS 4, MIS 5' and MIS 8 toward iprodione at 25°C after 1h of 567 568 contact time with solution of iprodione (50% ethanol / water, v/v). Bars represent standard 569 deviation. 570 571 Figure 7: Isotherms models of the adsorption of iprodione by MIS 4. Sorption isotherms 572 obtained at 25°C after 1h of contact time with solution of iprodione (50% ethanol / water, v/v) 573 (a) Langmuir plot 574 **(b)** Freundlich plot 575 (c) Temkin plot 576 577 Figure 8: Binding isotherme of MIS 5' and MIP toward iprodione at 25°C after 1h of contact 578 time with solution of iprodione (50% ethanol / water, v/v). Bars represent standard deviation. 579 580

**(a)** 

**(b)** 

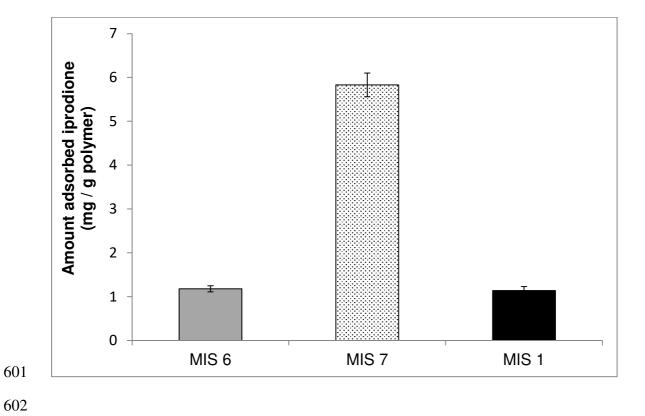
$$OCH_3$$
  $O$   $H_3CO-Si$   $N$   $NH_2$   $NH_2$ 

**(c)** 

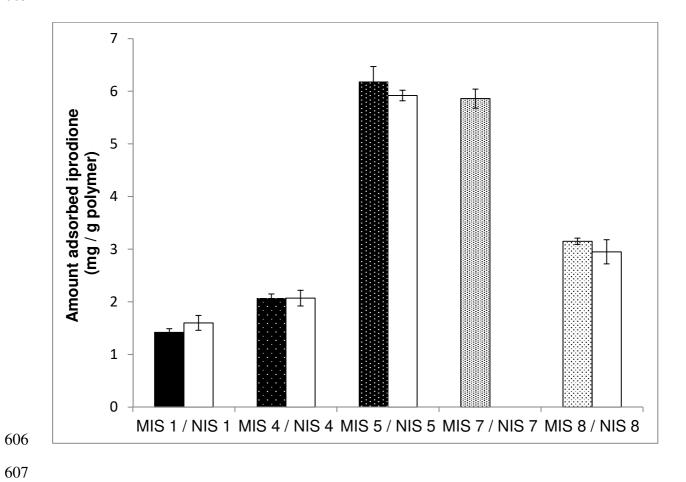
**(d)** 

$$H_2C$$
  $NH_2$ 

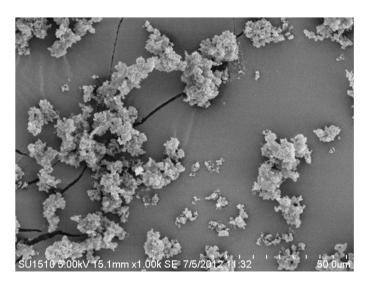
**(e)** 



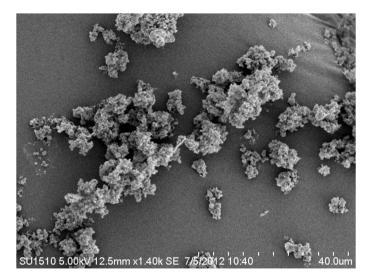
**Figure 3** 



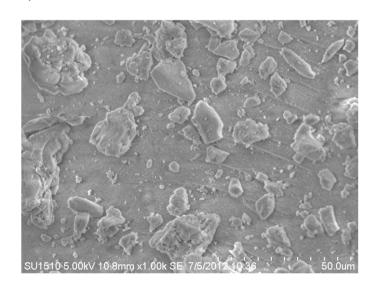
09 **a**)

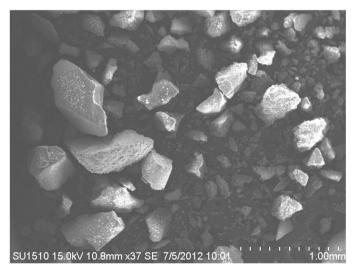


12 **b**)

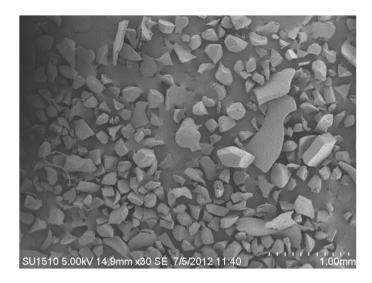


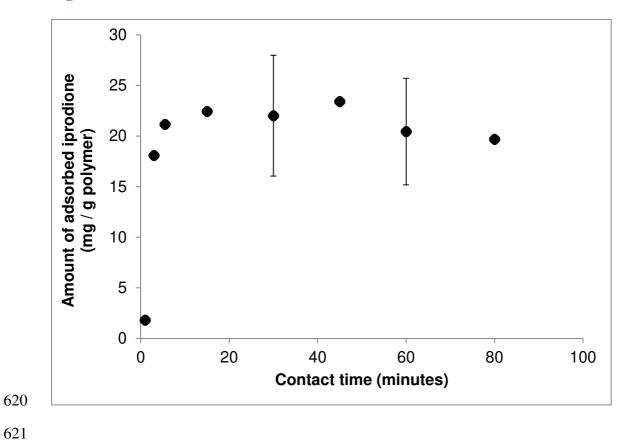
14 **c**)

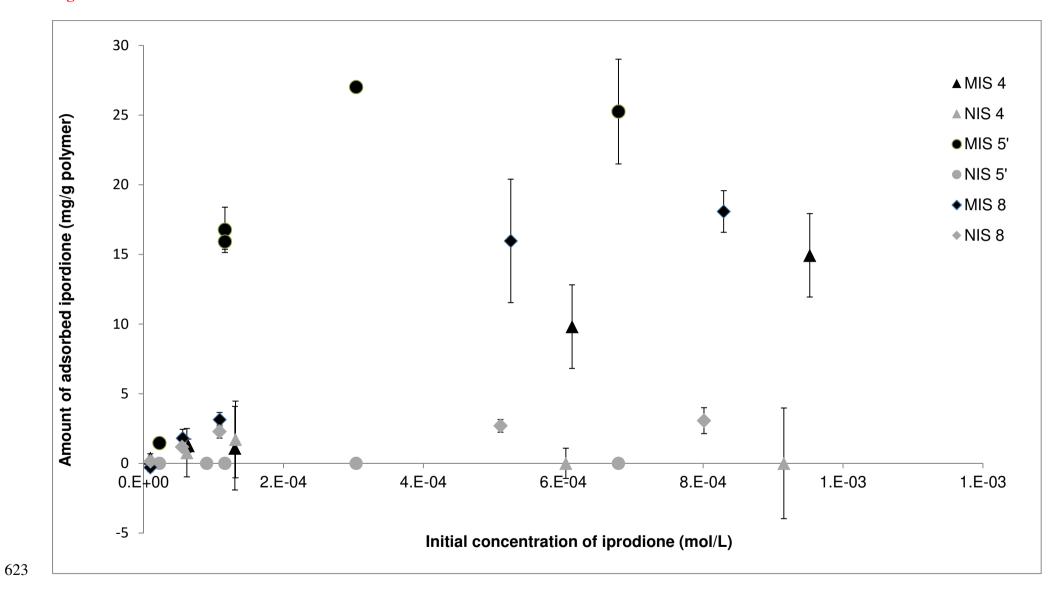




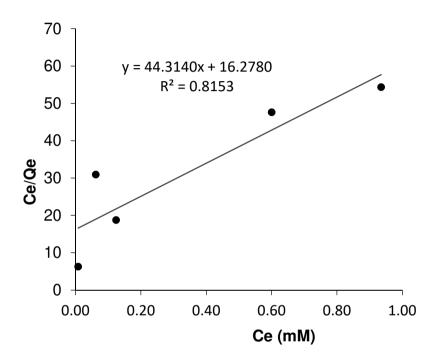
16 **d**)



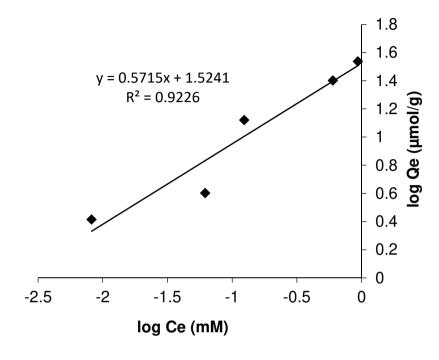




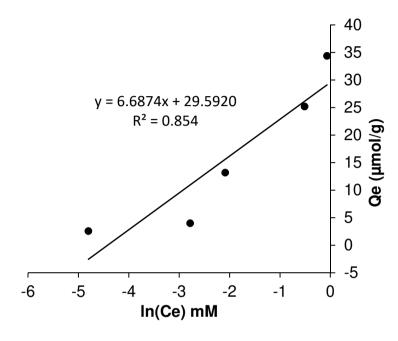
## **(a)**



**(b)** 



**(c)** 



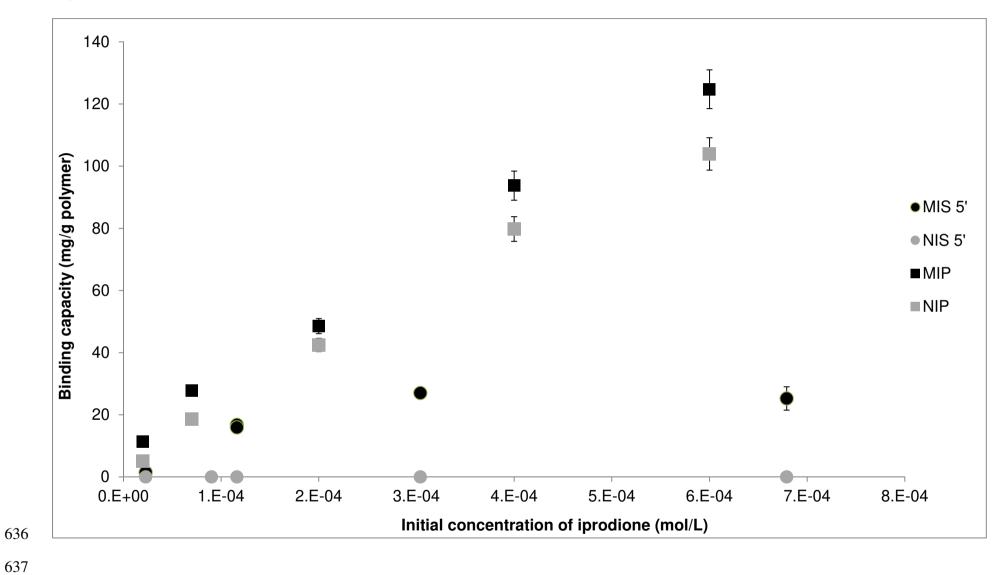


 Table 1: Nature of molecularly imprinted polymer using fungicide as template

Nature of molecularly imprinted polymer	Template	Nature of sample	Functional monomer	Reference
	imidazole fungicides (climbazole, clotrimazole, miconazole)	river water	methacrylic acid (MAA)	32
	dicarboximide fungicide (iprodione)	white wine	methacrylamide (MAM)	8
	imidazole fungicide (climbazole)	river and tap water	methacrylic acid (MAA)	33
	triazine (terbutylazine)	river water	methacrylic acid (MAA)	34
	triazine (terbutylazine)	surface waters and an industrial effluent	methacrylic acid (MAA)	35
	triazine (terbutylazine)	groundwater	methacrylic acid (MAA)	36
	triazine (terbutylazine)	river water	methacrylic acid (MAA)	37
Molecularly	triazine (terbutylazine)	tap water and groundwater	methacrylic acid (MAA)	38
imprinted	triazine (terbutylazine)	grape juice and soil extract	methacrylic acid (MAA)	39
polymer (MIP)	benzimidazole fungicide (thiabendazole)	river and tape water	methacrylic acid (MAA)	40
	chlorinated fungicide (fenarimol)	apple, banana, and tomato	methacrylic acid (MAA)	41
	benzimidazole fungicide (thiabendazole and carbendazim)	orange peel	methacrylic acid (MAA)	42
	anilinopyrimidine fungicide (pyrimethanil)	red wine	methacrylic acid (MAA)	43
	benzimidazole fungicide (thiabendazole)	spiked river, tap and well water	methacrylic acid (MAA)	44
	triazole fungicides (triadimefon)	cucumber	methacrylic acid (MAA)	45
	triazole fungicide (difenoconazole)	tap water and grape juice	methacrylic acid (MAA)	46
	morpholine fungicide (dimethomorph)	ginseng powder	methacrylic acid (MAA)	47
Molecularly imprinted sol-	dicarboximide fungicide (iprodione)	white wine	(3-Aminopropyl)trimethoxysilane (APTMS)	12
gel polymer (MIS)	dicarboximide fungicide (iprodione)	hydroalcoholic medium	(3-Aminopropyl)trimethoxysilane (APTMS)	48

Triazine (atrazine)	sugar cane juice	(3-Aminopropyl)trimethoxysilane	49
		(APTMS)	

**Table 2:** Samples codes of different MIS and NIS synthesized for this study depending on different natures of functional monomers and different molar ratios of reagents.

template / functional monomer / crosslinker molar	1/4/20	1/4/20	1/4/50	1/4/59	1/4/20	1/4/20	1/4/59
ratio							
Functional monomer	APTMS	APTMS	APTMS	APTMS	Urea	Aniline	Aniline
Ethanol / water ratio (mL)	2.3 / 1.3	1.15 / 0.44	1.15 / 0.44	1.15 / 0.44	2.3 / 1.3	2.3 / 1.3	1.15 / 0.44
Code number of MIS synthetized	MIS 1	MIS 4	MIS 5	MIS 5'	MIS 6	MIS 7	MIS 8
Code number of NIS synthetized	NIS 1	NIS 4	NIS 5	NIS 5'	NIS 6	NIS 7	NIS 8

Table 3: Imprinting factors obtained for MIS ans MIP depending on the nature of the solution (10 % or 50 % (v/v) ethanol / water solution). In each line, mean values were calculated using three repetitions  $\pm$  standard deviation. Means with different superscripts (a – c) are significantly different at p < 0.05 using Fisher t-test.

Nature of the solution	MIS 1	MIS 4	MIS 5	MIS 5'	MIS 6	MIS 7	MIS 8	MIP
from 10 % (v/v) ethanol/water solution	$0.96 \pm 0.11$	$0.99 \pm 0.07$	$1.15 \pm 0.42$	nd	nd	nd	$1.13 \pm 0.06$	nd
from 50 % (v/v) ethanol/water solution	nd	99 ± 31 b	nd	271 ± 43 °	nd	nd	6 ± 2 a	2 ± 0.1 a

Table 4: Adsorption isotherms constants from Langmuir, Freundlich, Temkin models obtained at 25°C with 50 % (v/v) ethanol / water solution. In each line, mean values were calculated using three repetitions  $\pm$  standard deviation. Means with different superscripts (a – c) are significantly different at p < 0.05 using Fisher t-test.

Isotherms	Constants	MIS 4	MIS 5'	MIS 8	MIP
	$r^2$	$0.91 \pm 0.02$	$0.83 \pm 0.05$	$0.93 \pm 0.03$	$0.97 \pm 0.05$
	n	$0.75 \pm 0.17^{a}$	$0.86 \pm 0.04^{\text{ a}}$	$0.91 \pm 0.10^{a}$	$0.47 \pm 0.02^{\mathbf{b}}$
Freundlich	$K_{F}$ (mM <sup>-1</sup> )	38 ± 5 a	202 ± 35 °	76 ± 11 b	$490 \pm 24^{d}$
	$K_{(K MIN-K MAX)} (mM^{-1})$	$3.6 \pm 0.4^{\text{ a}}$	$6.3 \pm 0.3$ b	3.8 ± 0.2 a	32 ± 1.6 °
	N (µmol/g)	$12 \pm 5.2^{a}$	$31.8 \pm 6.7$ b	15.5 ± 1.8 a	229 ± 11.5 °
	$r^2$	$0.76 \pm 0.09$	$0.89 \pm 0.09$	$0.89 \pm 0.09$	$0.82 \pm 0.04$
Temkin	K <sub>T</sub> (g/μmol)	57 ± 23 a	75 ± 14 a	75 ± 14 a	$312 \pm 15^{\text{ b}}$
	B ((µmol/g)/mM)	9.3 ± 2.5 a	$24.5 \pm 4.4^{\text{ b}}$	$24.5 \pm 4.4^{\text{ b}}$	$62 \pm 3^{c}$
	b (J/mol)	282 ± 81 °	104 ± 19 b	104 ± 19 b	40 ± 2 a
Langmiur	$r^2$	$0.30 \pm 0.45$	$0.61 \pm 0.13$	$0.43 \pm 0.36$	$0.80 \pm 0.04$

